

The Unique Deletion, Insertion and Point Mutation in The Upstream of Nucleocapsid (N) Gene of Different Covid-19 Variants Suggests Differential Translational Mechanisms of N-Protein

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Abstract

The nucleocapsid protein (N protein; 219 AAs) of SARS-CoV-2 was synthesized from terminal part of the ~30kb genome and a 31ERS three AAs deletion was reported in all Omicron variants but not in Alpha, Beta, Gamma and Delta variants. We found that upstream of N-gene which also a part of ORF8 gene has different deletion, insertion and point mutations in different corona virus lineages. As for example, 119DF two amino acids (5'-28243GATTTC-3') deletion was found in ORF8 protein of Delta variants located adjacent to the N-gene upstream. Similarly, four nucleotides (5'-AACCA-3') insertion at 28262 position was found in the Gamma (P.1) variant only. Whereas all Omicron variants (BA.1/BA.2/BA.4/BA.5) and subvariants (XBB.1.5, BQ.1.1) which were spreading now with mild diseases, have no such deletion or insertion. Some point mutations T>C at 28247 position were found in Alpha variants in that small region of N-gene upstream. Moreover, "GAT" sequence at 28279 position in Gamma variant similar to Wuhan and Beta (B.1.351), was changed to "CTA" in Alpha variant (B.1.1.7) at the N-gene upstream. The extra "A" at 28273 position in Alpha variant (also found in Wuhan and Gamma) was equally distributed ("AAA" vs "AAAA") as judged by BLAST-N search but it was not instrumental if there was a sequence error. The differential hairpin structures were obtained with upstream of Delta and Gamma but no such difference was noticed with hairpin structures of different Omicron N-gene upstream. We suggest that hairpin structures with higher δG specifically in Gamma variant (P.1) may reduce the translation of N-protein and may be the one of the causes of extinct of that lineage of corona virus.

Keywords: COVID-19, N-Gene Upstream, Translation Control, Hairpin Structure, Orf8 Gene, ERS N-Protein Deletion.

1. Introduction

Corona virus pathogenesis has rippled this Earth with few million deaths worldwide [1]. COVID-19 was detected in March-2019 and whole genome sequencing was available from December, 2019 onwards [2]. During three years period many mutations in the diverse variants of COVID-19 genomes were reported in the NCBI SARS-CoV-2 Database [3]. However, related SARS and MERS RNA viruses were known since 2003 and considerable molecular biology of such viruses have been reported in the PubMed [4,5]. Most astonishing fact was large polyprotein (7096 AAs) synthesis in the infected cells and such proteins were proteolytically cleaved into 16 polypeptides with important biological functions [6]. The Nsp1 protein is 180aa (regulatory factor), nsp2 is 638aa (RNA topoisomerase), nsp3 is ~1945aa (C3 protease), nsp4 is 500aa (membrane factor), nsp5 is ~305aa (C5 protease), nsp6 is 290aa (membrane factor), nsp7 is 183aa (accessory protein to replication), nsp8 is 198aa (accessory protein to replication), nsp9 is 113aa (RNA binding factor), nsp10

is 139aa (RNA binding factor), nsp11 is only 13aa (unknown function), nsp12 is 918aa (RNA-dependent RNA polymerase), nsp13 is 601aa (RNA helicase-capping methyltransferase), nsp14 is 527aa (exoribonuclease-methyltransferase), nsp15 is 346aa (endoribonuclease-recombinase), nsp16 is 298aa (2'-O Uridine rRNA methyltransferase) [9-22]. On the country, structural spike protein is 1273aa long and other structural proteins (M, N, E) of corona virus are relatively very small [24-29]. The regulatory proteins like orf3a, orf7a, orf7b, orf8 and orf10 were also characterized having interacted with many cellular proteins [30-35]. The N-protein is a strong RNA-binding protein (419aa) and requires long structured RNA as substrate [36]. Importantly, a 31ERS deletion of the protein was found in all omicron viruses including BA.2.75, BF.7, BQ.1, BQ.1.1, BQ.1.1.1 and XBB.1.5 sub subvariants [34,37]. N-protein of SARS-CoV-2 has 91% and 49% similarity to SARS-CoV and MERS-CoV respectively and is predicted to be predominantly a basic nuclear protein [38]. The N-protein has a modular structure with an N-terminal

RNA-binding domain (RBD) and a C-terminal dimerization domain (CTD), including three well characterized intrinsically disordered regions (IDRs) between the RBD and CTD [39-41]. The protein oligomerizes through its CTD and disordered C-terminal tail whereas undergoes liquid-liquid phase separation with RNA mediated by its RBD and central disordered region [42]. Cells transfected with N-protein showed a G1/S phase block with an increased expression of tubulin isomers like TUBA1C and TUBB6 [41]. The twelve phosphorylated sites and nine potential protein kinase sites in N-protein suggested as promising targets for drug discovery and development for of a recombinant virus vaccine [38,43]. The R203K and G204R

mutations in the N-protein destabilized and decreased overall structural flexibility [36]. The N-protein (419aa) is required for viral life cycle and strongly binds to viral RNA regulating its replication as well as transcription of sub-genomic RNAs. A novel DNA-aptamers binding to the SARS-CoV-2 nucleocapsid protein and its variants, Delta and Omicron was discovered recently improving diagnostic tools for SARS-CoV-2 [44,45]. The replication of COVID-19 virus has been investigated but translational mechanisms were largely unknown [46]. We have reported here the sequence variation in the small upstream of N-gene of different corona virus variants.

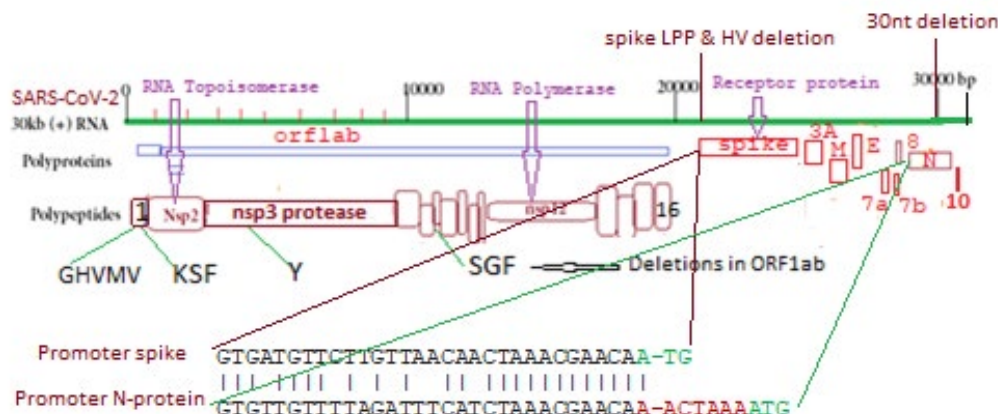


Figure.1: Structure of COVID-19 and similarity between promoters of spike protein and N-protein. The deletions in different omicron lineages were also seen and such viruses like BQ.1, XBB.1.5 and BF.7 were less pathogenic.

2. Methods

We searched PubMed to get idea on published papers on N-gene upstream and also searched SARS-CoV-2 NCBI database using BLAST-N and BLAST-X search methods to get sequences. Multi-alignment of protein was done by MultAlin software [47,48] and multi-alignment of DNA by CLUSTAL-Omega software, EMBL-EBI [49,50]. The ORF1ab mutants was obtained by BlastN search of deletion boundary of 60-100nt sequence and then analyzing the sequences with 95-100% similarities. The Hairpin structure of ~ 120-200nt sequence was done by OligoAnalyzer 3.1 software (Integrated DNA Technologies) [51,52]. The protein 3-D structure of N-protein was determined by SWISS-Model software [53,54].

3. Results

BLAST search and Multi-alignment demonstrated a variation in the upstream of N-protein among Wuhan, Beta, Gamma, Delta corona virus sequences (figure-2). But all omicron variants N-gene upstream sequences were found homogeneous and no 5'-AAGC-3' sequence insertion at 28262 position as found in Gamma (P.1) variant. Moreover, in Alpha (B.1.1.7) variant N-protein upstream, some mutations were found. Further, in Delta variant, a "DF" two amino acid deletions (5'-28243GATTTC-3') in the small ORF8 protein carboxy-terminal was prominent at that locus (figure-3). Previously, we demonstrated hotspot deletion sites in the ORF7a gene and three TAA termination codons creating short chimera proteins [32-34]. Thus, sequence variation in that region of 3'-end of the SARS-CoV-2 may be one of the reasons of viral extinction or slower replication and

pathogenicity and N-protein was implicated in replication and pathogenicity. Indeed, downstream of N-gene, another 26nt deletion at the 3'-UTR was prominent in most omicron corona viruses [34]. Surely, such big deletion in the 3'-UTR slower viral replication and viral titre.

In Table-1, we designed many oligonucleotides to check the abundance of different N-gene upstream mutants in the SARS-CoV-2 Database. The Alpha variant sequence (5'-cgt gtt gta gat ctg ttc tct aaa cga aca aac-3') at 28233 position at the upstream of ORF8 termination codon was justified and "tt" insertion-like alignment was true but point mutation. The Delta variant sequence (5'-ggt cgt gtt tta atc taa acg aac aaa c-3') was also justified. Similarly, Gamma "aac" insertion in Gamma variant COVID-19 sequence (5'-atc taa acg aac aaa caa act aaa atg tct gat aat gga c-3') at 28253 position (accession no. NC_045512.2) was true (>998 sequences). The Alpha variant variants sequence (5'-atc taa acg aac aaa cta aat gtc tct aaa tgg ac-3') was also justified (>5000 sequences). But one nucleotide "A" addition abolished the specificity of Alpha whereas similar addition of one "A" to other positions did retain specificity (>5000). On the contrary, further deletion of "AA" abolished the Wuhan and Beta specificities at the both positions (Table-1). It appeared that "AAA" vs "AAAA" at the middle of N-gene upstream equally distributed in the database (>5000 sequences). Thus, if it is a sequencing error or indeed "AAA" was the mutant form was not clear as "AAAA" at that position found in Wuhan (B.0) as well as Gamma and Alpha variants but not valid in Delta variant (preference of "AAA"). Moreover, "GAT" sequence at 28279

position in Gamma variant similar to Wuhan and Beta (B.1.351) variants, was changed to “CTA” in Alpha (B.1.1.7) variant at the N-gene upstream. We concluded that the upstream sequences of the N-protein were varied in different Wuhan, Beta, Alpha and Delta variants. Such changes may increase or decrease the production of N-protein but such data is very limited in the PubMed. The N-protein binds to corona virus RNA genome and regulates the replication as well as viral mRNA synthesis and expected its expression will be tightly regulated in host cells [55,56].

We performed the hairpin secondary structure of the N-gene upstream of different corona virus variants and detected minor changes in nob-like structures with different δG (figure-4). Indeed, we found higher δG in Alpha variant (-7.9 Kcal/mole) and to some extent in Delta variant (-9.1 Kcal/mol; Table-2). However, “AGA” insertion in Gamma variant did not change

the overall secondary stem-loop structure of N-gene upstream sequences (-9.8 Kcal/mol) as demonstrated in figure-4.

In figure-5, we demonstrated the similarity between promoters of Spike protein verses N-protein. But such similarity was not detected in the promoter of ORF1ab or M proteins (data not shown). We found the 30nt deletion in the 3'-UTR of Omicron corona viruses (BA.2, BA.4, BA.5, BF.7, BQ.1 and XBB.1.5 etc.). Such deletions including other spike deletions (24LPP and 69HV) and also spike insertions (215EPE and 249 RWMD) may be significant to lower the coronaviruses pathogenicity [32-34]. Overall, transcriptional differences among the deadly coronavirus B.1.1.7 and B.1.617.2 lineages as well as in recent omicron sub subvariants (XBB.1.5, BQ.1.1.1) never been investigated before. We have first shown here the minor but important variations among the VOC of coronaviruses with possible difference in transcriptional regulations.

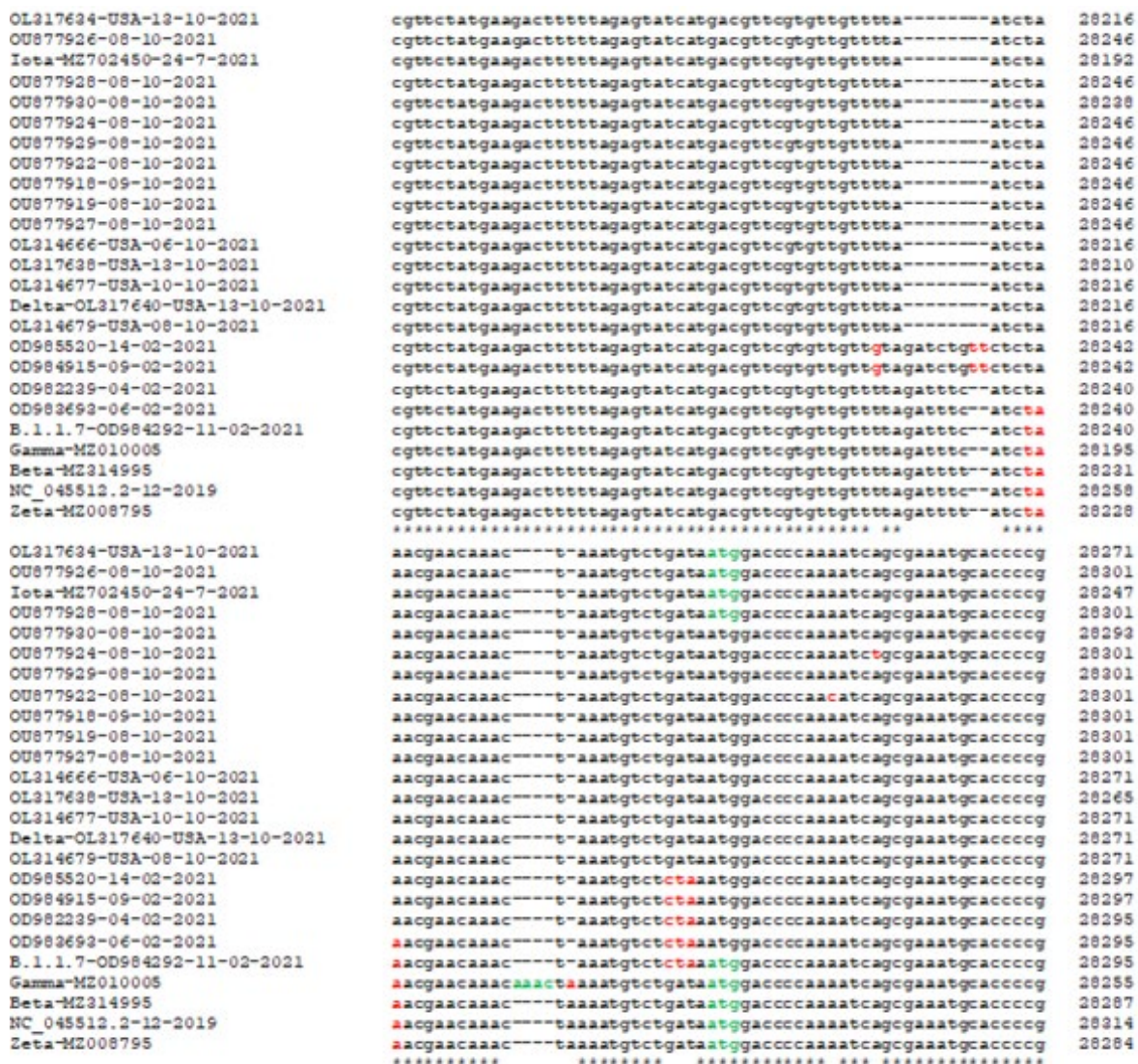


Figure.2: Multi-alignment of SARS-CoV-2 Delta, Alpha, Beta, Iota, Zeta and Gamma sequences showing different deletions and insertions surrounding N-gene. A “GATTTTC” sequence variation was important between Alpha and Delta Variants. A two nucleotides (TT) insertion was found upstream of ORF8 termination codon of Alpha variant and four nucleotide (AAAC) insertion was found upstream of N-protein in Gamma variant. Moreover, “GAT” was changed into “CTA” in Alpha variant than all other variants (Wuhan, Delta, Beta, Zeta and Iota). Further, a single nucleotide “a” deleted in Delta, Alpha and Iota variant as compared to Wuhan virus including Beta and Zeta variants.

BA.1.1.1-OP606805-6-1-2022	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28146
BA.1.1-ON394519	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28198
BA.1.1.2-ON394520	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28198
BA.1.1.18-ON386282	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28202
B.1.1.529-OL677199-N11	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28198
B.1.1.529-OL677199	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28198
BA.4.1.1-OP051187	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	27823
BA.4.0-OP258049	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28181
BA.4.1.6-OP257501	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28193
BA.4.1-OP436295	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28215
BA.4.1-OP257429	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.2-OP437162	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.1.1-OP307754	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28017
BA.4.2-OP306354	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28138
BA.4.1-ON991461	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28240
BA.4.2-OP257613	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.2-OP257529	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.5-OP257738	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.0-OP258051	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.4-OP257734	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.6-OP258130	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.6-OP257669	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.6-OP258078	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.4-OP258001	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.4.4-OP257777	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
B.2.75-OP571747	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	57968
BA.4.4-OP257451	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28190
BA.2-OM901219	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28230
BA.5.2.2-OP257545	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28199
BA.2.3-OP257551	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28199
BA.5.1.7-OP257528	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28199
BA.5.6-ON999542	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28177
BA.5.2-ON999606	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28210
BA.5-ON658807	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28210
BA.5.1-OP237923	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28195
BA.5.5-OP237919	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	27907
BA.5.2.1-OP238284	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28243
BA.5.2.1-OP238223	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28195
BA.5.2.1-OP237918	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28241
BA.5.2.1-OP238183	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28195
Gamma-ON017297-18-6-2021	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28266
Delta-ONS07031-20-12-2021	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28218
Alpha-MZ253074-3-5-2021	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28208
Wuhan-MT049951-17-1-2020	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28273
NC_045512.2-12-2019	ttagagtatcatgacggttcggtggttttagatctctaaacgaaca-----aacttaa	28273

Figure.3: The N-upstream of omicron corona viruses (BA.1, BA.2, BA.4 and BA.5 subvariants) have similar N-upstream as compared to Alpha, Gamma and Delta variants. The “GATTC” deletion in Delta variant and the ”ACA” insertion in Gamma variant as well as in Alpha variant were also shown.

No.	Oligonucleotides sequences and position (NC_045512)	Population	Variant
P1	(28233)5'-cgtggtggtgtagatctgctctaaacgaacaac-3'	>5000	Alpha
P2	(28233)5'-cgtggtggtgtagatctgctctaaacgaacaac-3'	33	Alpha*
P3	(28233)5'-cgtggtggtgtagatctgatctaaacgaacaac-3'	0	Alpha**
P4	(28253)5'-cgtggtggtgtagatctgctctaaacgaacaac-3'	1	Alpha***
P5	(28233)5'-cgtggtggtgtagatctgctctaaacgaacaac-3'	0	Alpha****
P6	(28256)5'-gttcggtggttttaactaaacgaacaac-3'	>5000	Delta
P7	(28256)5'-gttcggtggttttatctaaacgaacaac-3'	0	Delta*
P8	(28256)5'-gttcggtggttttaactaaacgaacaac-3'	6	Delta**
P9	(28256)5'-gttcggtggttttaactaaacgaacaac-3'	0	Delta***
P10	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	>998	Gamma
P11	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	20	Gamma*
P12	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	0	Gamma**
P13	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	0	Gamma***
P14	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	>5000	Alpha
P15	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	0	Alpha*
P16	(28253)5'-atctaaacgaacaacaaactaaaatgtctgataatggac-3'	0	Alpha**

P17	(28253)5'-atctaaacgaacaaactaaatgtctctaatggac-3'	>4987	Alpha***
P18	(28253)5'-atctaaacgaacaaactaaatgtctgataatggac-3'	>4999	Wuhan/Beta
P19	(28253)5'-atctaaacgaacaaactaaatgtctgataatggac-3'	>5000	Wuhan*
P20	(28253)5'-atctaaacgaacaaactaaatgtctgataatggac-3'	2	Wuhan**
P21	(28253)5'-atctaaacgaacaaactaaatgtctgataatggac-3'	110	Wuhan***

Table-1: Oligonucleotides used to BLAST-N search for the identification of different COVID-19 variants with mutated N-gene upstream sequences

Variant	DNA sequences	Delta G (Kcal/mole)
Wuhan	5'cgttctatgaagacttttagagtatcatgacgttcgtgtgttttagatttcataaacgaacaaa ctaaaatgtctgataatggaccccaaatcagcgaaatgcaccccg-3'(114nt)	-9.896
Alpha	5'cgttctatgaagacttttagagtatcatgacgttcgtgtgttagatctgttctctaaacgaacaa actaaatgtctctaaatggaccccaaatcagcgaaatgcaccccg-3'(115nt)	-7.929
Gamma	5'cgttctatgaagacttttagagtatcatgacgttcgtgtgttttagatttcataaacgaacaaac aaactaaaatgtctgataatggaccccaaatcagcgaaatgcaccccg-3'(118nt)	-9.896
Delta	5'cgttctatgaagacttttagagtatcatgacgttcgtgtgttttaataacgaacaaactaaatg tctgataatggaccccaaatcagcgaaatgcaccccg-3'(107nt)	-9.147
Omicron	5'cgttctatgaagacttttagagtatcatgacgttcgtgtgttttagatttcataaacgaacaaa ctaaaatgtctgataatggaccccaaatcagcgaaatgcaccccg-3'(114nt)	-9.896

Table-2: Delta G of Hairpin structures of N-gene upstream regulating translation of N-protein

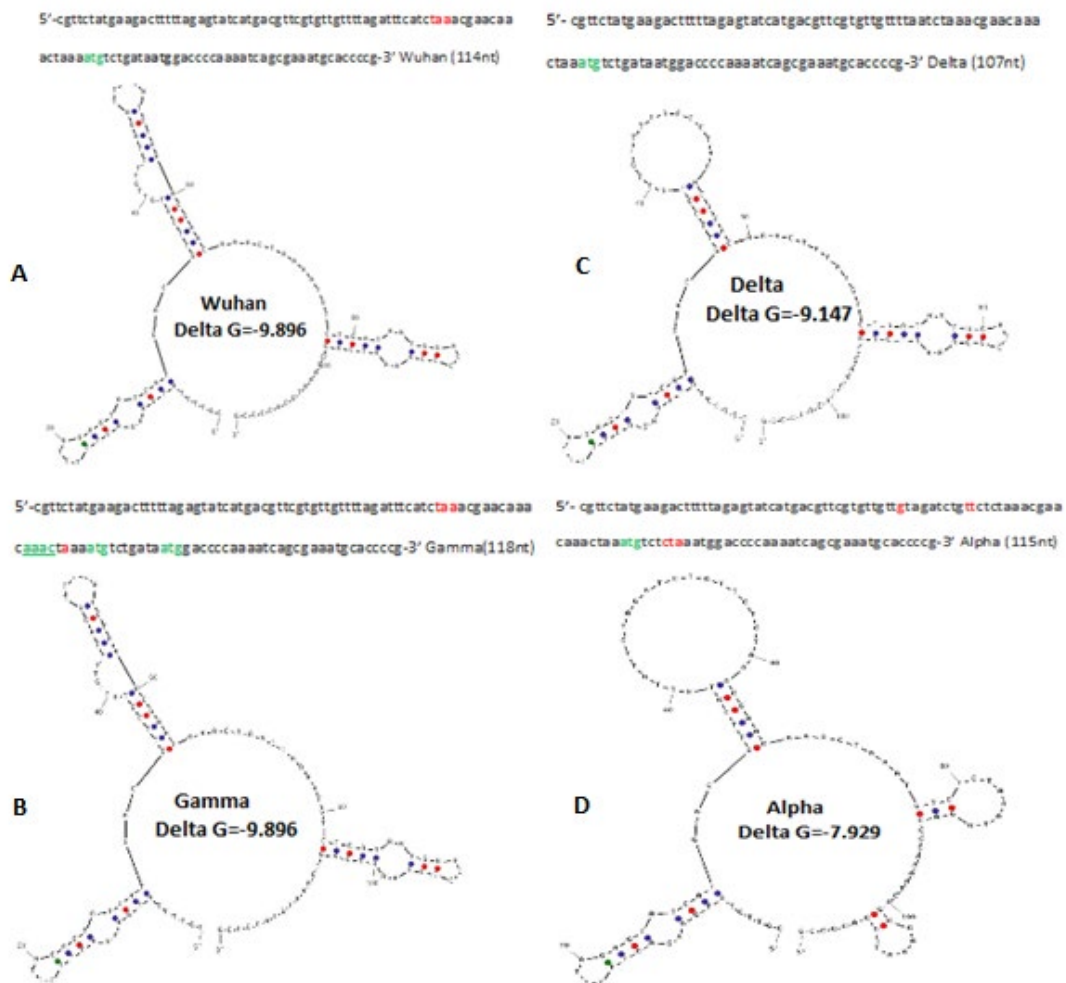


Figure.4: Hairpin structures of N-gene upstream of different corona virus variants. The hairpin structure of omicron lineages was similar to Gamma variant and was not shown here.

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P-S   GTGATGTTCTTGTTAACTAAACGAACAA-TG
      ||| |||| | | | || |||||
P-N   GTGTTGTTTTAGATTTTCATCTAAACGAACAA-ACTAAAATG

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Figure.5: Blast-2 similarity between Spike gene and N-gene upstream showing similarity. It was shown a 5'-AACTAAA-3' insertion in the ATG codon upstream in N-gene. The canonical TATATAAT promoter sequence was not detected. The SP3 (GGCCGGG), Myc (CACGTG), Oct1 (ATGCAAAT) eukaryotic promoter sequences were not detected

4. Discussion

In this communication, we reported the sequence variation among the different corona virus variants at the N-protein upstream as well as coding region. We recently reported the variations (Deletions, Insertions and Point Mutations) in the ORF1ab, ORF7a and ORF8 proteins [32-34]. Similarly, Spike protein deletions and huge mutations in omicron variants were well documented [25]. As for example, 24LPP deletion was found in most omicron corona viruses and 69HV spike deletion was found in omicron BA.1, BA.4 and BA.5 variants but not in omicron BA.2 variant. Similarly, 69HV deletion was first detected in B.1.1.7 lineages but not in B.1.617.2 lineages. Instead in B.1.617.2 lineages 157FR spike deletion was prominent. On the other hand, in omicron BA.1 lineage, both 69HV, 143VYY, 212L deletions as well as 215EPE insertion in the spike was documented [26]. Special D614G, N501Y, E484K, and L452R point mutations were instrumental in different corona virus variants and such mutations increased transmission and immune-escape [3,57]. The N501Y was found in B.1.1.7 but also transmitted in Beta, Gamma and Omicron variants but D614G mutation was found in all variants since its appearance in March, 2020. The E484K point mutation was prominent in Beta (B.1.351), Gamma (B.1.1.28.1) and Iota (B.1.526) variants as well as Mu (B.1.621) while at the same locus E484Q was reported in Kappa (B.1.617.1) variant and E484A in Omicron BA.1 (B.1.1.529) variant. The L452R point mutation was found in many COVID-19 variants like Delta (AY.103 and B.1.617.2) and Epsilon (B.1.429) variants while at the same locus L452Q point mutation was reported in Lambda (C.37) variant [6,23]. Thus, mutational landscapes continued since appearance of coronaviruses in 2003 and surely such minor variances in the subvariant populations determine its pathogenicity and omicron sub-lineages are very mild pathogenic [58].

Importantly, the deletions and mutations in the ORF1ab polyprotein (7092-7096 AAs) were reported at low rate compare to spike protein (1268-1273 AAs). The most important P4715L mutation in the RdRp was very dominant and T95I nsp2 RNA topoisomerase mutation was reported in Delta, Iota and few Omicron variants. It was a debate as D614G spike mutation increased 80% transmission in presence or absence of P4715L mutation in the RNA-dependent RNA polymerase. The 3675SGF deletion in the ORF1ab was very prominent in most corona virus variants except Delta variant and 141KSF deletion was favored in omicron BA.4 variant and subvariants while the 82GHVMV locus deletions at the upstream of 141KSF, were reported in few Omicron variants recently [8].

This lineage specific changes were not new and our finding of

N-gene upstream variation in sequence was important as many point mutations and deletions were found drastically enhanced viral transmission with increased pathogenicity. Obviously, characterization of corona virus sequences was done worldwide to tract specific VOC which may need extra caution to neighbor countries to control epidemic spread. The vaccination so far saved the world against Alpha and Delta variants but >30 mutations in the spike caused some doubt on early spike gene based genetic vaccines [60]. Recently, many laboratories worldwide engaged with yeast cells-expressed protein vaccine based on altered spike protein of Omicron corona viruses. However, adeno-vector based DNA-vaccine and mRNA vaccine were found good based on spike protein [59,60] as well as other proteins like RNA topoisomerase (nsp2) and uridine methyl transferases (nsp16) [9,18]. The co-infection of multidrug resistant bacteria in patients were reported and thus infections were very serious as most antibiotics failed to cure such pan-drugs resistant infections [61,62]. We recently found root and bark herbal extracts from Suregada multiflora (*Gelonium multiflorum*) and *Cassia fistula* (gloden showers) could combat such MDR bacteria [63-65]. Apart from N-protein, other small protein like nsp1 (180AAs) has profound role controlling virus replication and pathogenicity [66,67].

5. Conclusion

The genetic changes in RNA viruses have been well documented and ubiquitous. While the changes in N-gene upstream in different corona virus variants may throw some insights into extinct of specific lineage with time. Undoubtedly, we first highlights the possible differential regulation of N-gene in different SARS-CoV-2 variants.

Acknowledgement

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Conflict of interest

The author declares no competing interest and the data discussed here was computer generated.

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