

## The Immune Cells and Its Link to COVID-19

Archana Tripathy

Disease Biology Laboratory, School of Biotechnology, Kalinga Institute of Industrial Technology (KIIT) deemed to be University, Bhubaneswar-751024, Odisha, India

### \*Corresponding author

Archana Tripathy, Disease Biology Laboratory, School of Biotechnology, Kalinga Institute of Industrial Technology (KIIT) deemed to be University, Bhubaneswar-751024, Odisha, India

Submitted: 17 Jun 2020; Accepted: 22 Jun 2020; Published: 06 July 2020

### Abstract

World health organization has declared SARS-CoV-2 infection as a worldwide pandemic on March 11, 2020 and it is continuously affecting public health throughout the globe. The disease progresses from mild symptoms to a pneumonia like condition with severe inflammation of the respiratory tract due to cytokine release or cytokine storm that is the major characteristic of this disease. T cells numbers decrease and become exhausted in COVID positive patients this might be due to excessive amount of IL 10, IL 6, and TNF $\alpha$ . CD8+T cells and NK cells have showed functional impairment on differentiation, maturation and adequate amount cytokine production which lead to compromise the host immune response against SARS-CoV-2 infection. IFN $\gamma$  behaves as a protective cytokine at early or recovery stages and at severe stage, it acts as more pathogenic by inducing anti-viral responses. This review has summarized the current states of immune responses regarding SARS-CoV-2 infection. It might be helpful on offering new understandings and therapeutic approaches for COVID-19.

**Keywords:** SARS-CoV-2, T Cells, NK Cells, B Cells, IFN $\gamma$

### Introduction

The onset of the new decade has been with the spreading of a novel corona virus disease COVID-19 from the Chinese province of Wuhan to more than 200 countries across the world. This worldwide spread has led the World Health Organization to declare COVID-19 a pandemic on 11 March 2020. Clinical symptoms of COVID-19 are mild fever, cough, dyspnea and in few severe cases it leads to respiratory failure or death [1, 2]. SARS-CoV-2 is closely connected with MERS (Middle Eastern Respiratory Syndrome) and SARS-CoV-1 coronaviruses, prevailing local outbreaks in 2012 and 2003 respectively [3]. The entry of corona virus into the host cells happens by attachment of viral spike (S) protein to host ACE2 (angiotensin-converting enzyme 2) receptor and resulting increase serum Angiotensin II level [4].

Respiratory viruses have induced excessive inflammatory responses, which might result in robust cytokine production [5, 6]. The reason of dysregulation of immune system due to corona virus till remains elusive. Scientific community still investigates how SARS-CoV-2 infections disturb the immune homeostasis and activate inflammatory responses in host. A study suggested, corona virus might effectively mislead the innate immunity causing a delayed or inadequate response in patient [7]. As SARS-CoV-2 is a RNA virus, the genomic content can bind to the pattern recognition receptors (PRR) like cytosolic RLRs (RIG1 like receptor) and extra or intra cellular TLRs (Toll like receptor). The downstream pathways followed by

COVID infection via PRRs start triggering the secretion of certain cytokines. IFN $\gamma$  is the prominent cytokine that plays a vital role in inducing antiviral responses by transmitting signal via JAK/STAT pathway for the activation of interferon-stimulated genes [8]. It was also found, IFN $\gamma$  is protective in early stage of disease and during severity, it becomes pathogenic [9]. HLA (Human leukocyte antigen) alleles are complex polymorphic components of the viral peptide presentation process that initiate adaptive immune response to eliminate the infection. Recently an in silico analysis has identified certain HLA alleles having more predicted SARS-CoV-2 peptide that might provide evidence of genetic susceptibility or resistance to SARS-CoV-2 infection [10]. Several studies have explained about the status of Natural killer (NK) cells during infection, they found reduction in the number of NK cells, diminishing surface receptors and impairment in the production of cytokines [11-14]. However, CD8+T cells number is reduced in COVID-19 patients, their cytotoxic activity maintained by producing proinflammatory molecules [15]. At the same time, severe patient showed a decrease in number of CD4+T cells and helper CD4+T cells number was enhanced in recovering patients along with elevated levels of Granzyme A, Granzyme B and Perforin [16-18].

This review focuses on the response of innate and adaptive immune cells in the context of SARS-CoV-2 infection that might be helpful for understanding the overall scenario of host defense mechanism against COVID-19.

## Innate immune cell response to SARS-CoV-2

Innate immunity is non-specific to any particular pathogen; by screening the conserve characteristics or features of pathogen, it is quickly activated to destroy invaders. Innate immunity comprises of wide range of lymphoid and myeloid cell types. Natural killer cells were found significantly higher in blood of COVID-19 patients compared to healthy and its count reduced gradually during severity of the disease [19]. Initial contact of SARS-CoV-2 with host cells via ACE-2 (angiotensin-converting enzyme 2) receptor expressed on cells in blood vessels, gastrointestinal tract, heart, kidney and specifically epithelial cells of alveoli [3, 20, 21]. SARS-CoV-2 infection has down regulated ACE2 expression, which might compromise the innate response of COVID-19 patients [4]. NKG2A (NK group 2 member A) is a heterodimeric inhibitory receptor prominently expressed in NK (Natural killer) cell and activation of NKG2A transmits the inhibitory response by suppressing cytotoxicity and cytokine secretion of these cells [22]. NKG2A over expression has been recently demonstrated in NK cells and CD8+T cells of SARS-CoV-2 infected patients which might be another reason for suppression of immune response against COVID-19 [19]. In COVID-19 patients GM-CSF (Granulocyte-macrophage colony-stimulating factor) producing CD14+HLA-DR inflammatory monocytes (IM) have been found higher in number by flow cytometric analysis [23]. Single cell transcriptomic data also revealed the cellular expansion of CD14+IL 1 $\beta$ + monocyte and detected low IL 1 $\beta$  level in severe COVID-19 patients [24]. Data suggested elevated pro inflammatory cytokine like IL 6, IL 7, IL 2, IFNY, Interferon gamma-induced protein 10 (IP 10), Monocyte chemo attractant protein-1 (MCP-1), Macrophage Inflammatory Proteins (MIP), Granulocyte-colony stimulating factor (GCSF) and GM-CSF have been found in COVID-19 patient, in addition, IL 6 was shown to be correlated with disease severity [1, 25, 26]. In vitro study has indicated SARS-CoV infection could induce macrophage pyroptosis and inflammasome activation via NLRP3 dependent pathway [27, 28]. Due to increased level of IFNY and GM-CSF in the blood plasma of a COVID-19 patient, signal transmitted via JAK/STAT pathway for the activation of interferon-stimulated genes which can enhance the amount of other cytokines. During MERS-CoV and SARS-CoV infections, the signals are transmitted via MAPK pathways by phosphorylating the intermediate protein sub-units [15, 28, 29].

Several studies have mentioned about NK cells association with COVID-19 severity and its number being less in blood in these patients [11-14]. Single cell transcriptomic study revealed equal presentation of these cells on the basis of some signature genes in lungs of both patient as well as healthy controls [30]. Number of CXCR3 ligand-producing monocytes was found higher and CXCR3 ligand was increased in lung tissue of SARS-CoV-2 infected patients which might help in migration of other immune cells to target. In other respiratory viral infection like influenza, majority of lung NK cells were nonresident and they were infiltrated from peripheral blood [30-32]. This might suggest the recruitment of NK cells in lungs of COVID-19 patient facilitated by CXCR3 pathway.

Percentage of CD16 expressing NK cells were decreased in peripheral blood upon SARS-CoV-2 infection [33]. In SARS-CoV-1 infection, Killer-Immunoglobulin Receptors (KIR) positive NK cells frequency was less compared to healthy individuals [34]. Collectively, data suggested in COVID-19 patient there was impairment in maturation or infiltration of NK cells as CD16 and KIR were essential for

development as well as recruitment of these cells to lungs. Peripheral blood NK cells of COVID-19 patients have showed decreased expression of Granulysin, Granzyme B, IFNY, TNF $\alpha$ , CD107a and Ksp37 which suggests an impaired cytotoxic nature [14, 35]. Non-cytotoxic innate lymphoid cells (ILCs) have not been studied well in the context of COVID-19. ILC 2 produces IL 13 which helps in recruitment of macrophages and induced hyperactivity in influenza infection [36].

Golonka et al. has emphasized the positive impact of TLR5 on SARS-CoV-2 infection [37]. They proposed TLR5 can induce cytokines which might restore the impaired response of innate cell. Data shows SARS-CoV infection induces up regulation of TLR7 in monocytes but it is not clear whether it can possibly trigger innate immune response against COVID-19 [38].

HLA alleles code for cell surface protein displayed on antigen presenting cells like dendritic cells and macrophages of innate immune system. Those are critical components of the viral antigen presentation process that confer severity of disease as well as differential viral susceptibility [10]. Individuals with the HLA-B\*46:01 genotype show high risk of SARS-CoV infection [39]. Several other HLA alleles including HLA-B\*54:01, HLA-B\*39:01 and HLAB\*13:01 were identified in severe acute respiratory syndrome and their association with disease has been well-explained [40]. Recently, in silico analysis revealed that HLA-B\*15:03 has highest predicted binding peptide and HLA-B\*46:01 has least possible conserve peptide for SARS-CoV-2 [10].

## T cell response to SARS-CoV-2

T lymphocytes play major role in viral infection, where CD8+T cells clear out the infected cells and CD4+T cells convey the signal to B cells for antibody formation, which together minimize viral burden. Overall reduction in number of CD4 and CD8+T cells was observed in SARS-CoV-1 infection; several current studies emphasize the occurrence of T cells lymphopenia in moderate and severe COVID-19 cases [41-43]. The percentage of CD8+T memory cells was higher than that of CD4+T memory cells in SARS-CoV survivors and it suggested virus specific T cells could confer long-term immunity [44]. Patients recovered from COVID-19 showed robust T cells response against viral N (nucleocapsid), M (membrane) and S (spike) proteins and one third cases of total recovery have expressed N specific T cell response [45]. Flow cytometric analysis revealed that 1.3% and 1.4% cells were COVID-19 specific CD8T and CD4+ T cells in patient respectively. Limited data available on phenotyping of CD4 and CD8+T cells by considering various surface markers. According to the percentage of CCR7 and CD45RA expressing T cells, CD4 central memory or CD8 effector memory and effector memory with RA positive cells were predominant in SARS-CoV-2 infected patients [46]. Spike protein specific response induced the expression of CD154 and CD137 on T cells and this T cells population was found in 83% of SARS-CoV-2 infected cases with increased expression of HLA-DR, CD38, and Ki-67 [47]. Certain functional and phenotypical modifications of T cells due to SARS-CoV-2 infection are analyzing continuously and some reports have suggested enhanced frequencies of activated T cells with expression of CD25, CD38, CD44, CD69, HLA-DR and Ki-67 in patient with COVID-19 [16-18, 24, 30, 45, 47]. CD8+T cells response seems to be stronger than CD4+T cells in SARS-CoV-2 infection on the basis of cytotoxic activity and effector like function [16, 17]. Follicular helper CD4+T cells number was enhanced in

recovering patients along with elevated levels of Granzyme A, Granzyme B and Perforin [16, 18]. Xu et al. found a significant decrease in the number of both CD8 and CD4+T cells in SARS-CoV-2 infected patients and also confirmed the cells were highly activated by expressing HLA-DR (CD4 3.47%) as well as CD38 (CD8 39.4%) [15]. In addition, high proportions of CCR6+Th17 cells were also identified in COVID-19 positive cases [15].

CD8+T cells found were rich in cytotoxic granules releasing Granulysin (64.2%), Perforin (31.6%) and both Granulysin and Perforin (30.5%) in COVID-19 positive patients [15]. As shown by recent study, increased level of pro inflammatory molecule reported due to COVID-19 infection and enhanced level of IFN $\gamma$ , IL 1 $\beta$ , CCL2 and CXCL10 strongly induced activation of Th1 cell function [1]. Another study highlighted that SARS-CoV-2 infection has also increased Th2 response by secreting IL 10 and IL 4, which are anti-inflammatory in nature [48]. TNF $\alpha$ , CXCL10 and CCL2 concentration was seen higher in infected patients requiring ICU admission compared to less severe patients [48]. COVID-19 infected patient showed increased Th17 response due to the excessive presence of IL 6 driven by viral infection. So increased IL 6 level in blood plasma plays an indicator for disease severity and might be used as a prognostic indicator for COVID-19 [49-51]. Chen et al recently reported that IFN $\gamma$  expressing CD4+T cells were markedly lower in severe case than moderately infected patients [42].

### B cells and COVID- 19

B cells show humoral immune response by producing antibody to clear the pathogen and memory cells to prevent reinfection. SARS-CoV-2 infection induces B cell response by generating virus specific IgG, IgM and IgA along with neutralizing IgG antibodies after certain days of infection [52]. Huang et al. well described the kinetics of antibody reaction against SARS-CoV-2 [52]. Study declares, within 7 to 14 days after the onset of COVID-19 symptoms, virus specific antibody is produced in a detectable range in the blood plasma of patient [53-55]. Common antibody detected are against internal nucleocapsid protein and the external spike glycoprotein of SARS-CoV-2 [56-58]. Binding domain of spike glycoprotein is more immunogenic and antigen binding site of antibody binds to it leads to neutralize and also blocks ACE2 to avoid viral entry to host cells [57, 59]. Receptor binding domain specific CD19 positive B cells were sorted from COVID-19 patients after 9-28 days of infection and carried for gene sequencing. From the sequencing data, it was demonstrated that the monoclonal antibodies had relatively no or less somatic mutations with diverse repertoire and variable binding reactivity [57].

### Conclusion

COVID-19 pandemic brings a high degree of severity in individuals with compromised immune system. Immune cells with limited function represent the major barrier for COVID-19 recovery. During prolonged infection of SARS-CoV-2, cytotoxic T cells and NK cells start losing their cytotoxic properties that are required for eradicating viruses.

Cytokine storm is an unfavorable circumstance that occurs due to robust inflammatory reactions in case of SARS-CoV-2 infection [60, 61]. This condition could be considered as major contributor to inflammation and excessive swelling in respiratory tract. Several reports have validated, cytokine storm is mostly obtained from monocytes, macrophages

and T cells by secreting excess amount of cytokines like TNF $\alpha$ , IL 6 and IL 10 [60, 61]. Based on Diao et al. investigation, secretion of cytokine has not been originated from T cells and cytokine storm might promote necrosis or apoptosis of T cells that consequently lead to a decrease in number of cells [62]. Therefore, it needs further investigation to know the sources of cytokine release and its mechanism of production in COVID-19 patients.

In vitro study demonstrated that virus specific antibody inversely correlates with viral load and positively correlates with rate of neutralization. It indicates a successful neutralization process in majority of individuals with COVID-19 and explains the greater association of higher antibody titers with more severe cases [55, 63, 64]. This study also suggests robust antibody alone is not sufficient to neutralize the viral peptide in severe cases. So further studies are needed to determine the extent of antibody response towards disease pathophysiology.

Based on availability of data, T cell exhaustion happens in COVID-19, which affects adaptive immunity by losing the effector function like differentiation, maturation and cytokine production. Over expression of NKG2A receptor induces inhibitory response in NK cells as well as CD8+T cells of COVID-19 positive patients resulting impairment in cytotoxic properties which over all compromises the anti-viral immune responses.

Questions regarding the contribution of antibody, restoration of impaired immunity and mechanism of cytokine production during COVID-19 infection are urgently required to address. Accordingly, more in vitro, ex vivo and in silico experiments with animal study by considering broad range of participants are important for the new findings regarding COVID-19 infection.

### References

1. Huang C, Wang Y, Li X, Ren L, Zhao J, et al. (2020) Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *The Lancet* 395: 497-506.
2. Wang D, Hu B, Hu C, Zhu F, Liu X, et al. (2020) Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *Jama* 323: 1061-1069.
3. de Wit E, van Doremalen N, Falzarano D, Munster VJ (2016) SARS and MERS: recent insights into emerging coronaviruses. *Nature Reviews Microbiology* 14: 523.
4. Zou Z, Yan Y, Shu Y, Gao R, Sun Y, et al. (2014) Angiotensin-converting enzyme 2 protects from lethal avian influenza A H5N1 infections. *Nat Commun* 5: 3594.
5. De Jong MD, Simmons CP, Thanh TT, Hien VM, Smith GJ, et al. (2006) Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia. *Nature medicine* 12: 1203-1207.
6. Huang KJ, Su IJ, Theron M, Wu YC, Lai SK, et al. (2005) An interferon- $\gamma$ -related cytokine storm in SARS patients. *Journal of medical virology* 75: 185-194.
7. D'Elia RV, Harrison K, Oyston PC, Lukaszewski RA, Clark GC (2013) Targeting the "cytokine storm" for therapeutic benefit. *Clin Vaccine Immunol* 20: 319-327.
8. Schoggins JW, Wilson SJ, Panis M, Murphy MY, Jones CT, et al. (2011) A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature* 472: 481-485.
9. Channappanavar R, Fehr AR, Zheng J, Wohlford-Lenane C,

- Abrahante JE, et al. (2019) IFN-I response timing relative to virus replication determines MERS coronavirus infection outcomes. *The Journal of clinical investigation* 129: 3625–3639.
10. Nguyen A, David JK, Maden SK, Wood MA, Weeder BR, et al. (2020) Human leukocyte antigen susceptibility map for SARS-CoV-2. *Journal of virology* 94: e00510-e00520.
  11. Song C-Y, Xu J, He J-Q, Lu Y-Q (2020) COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients. *MedRxiv*.
  12. Wang W, He J, Wu S, Lie p, Huang l, et al. (2020) The definition and risks of cytokine release syndrome-like in 11 COVID-19-infected pneumonia critically ill patients: disease characteristics and retrospective analysis. *Medrxiv*.
  13. Yu L, Tong Y, Shen G, Fu A, Lai Y, et al. (2020) Immunodepletion with Hypoxemia: A Potential High Risk Subtype of Coronavirus Disease 2019. *medRxiv*.
  14. Zheng M, Gao Y, Wang G, Song G, Liu S, et al. (2020) Functional exhaustion of antiviral lymphocytes in COVID-19 patients. *Cellular & molecular immunology* 17: 533-535.
  15. Xu Z, Shi L, Wang Y, Zhang J, Huang L, et al. (2020) Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *The Lancet respiratory medicine* 8: 420-422.
  16. Thevarajan I, Nguyen TH, Koutsakos M, Druce J, Caly L, et al. (2020) Breadth of comorbid immune responses prior to patient recovery: a case report of non-severe COVID-19. *Nature medicine* 26: 453-455.
  17. Yang X, Dai T, Zhou X, Qian H, Guo R, et al. (2020) Analysis of adaptive immune cell populations and phenotypes in the patients infected by SARS-CoV-2. *medRxiv*.
  18. Zheng H-Y, Zhang M, Yang C-X, Zhang N, Wang X-C, et al. (2020) Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients. *Cellular & molecular immunology* 17: 541-543.
  19. Zheng M, Gao Y, Wang G, Song G, Liu S, et al. (2010) Functional Exhaustion of antiviral lymphocytes in COVID-19 patients. *Cell Mol Immunol* 10. <http://science.sciencemag.org/>
  20. Fukao T, Fukuda Y, Kiga K, Sharif J, Hino K, et al. (2007) An evolutionarily conserved mechanism for microRNA-223 expression revealed by microRNA gene profiling. *Cell* 129: 617-631.
  21. Zhao Y, Zhao Z, Wang Y, Zhou Y, Ma Y, et al. (2020) Single-cell RNA expression profiling of ACE2, thereceptor of SARS-CoV-2. *Biorxiv*.
  22. Wong C, Lam C, Wu A, Ip W, Lee N, et al. (2004) Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome. *Clinical & Experimental Immunology* 136: 95-103.
  23. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Akinosoglou K, Antoniadou A, et al. (2020) Complex immune dysregulation in COVID-19 patients with severe respiratory failure. *Cell host & microbe* 27: 1-9.
  24. Guo C, Li B, Ma H, Wang X, Cai P, et al. (2020) Tocilizumab treatment in severe COVID-19 patients attenuates the inflammatory storm incited by monocyte centric immune interactions revealed by single-cell analysis. *BioRxiv*.
  25. Wan S, Yi Q, Fan S, Lv J, Zhang X, et al. (2020) Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP). *MedRxiv*.
  26. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, et al. (2020) COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet (London, England)* 395: 1033.
  27. Chen I-Y, Moriyama M, Chang M-F, Ichinohe T (2019) Severe acute respiratory syndrome coronavirus viroporin 3a activates the NLRP3 inflammasome. *Frontiers in microbiology* 10: 50.
  28. Shi C-S, Nabar NR, Huang N-N, Kehrl JH (2019) SARS-Coronavirus Open Reading Frame-8b triggers intracellular stress pathways and activates NLRP3 inflammasomes. *Cell death discovery* 5: 1-12.
  29. Lim YX, Ng YL, Tam JP, Liu DX (2016) Human coronaviruses: a review of virus–host interactions. *Diseases* 4: 26.
  30. Liao M, Liu Y, Yuan J, Wen Y, Xu G, et al. (2020) The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing. *MedRxiv*.
  31. Chu H, Chan JF-W, Wang Y, Yuen TT-T, Chai Y, et al. (2020) Comparative replication and immune activation profiles of SARS-CoV-2 and SARS-CoV in human lungs: an ex vivo study with implications for the pathogenesis of COVID-19. *Clinical Infectious Diseases* 2020: ciaa410.
  32. Gasteiger G, Fan X, Dikiy S, Lee SY, Rudensky AY (2015) Tissue residency of innate lymphoid cells in lymphoid and nonlymphoid organs. *Science* 350: 981-985.
  33. Wang F, Nie J, Wang H, Zhao Q, Xiong Y, et al. (2020) Characteristics of peripheral lymphocyte subset alteration in COVID-19 pneumonia. *The Journal of infectious diseases* 221: 1762-1769.
  34. National Research Project for SARS BG (2004) The involvement of natural killer cells in the pathogenesis of severe acute respiratory syndrome. *American Journal of Clinical Pathology* 121: 507-511.
  35. Wilk AJ, Rustagi A, Zhao NQ, Roque J, Martinez-Colon GJ, et al. (2020) A single-cell atlas of the peripheral immune response to severe COVID-19. *medRxiv*.
  36. Chang Y-J, Kim HY, Albacker LA, Baumgarth N, McKenzie AN, et al. (2011) Innate lymphoid cells mediate influenza-induced airway hyper-reactivity independently of adaptive immunity. *Nature immunology* 12: 631-638.
  37. Golonka RM, Saha P, Yeoh BS, Chattopadhyay S, Gewirtz AT, et al. (2020) Harnessing innate immunity to eliminate SARS-CoV-2 and ameliorate COVID-19 disease. *Physiol Genomics* 52: 217-221.
  38. Hu W, Yen Y-T, Singh S, Kao C-L, Wu-Hsieh BA (2012) SARS-CoV regulates immune function-related gene expression in human monocytic cells. *Viral immunology* 25: 277-288.
  39. Lin M, Tseng H-K, Trejaut JA, Lee H-L, Loo J-H, et al. (2003) Association of HLA class I with severe acute respiratory syndrome coronavirus infection. *BMC Medical Genetics* 4: 9.
  40. Ng MH, Lau K-M, Li L, Cheng S-H, Chan WY, et al. (2004) Association of human-leukocyte-antigen class I (B\* 0703) and class II (DRB1\* 0301) genotypes with susceptibility and resistance to the development of severe acute respiratory syndrome. *Journal of Infectious Diseases* 190: 515-518.
  41. He Z, Zhao C, Dong Q, Zhuang H, Song S, et al. (2005) Effects of severe acute respiratory syndrome (SARS) coronavirus infection on peripheral blood lymphocytes and their subsets. *International journal of infectious diseases* 9: 323-330.
  42. Chen G, Wu D, Guo W, Cao Y, Huang D, et al. (2020) Clinical and immunological features of severe and moderate coronavirus

- disease 2019. The Journal of clinical investigation 130: 2620-2629.
43. Nie S, Zhao X, Zhao K, Zhang Z, Zhang Z, et al. (2020) Metabolic disturbances and inflammatory dysfunction predict severity of coronavirus disease 2019 (COVID-19): a retrospective study. medRxiv.
  44. Ng O-W, Chia A, Tan AT, Jadi RS, Leong HN, et al. (2016) Memory T cell responses targeting the SARS coronavirus persist up to 11 years post-infection. Vaccine 34: 2008-2014.
  45. Dong C, Ni L, Ye F, Chen M-L, Feng Y, et al. (2020) Characterization of anti-viral immunity in recovered individuals infected by SARS-CoV-2. medRxiv.
  46. Weiskopf D, Schmitz KS, Raadsen MP, Grifoni A, Okba NM, et al. (2020) Phenotype of SARS-CoV-2-specific T-cells in COVID-19 patients with acute respiratory distress syndrome. medRxiv.
  47. Braun J, Loyal L, Frensch M, Wendisch D, Georg P, et al. (2020) Presence of SARS-CoV-2 reactive T cells in COVID-19 patients and healthy donors. medRxiv.
  48. Zhang C, Wu Z, Li J-W, Zhao H, Wang G-Q (2020) The cytokine release syndrome (CRS) of severe COVID-19 and Interleukin-6 receptor (IL-6R) antagonist Tocilizumab may be the key to reduce the mortality. International journal of antimicrobial agents 2020: 105954.
  49. Chen L, Liu H, Liu W, Liu J, Liu K, et al. (2020) Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia. Zhonghua jie he he hu xi za zhi= Zhonghua jiejie he huxi zazhi= Chinese journal of tuberculosis and respiratory diseases 43: E005-E.
  50. Henry BM, De Oliveira MHS, Benoit S, Plebani M, Lippi G (2020) Hematologic, biochemical and immune biomarker abnormalities associated with severe illness and mortality in coronavirus disease 2019 (COVID-19): a meta-analysis. Clinical Chemistry and Laboratory Medicine (CCLM) 58: 1021-1028.
  51. Ulhaq ZS, Soraya GV (2020) Interleukin-6 as a potential biomarker of COVID-19 progression. Médecine et Maladies Infectieuses 50: 382-383.
  52. Huang AT, Garcia-Carreras B, Hitchings MD, Yang B, Katzelnick LC, et al. (2020) A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease. medRxiv.
  53. Haveri A, Smura T, Kuivanen S, Österlund P, Hepojoki J, et al. (2020) Serological and molecular findings during SARS-CoV-2 infection: the first case study in Finland, January to February 2020. Eurosurveillance 25: 2000266.
  54. Lou B, Li T-D, Zheng S-F, Su Y-Y, Li Z-Y, et al. (2020) Serology characteristics of SARS-CoV-2 infection since exposure and post symptom onset. European Respiratory Journal 2020: 2000763.
  55. Okba NM, Müller MA, Li W, Wang C, GeurtsvanKessel CH, et al. (2020) Severe acute respiratory syndrome coronavirus 2-specific antibody responses in coronavirus disease 2019 patients. Emerging infectious diseases 26.
  56. Amanat F, Krammer F (2020) SARS-CoV-2 vaccines: status report. Immunity 52: 583-589.
  57. Ju B, Zhang Q, Ge X, Wang R, Yu J, et al. (2020) Potent human neutralizing antibodies elicited by SARS-CoV-2 infection. BioRxiv.
  58. To KK-W, Tsang OT-Y, Leung W-S, Tam AR, Wu T-C, et al. (2020) Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: an observational cohort study. The Lancet Infectious Diseases 20: 565-574.
  59. Wu F, Wang A, Liu M, Wang Q, Chen J, et al. (2020) Neutralizing antibody responses to SARS-CoV-2 in a COVID-19 recovered patient cohort and their implications.
  60. Kany S, Vollrath JT, Relja B (2019) Cytokines in Inflammatory Disease. International Journal of Molecular Sciences 20: 6008.
  61. Maltese NM, Paola Lucia Minciullo, Antonino Catalano, Giuseppe Mandraffino, Marco Casciaro, et al. (2016) Inflammaging and Anti-Inflammaging: The Role of Cytokines in Extreme Longevity. Arch Immunol Ther Exp (Warsz) 64: 111-126.
  62. Diao B, Wang C, Tan Y, Chen X, Liu Y, et al. (2020) Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19). Frontiers in Immunology 11: 827.
  63. Zhao J, Yuan Q, Wang H, Liu W, Liao X, et al. (2020) Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019. Clinical Infectious Diseases 2020: ciaa344.
  64. Wölfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, et al. (2020) Virological assessment of hospitalized patients with COVID-2019. Nature 581: 465-469.

**Copyright:** ©2020 Archana Tripathy. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.