

Review on Goat Mastitis and Associated Bacterial Zoonoses in Raw Milk from Mastitis Infected Dairy Goat

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Abstract

Goat milk has medicinal value for children and contributes much more for the wellbeing of human baby. Goat milk provides more nutritional value than dairy cow's milk. Despite the large number of goats and their contributions to the livelihood of the farmers, goat productivity is low due to prevalence of different diseases. One of the major diseases that affect the dairy goats is mastitis. Mastitis is an inflammation of the mammary gland, caused by over 150 different contagious or environmental micro-organisms. Mastitis in goats constitutes an enormous animal health problem. In addition to causing health problems, inflammation of the mammary glands can also cause poor quality of milk. Milk produced by goats with mastitis presents a serious risk in terms of public health as it can be linked to milk-borne diseases for humans. Raw milk from Mastitis infected goat is usually colonized by a variety of many zoonotic pathogens such as enterohaemorrhagic *Escherichia coli*, *Salmonella typhimurium*, *Listeria monocytogenes* and *Staphylococcus aureus*, *Brucella melitensis*, *Mycobacterium bovis*. These pathogens in milk have been linked to mastitis milk. These zoonotic organisms may lead to health problems in the human population such as tuberculosis (TB), brucellosis, haemorrhagic enteritis, salmonellosis and food poisoning. Transmission of pathogens from animals to humans can occur via consumption of milk, especially when these products are consumed raw.

Keywords: Bacteria, Mastitis, Milk, Zoonoses

1. Introduction

Goat production is one of the low resources demanded and efficient farming types, since goats have broad feeding habit, adaptation to unfavorable environmental conditions, low cost of maintenance, inherent suitability for small scale production and their short reproductive cycle. These provide goats with comparative advantage over cattle and sheep to suit the circumstances of especially resource poor livestock keepers [1]. Since goats browse different variety of trees and shrubs, goat owners believed that goat milk has medicinal value for children and contribute much more for the wellbeing of human baby [2].

Milk production in goats is an active and emergent business in harsh climate areas where large ruminants cannot be reared or are difficult to rear and it largely contributes to the mainstream dairy milk production [3]. In addition to this, goat milk provides more nutritional value than dairy cow's milk [4]. Highly milk productive goats are able to produce milk as much as 20 times their body weight [5]. Goat milk is highly nutritious and has a similar nutritional profile to those of human's breast milk [6]. Despite the large number of goats and their contributions to the

livelihood of the farmers, goat productivity is low due to prevalence of different diseases and parasites [7]. One of the major diseases that affect the dairy goats is mastitis [8].

Mastitis is an inflammation of the mammary gland, caused by over 150 different contagious or environmental micro-organisms [9]. In lactating dairy goats, the inflammation of the mammary gland is one of the most common infectious diseases [10]. It occurs after several pathogens invades and colonizes the secretory tissue leading to inflammation of the mammary gland [11]. Mastitis in goats constitutes an enormous animal health problem [12].

In addition to causing hygiene and health problems, inflammation of the mammary glands can also cause economic losses due to reduced milk production, poor quality of milk [13]. Mastitis also poses a threat to human health due to the risk of transmitting zoonotic pathogens through ingestion of contaminated milk [14,15]. Milk produced by goats with mastitis presents a serious risk in terms of public health as it can be linked to milk-borne diseases for humans [16]. Though milk is an important

food product, milk and milk by-products can harbour a variety of zoonotic pathogens which cause zoonoses [17].

Zoonoses are infections that can spread from animal to man [18]. The World Health Organization (WHO) defines zoonosis as diseases that can be transmitted between humans and animals [19]. Zoonotic pathogens may be bacterium, virus, fungus or other communicable disease agents [20]. These zoonotic pathogens may contaminate milk either whilst the milk is still in the udder [21]. Raw milk is usually colonized by a variety of many zoonotic pathogens such as enterohaemorrhagic *Escherichia coli*, *Salmonella typhimurium*, *Listeria monocytogenes* and *Staphylococcus aureus*, *Brucella melitensis*, *Mycobacterium bovis* [22]. These pathogens represent an important source of food-borne pathogens [23].

These zoonotic organisms may lead to health problems in the human population such as tuberculosis (TB), brucellosis, hemorrhagic enteritis, salmonellosis and listeriosis [24]. The population that is at higher risk of being infected with this milk borne pathogens includes people who are immunocompromised, the elderly, pregnant women and children [25].

Transmission of pathogens from animals to humans can occur via consumption of milk, especially when these products are consumed raw. Raw milk and products from raw milk containing pathogenic microorganisms can enter the food chain and be responsible for food poisoning episodes among consumers [26]. The transfer of heat-stable toxins produced by mastitis-causing pathogens in milk is another serious potential concern [27]. Pasteurization reduces the number of viable micro-organisms but often does not destroy toxins produced by bacterial pathogens, hence the concern when raw milk is consumed or when pasteurization is faulty [28]. While most milk in developed countries is tested for high SCC and pasteurized before it reaches consumers, in developing countries, milk is often consumed unpasteurized [29]. For this reason, utilization of both raw untreated milk and raw milk cheeses has frequently been associated with foodborne illness [30].

The Objective of this paper is to;

- review the most recent literature about goat mastitis and how intramammary infections in this species affect the milk safety.
- review the public health importance of pathogenic agents involved in the etiology of mastitis.

2. Literature Review

2.1. Mastitis Definition

Mastitis is a parenchymal inflammation of the mammary gland, characterized by physical, chemical, and usually bacteriological changes in milk and pathological changes in glandular tissues [31]. Mastitis cases can be divided on the basis of origin into environmental and contagious [32]. Environmental mastitis is caused by bacterial microorganisms from the surrounding environment, referred to as environmental pathogens, whereas contagious mastitis is due to spread from other infected quarters [33]. Environmental mastitis is caused by microorganisms present in the animals surrounding area. These pathogens infect the udder via the teat canal [34]. The main reservoirs of contagious

pathogens are the rectal, rumen, and genital areas in addition to the mammary gland [35]. The infection is spread during milking time when infected milk contacts an uninfected mammary gland and bacteria then penetrate the teat canal [36].

There are many microorganisms that cause mastitis in goats [37]. Unappropriate milking techniques and unsuitable hygiene conditions increases the infections [38]. Mastitis is a complex disease resulting from the interaction between the agent, animal and the environment, associated with the presence of microorganisms in most cases [39]. It is an important animal health and public health problem, with great economic repercussion in practically every country in the world [40]. A greater number of zoonotic pathogens including *brucella (b.) Melitensis*, *campylobacter spp.*, *escherichia coli*, *mycobacterium spp.*, *salmonella spp.*, and *staphylococcus aureus* were isolated from milk [41]. The prevalence of these pathogens in the milk was associated with the occurrence of diseases in the animals including goat mastitis [42].

2.2. Epidemiology

2.2.1. Geographical Distribution

Mastitis is the most prevalent production disease in dairy herds worldwide [43]. It is a well-documented disease with a heavy burden in both, developed and developing countries [44]. It is a global health problem of lactating animals and it considered one of the most important diseases of domestic animals, caused by several etiologic agents [45].

The study of a disease in a population requires an understanding of the relationships between organisms, hosts and their environment [46]. Moreover, epidemiological investigations include an increase in the understanding of the pathogenesis, maintenance and for infectious agents transmission of disease also when and where a disease may occur to enable the development of suitable control techniques where the delay in epidemiological interference discovery is probably due to the lack of long-term surveys on the incidence of infections as reported [47].

2.2.2. Risk Factors

Environmental risk factors: - Udder disease is affected by a number of external environmental factors (management, manner of milk removal and milking technology, season) [48]. It also including, milking hygiene, management practice, and geographical locality which influence the type and the frequency of isolation of organisms cause mastitis and in particular the routine of machine milking [49,50]. Predisposing factors such as poor management and hygiene, teat injuries and inefficient use of milking machines are known to accelerate the entry of infectious agents and the course of the disease [51]. The high percentage of the subclinical mastitis could be due to a lack of hygiene and to the practice of traditional breeding of extensive type, which favors diseases [16].

Host risk factors: - Internal factors (include physiological status of the body, like stage and order of lactation, oestrus, udder shape, feeding, number of lactation days) are important for the occurrence of mastitis [52]. High stocking density, particularly in intensively managed herds/flocks or during the suckling period, may result in large air concentrations of total microorgan-

isms, coliform bacteria and staphylococci [53]. These effects are probably associated with incorrect ventilation and high relative humidity. The multiplication of various bacteria on the skin (and in the litter) can be subsequently enhanced [54].

Pathogenic Risk Factors: Several pathogens can cause mastitis, *Staphylococcus* spp. are the most frequently diagnosed causal microorganisms of intramammary infection (IMI) goats and it secretes several toxins contributing to the pathogenesis of mastitis and also plays a role in food borne diseases; even with pasteurized milk because of the thermo-stable enterotoxins [55]. Staphylococcal alpha-hemolysin or alpha-toxin is the most studied and characterized cytotoxin, and is considered a main pathogenicity factor because of its hemolytic, dermonecrotic and neurotoxic effects [56].

2.2.3. Transmission

The group of environmental agents is present in organic matter as soil and feces, in the bed of animals, in water and in the air. The infection occurs mainly in the inter-milking period, but may also occur during milking [57]. Infections by contagious agents occur predominantly during milking, and microorganisms opportunistically invade the mammary gland, causing an immediate inflammatory response [9]. Udder massage and stripping induce air intakes leading to impact [50]. Cluster removal by the milker may also induce impact, since it is often performed without previous vacuum cutting off (the automatic cluster removal is developing) [58]. Bacteria are also transported passively by liners. However, the intra mammary infections (IMI) prevalence's do not seem to be significantly different between dairy (hand or machine milked) and meat flocks [59]. Transmission is also possible by "milk-robber" lambs (buccal carriage) and may be important for staphylococci, Pasteurellaceae, parapoxvirus (contagious ecthyma) [60]. Infection of the udder usually takes place directly through teat canal [61].

2.3. Pathogenesis

The first line of udder defense against pathogens is the teat end. It is open and closed by a sphincter composed of smooth muscles that serves as a barrier to prevent pathogens from entering the canal and prevent milk from escaping [62]. The teat canal is lined with the stratified squamous epithelium, which creates keratin to fill the canal between 30 min and two hours after milking [63]. This time span may vary, creating an opportunity for bacteria near the opening to enter the teat canal [64]. In order to establish the infection, the etiologic agent must surpass the terminal portion of the ceiling, since the integrity of the ceiling is the first line of defense [62]. The bacteria enter the gland through the streak canal and multiply within the udder cells or in the teat duct [11].

The pathogenesis of *Staphylococcus aureus* mastitis is very complex [66]. It is associated with various surface proteins and virulence factors that are differentially expressed at various phases of the infection [67]. This process entails three key steps, that is adhesion, invasion and evasion [68]. In brief, the first step in the pathogenesis process is adhesion to epithelial cells and extracellular matrix, which permits the bacteria to avoid being flushed out of the udder from milk flux pressure [69]. In the second step of this process, *Staphylococcus aureus* again expresses different

virulence factors to establish infection by invasion into host cells and tissues [70]. The final step in the pathogenetic process is an evasion of the host immune response ((Nesse et al., 2023). Here, *S. aureus* escapes the host immune response by producing the various virulence factors that helps it not only to evade but also modulate the host immune response in its favor [69]. This mechanism is not only employed by *Staphylococcus aureus* but other mastitis causing bacteria such as *Streptococcus* spp. [71]. Therefore, adherence of microbial agent to teat epithelial tissue permits them to invade or penetrate this protective barrier and migrate to the teat duct [70].

2.4. Clinical Signs

Mastitis is a disease that occurs in several different forms. Generally, in animal, mastitis is divided into clinical and sub-clinical forms [72]. In cases where there are no visible changes in appearance of milk and udder but the milk composition is altered with presence of bacteria accompanied by decreased milk production then subclinical mastitis is diagnosed [73]. Subclinical mastitis is the term used for the inflammation of udder that cannot be detected by clinical methods such as inspection, palpation and organoleptic examination [74]. It is the most important diseases which cause change in the milk composition and any change in its percentage in turn affect the suitability of milk processing and the quality of its products [75].

Clinical mastitis is the term used for inflammation of the mammary gland that are present with obvious symptoms and is characterized by visible abnormalities in the udder or milk of infected animal [76]. Clinical sign in case of clinical mastitis is characterized mainly by appearances of changes in the milk such as flakes and clots and presence of signs of inflammation on the mammary glands such as swelling, heat, pain, and edema [77]. Systemic signs on the animal infected with clinical mastitis including fever, rapid pulse, appetite loss, dehydration, and depression [78].

2.5. Diagnosis

Diagnostic procedures used in mastitis include clinical examination, bacteriological tests, cytological examination of milk (direct by using fluoro-optoelectronic counters and microscopic cell counting, indirect by using the California Mastitis Test or the White Side Test), measurement of milk electrical conductivity and imaging techniques (ultrasonography, endoscopy, infrared thermography) [79]. Appropriate samples for the relevant diagnostic techniques include udder or teat skin swabs, teat duct material, milk, mammary tissue and blood samples [80].

Diagnosis of clinical mastitis is usually straight forward, based on findings of the clinical examination (swollen and painful udder, abnormal milk, high rectal temperature, lameness on the side of the affected gland) [81]. Further diagnostic tests (especially microbiological examinations) will support etiological diagnosis of mastitis, which is important for effective treatment [82]. In contrast, diagnosis of subclinical mastitis requires application of specific tests [83]. California Mastitis Test (CMT) and Somatic Cell Count (SCC) are regarded as the best indirect tests to diagnose intra-mammary infections (IMIs) in goats, when they are interpreted correctly [84]. Different threshold values for somatic cell counts have been proposed [85].

2.6. Treatment

Mastitis can be treated by the use of antibiotics through intramuscular as well as by intra mammary route followed by the identification of responsible pathogen as well as antibiotic sensitivity test to ensure proper treatment [86]. Microorganisms associated with mastitis in dairy goats are commonly controlled with antibiotics [87]. Dry off treatment has the advantage of antibiotics being used when the animal is not being milked, so that there is no milk loss and no antibiotic contamination of the bulk tank milk [88].

2.7. Control and Prevention

The fight against mastitis should not be limited to treating isolated clinical episodes, but also requires the surveillance and control of both, clinical and subclinical mastitis [89]. The knowledge of risk factors and etiological agents involved are also important in order to recommend specific and efficient control measures for both clinical and subclinical mastitis [45]. Farm management systems, milking management practices, breed considered or technical skills of the investigators are among the factors associated with the variability in the prevalence of goat mastitis between research reports [90]. The main control principles include: sound husbandry practices and sanitation, post milking teat dip, treatment of mastitis during non-lactating period, and culling of chronically infected animals [91].

Prevention of mastitis in small ruminants can be described upon programs that include vaccination, culling, application of good husbandry control and proper maintenance of milking machines, better milking routine without over milking and removal of clusters without impacts, and hygiene after milking [92]. Improved techniques depend on a better understanding of the nature of predisposing factors and breeding for resistance are the approach that is considered as a sustainable method for mastitis control [34].

2.8. Public Health Importance of Goat Mastitis Associated Bacterial Zoonoses

Contaminated milk can also serve as sources of a number of milk-borne infections to humans [93]. Milk produced by goats with mastitis presents a serious risk in terms of public health as it can be linked to milk-borne diseases for humans [16]. Some mastitic milk carries bacteria that can cause severe human illness. Pasteurization reduces the number of viable microorganisms but often does not destroy toxins produced by bacterial pathogens, hence the concern when raw milk is consumed or when pasteurization is faulty [94,95]. The transfer of heat-stable toxins produced by mastitis-causing pathogens in milk is another serious potential concern [96]. The main pathogens causing infections and toxins related to the consumption of mastitic milk and its derivatives are *Staphylococcus aureus*, *Salmonella* spp., *Escherichia coli*, *Mycobacterium bovis* and *Brucella melitensis* [28].

Staphylococcus aureus are the most frequently diagnosed causal microorganisms of IMI in goats. Intramammary infections caused by these pathogens need to be treated with particular caution, because this bacterium is responsible for both acute clinical mastitis (gangrenous mastitis) and subclinical mastitis [13]. This pathogen can be found widely distributed in animals,

and it is a contagious pathogen that can be transmitted from doe to doe during unhygienic milking procedures [97]. The possible contamination by *Staphylococcus aureus* in raw milk might occur from infected mammary glands [98]. The toxin, because it is thermostable, can remain in the food even after heat treatment, favoring the occurrence of intoxication, characterized clinically by nausea, emesis, malaise, general weakness, diarrhea, headache and abdominal pain [84].

Salmonellosis is an infectious disease of humans and animals which is caused by the bacteria of genus *Salmonella* and is clinically characterized by septicemia, acute enteritis or chronic enteritis [99]. *Salmonella* are Gram negative motile bacilli and amongst the leading causes of food borne illness in humans [100]. *Salmonella* spp. has a low prevalence in the case of mastitis in goats, however, salmonellosis is considered to be the disease most commonly associated with outbreaks of food poisoning [101]. Currently, more than 2,000 serotypes of *Salmonella* species are described, with *Salmonella enterica* serotype Typhimurium being the most prevalent in infections for human and domestic animals [102]. *Salmonella* spp. may be excreted directly in the milk by an infected animal [103].

E. coli is a normal inhabitant of the intestines of animals and humans but its recovery from food may be of public health concern due to the possible presence of enteropathogenic and/or toxigenic strains which lead to severe gastrointestinal disturbance [104]. While most strains of *E. coli* do not cause disease in humans, some are known to cause severe illness due to the production of toxins and/or other virulence factors [105]. In recent years, new pathogenic bacteria have emerged throughout the food chain [106]. It has been reported, for instance, that new milk-borne bacterial pathogens with extremely severe health effects, such as *E. coli* O157:H7, have emerged [107]. *E. coli* O157 is the most common type of *E. coli* infection that cause intestinal infections, urinary infections, septicemia, meningitis, and other infections [108]. Contamination of milk with pathogenic *E. coli* occurs through faecal material present on teats and udder, or from the environment [25]. Faecal contamination of udders is one of the risk factors triggering pathogens to enter the raw milk [109].

Tuberculosis in small ruminants is characterized by progressive cachexia, dry, short and repetitive cough, mastitis and infertility, and localized or generalized lymphadenomegaly may occur. It is mainly caused by *Mycobacterium bovis*, although *M. avium* and *M. tuberculosis* have been isolated occasionally [110]. Tuberculosis (TB) is also a leading zoonotic infectious cause of human death worldwide, which is transmitted to humans through consumption of raw, unpasteurized or contaminated milk, dairy products and other animal products from infected animals (e.g. cattle, goat, deer, buffalo, sheep and camel) [111]. Humans infected with open tuberculosis due to *M. bovis* can transmit the bacteria to animals via the aerogenous route by spitting or coughing [112]. The symptoms in humans are a cough, fever, sputum that in the advanced stage of the disease can present blood, difficulty breathing and progressive weight loss [113].

Brucellosis is one classical example of milk-borne infection, *Brucella* spp being transmitted from goats to humans either through direct contact or through the milk of the infected ani-

mal, particularly since the appearance and taste of the milk are rarely affected by the presence of the bacteria [106]. Approximately two thirds of acute natural *B. melitensis* infections of goats during pregnancy lead to infection of the udder and excretion of the bacteria in milk during the subsequent lactation [114]. Progressively, intermittent shedding of the agent in milk occurs in animals with persistent infection of the udder [115]. *Brucella melitensis* may cause inflammation of the mammary tissue, which is the most probable cause of reduced milk production in infected animals [116]. Once transmitted to humans, *Brucella* is responsible for a type of granulomatous hepatitis or an acute febrile illness which can, at times, persist and progress to a chronically incapacitating disease with serious complications [117].

3. Conclusion and Recommendations

Mastitis is a significant disease of dairy farms throughout the world. The diagnosis of mastitis in goats is similar to that for cows and other animals. Subclinical mastitis is detectable by the monitoring of somatic cell counts but it needs careful interpretation due to the higher rate of epithelial cell sloughing and the presence of cytoplasmic masses in goat milk. Careful diagnosis and treatment of mastitis in goats affords the best opportunity for a successful outcome when therapy is required. The prevention of mastitis through the establishment of good husbandry practices, sanitation, sound milking procedures including post-milking-teat-dipping and treatment during the non-lactating period, and culling of chronically infected can alleviate the loss due to caprine mastitis. The major obstacle in treating mastitis is antibiotic resistance; therefore, susceptibility test should be taken before treatment. Zoonotic diseases pose a serious threat to human health in developing countries.

Based on the above conclusive remarks, the following points are forwarded as recommendation.

- The periodic assessment of preventive measures for mastitis should be practiced.
- Maintaining clean and hygienic environment around animals.
- Vaccines protecting against major pathogens like *Staphylococcus Aureus* should be developed.
- Further studies on the status, distribution and the risks of milk-borne infections should be properly assessed and their control and prevention methods should be followed.

Declarations

Author's Contributions

Mohammed Bedruddin: Read and approved the final version of the manuscript.

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Ethical Approval and Consent to Participate

The protocol of my current review whose approved by college of veterinary medicine veterinary public health department of Haramaya University, Ethiopia and the ethical clearance who waived due to no major involvement of humans and animal subject welfare of ethical issues.

Competing Interests

The author declare that they have no competing interests.

References

1. Zewdie, B., Urge, M., Tadesse, Y., & Gizaw, S. (2020). Arab Goat Husbandry, Breeding Practices and Marketing Systems in Western Lowland of Ethiopia. *East African Journal of Veterinary and Animal Sciences*, 4(1), 27-40.
2. Hidosa, D., & Hailu, S. (2019). Goat Feed Resource Inventory and Feed Balance in Hamer and Bena-Tsemay Woreda of South Omo Zone, South Western Ethiopia.
3. Muzammil, I., Saleem, M. I., Aqib, A. I., Ashar, A., & Mahfooz, S. A., et al. (2021). Emergence of pathogenic strains of *Staphylococcus aureus* in goat milk and their comparative response to antibiotics. *Pakistan Journal of Zoology*, 53(5), 1659-1667.
4. Serdyukova, Y. P., Kazarova, I. G., Zakurdaeva, A. A., Gorlov, I. F., & Anisimova, E.Y., et al. (2021). Fermented goat milk product: Improvement of the production technology. In *IOP Conference Series: Earth and Environmental Science*, 677(3), 032083.
5. Gökdağ, A., Engin, S., Barbara, C., & Flaviana, G. (2020). Milking characteristics, hygiene and management practices in Saanen goat farms: a case of Canakkale province, Turkey. *Italian Journal of Animal Science*, 19(1), 213-221.
6. Prosser, C. G. (2021). Compositional and functional characteristics of goat milk and relevance as a base for infant formula. *Journal of food science*, 86(2), 257-265.
7. Kaumbata, W., Banda, L., Mészáros, G., Gondwe, T., & Woodward-Greene, M. J., et al. (2020). Tangible and intangible benefits of local goats rearing in smallholder farms in Malawi. *Small Ruminant Research*, 187, 106095.
8. Akter, S., Rahman, M. M., Sayeed, M. A., Islam, M. N., & Hossain, D. (2020). Prevalence, aetiology and risk factors of subclinical mastitis in goats in Bangladesh. *Small Ruminant Research*, 184, 106046.
9. Negash, S. (2023). Review on bovine mastitis and assessments of its associated risk factors in lactating dairy cows. *Int. J. Adv. Res. Biol. Sci*, 10(3), 59-65.
10. Ariffin, S. M. Z., Hasmadi, N., Syawari, N. M., Sukiman, M. Z., & Ariffin, M. F. T. (2019). Prevalence and antibiotic susceptibility pattern of *Staphylococcus aureus*, *Streptococcus agalactiae* and *Escherichia coli* in dairy goats with clinical and subclinical mastitis. *J. Anim. Health Prod*, 7(1), 32-37.
11. Egyedy, A. F., & Ametaj, B. N (2022). Mastitis: Impact of Dry Period, Pathogens, and Immune Responses on Etiopathogenesis of Disease and its Association with Periparturient Diseases. *Dairy*, 3, 881-906.
12. El Seedy, F., Radwan, I., WH, H., & Ramadn, A. (2023). Antimicrobial and Virulence Characteristics of *Escherichia coli* Isolated from Mastitis and Endometritis Cases in Sheep and Goats. *Journal of Veterinary Medical Research*, 30(1), 12-18.
13. Paramasivam, R., Gopal, D. R., Dhandapani, R., Subbarayalu, R., & Elangovan, M. P. (2023). Is AMR in Dairy Products a Threat to Human Health? An Updated Review on the Origin, Prevention, Treatment, and Economic Impacts of Subclinical Mastitis. *Infection and Drug Resistance*, 155-178.

14. Sugrue, I., Tobin C., Ross R. P., Stanton, C., & Hill, C. (2019). "Foodborne pathogens and zoonotic diseases," in *Raw Milk - Balance Between Hazards and Benefits*. London, UK: Academic Press, 259–272.
15. Maity, S., & Ambatipudi, K. (2021). Mammary microbial dysbiosis leads to the zoonosis of bovine mastitis: A One-Health perspective. *FEMS Microbiol. Ecol.*, 97 faa241.
16. Gabli, Z., Djerrou, Z., & Bensalem, M. (2019). Prevalence of mastitis in dairy goat farms in Eastern Algeria. *Veterinary world*, 12(10), 1563.
17. Razooqi, M. A., Mounam, M. A. A., & Saleem, H. D. (2021). Article Review: Changes in Cows' Milk Quantity and Quality Due to Bacterial Contamination. *Nveo-Natural Volatiles & Essential Oils Journal*, 8(6), 2550-2561.
18. Rahman, M. T., Sobur, M. A., Islam, M. S., Ievy, S., & Hos-sain, M. J., (2020). Zoonotic diseases: etiology, impact, and control. *Microorganisms*, 8(9), 1405.
19. Haider, N., Rothman-Ostrow, P., Osman, A. Y., Arruda, L. B., & Macfarlane-Berry, L. (2020). COVID-19-zoonosis or emerging infectious disease? *Frontiers in Public Health*. *Frontiers*, 8, 763.
20. Guarner, J., & Jean, S. (2023). One Health: The Role of Pathology as it pertains to Diagnosis of Zoonoses and Discovery of Emerging Infections. *Modern Pathology*, 36(8), 100236.
21. Chengat Prakashbabu, B., Cardwell, J. M., Craighead, L., Ndour, A. P. N., (2020). "We never boil our milk, it will cause sore udders and mastitis in our cows"-consumption practices, knowledge and milk safety awareness in Senegal. *BMC Public Health*, 20, 1-12.
22. Pradhan, A. K., & Karanth, S. (2023). Zoonoses from animal meat and milk. In *Present Knowledge in Food Safety*. Academic Press, 394-411.
23. Wei, X. & Zhao, X. (2021). Advances in typing and identification of foodborne pathogens. *Current Opinion in Food Science*, 37, 52-57.
24. Libera, K., Konieczny, K., Grabska, J., Szopka, W., & Augustyniak, A. (2022). Selected livestock-associated zoonoses as a growing challenge for public health. *Infectious disease reports*, 14, 63-81.
25. Van Den Brom, R., De Jong, A., Van Engelen, E., Heuvelink, A. & Vellema, P. (2020). Zoonotic risks of pathogens from sheep and their milk borne transmission. *Small ruminant research*, 189, 106123.
26. Deneke, T. T., Bekele, A., Moore, H. L., Mamo, T., & Al-maw, G. (2022). Milk and meat consumption patterns and the potential risk of zoonotic disease transmission among urban and peri-urban dairy farmers in Ethiopia. *BMC Public Health*, 22, 222.
27. Sevin, S., Karaca, B., Haliscelik, O., Kibar, H., & Omeroglou, E. (2021). Postbiotics secreted by *Lactobacillus sakei* EIR/CM-1 isolated from cow milk microbiota, display antibacterial and antibiofilm activity against ruminant mastitis-causing pathogens. *Italian Journal of Animal Science*, 20, 1302-1316.
28. Pierezan, M. D., Maran, B. M., Maran, E. M., & Verruck, S., (2022). Relevant safety aspects of raw milk for dairy foods processing. *Advances in Food and Nutrition Research*, 100, 211-264.
29. Garcia, S. N., Osburn, B. I. & Cullor, J. S. A. (2019). one health perspective on dairy production and dairy food safety. *One Health*, 7, 100086.
30. Megersa, R., Mathewos, M., & Fesseha, H. (2019). Isolation and Identification of *Escherichia coli* from dairy cow raw milk in Bishoftu Town, Central Ethiopia. *Arch Vet Anim Sci*, 1(1).
31. Mbago, P. (2022). Bovine mastitis on selected farms in Kamwenge District: prevalence and antibiograms of the causative bacteria. Makerere university.
32. Bertolini, A. B., Prado, A. M., Thyssen, P. J., De Souza Ribeiro Mioni, M., & De Gouvea, F.L.R. (2022). Prevalence of bovine mastitis-related pathogens, identified by mass spectrometry in flies (Insecta, Diptera) captured in the milking environment. *Letters in Applied Microbiology*, 75, 1232-1245.
33. Jadhav, P. & Das, D. (2021). Innovative Approach for Detection and Control of Mastitis in Dairy Animals. *International Journal of Agriculture Sciences*, 0975-3710.
34. Cheng, W. N., & Han, S. G. (2020). Bovine mastitis: Risk factors, therapeutic strategies, and alternative treatments-A review. *Asian-Australasian journal of animal sciences*, 33, 1699.
35. Samad, M. (2022). Review on mastitis in dairy lactating animals and their public health importance: the 56 years Bangladesh perspective. *J. Vet. Med. OH Res*, 4, 33-114.
36. Cobirka, M., Tancin, V., & Slama, P. (2020). Epidemiology and classification of mastitis. *Animals (Basel)*, 10(12), 2212.
37. Al Dujaily, A. H., & Mahmood, A. K. (2021). The effectiveness of biogenic silver nanoparticles in the treatment of caprine mastitis induced by *Staphylococcus aureus*. *Iraqi Journal of Veterinary Sciences*, 35, 73-78.
38. Arsenopoulos, K. V., Sioutas, G., Triantafyllou, E., Gelasakis, A. I., & Papadopoulos, E. (2021). Will Fly Repellency Using Deltamethrin Reduce Intramammary Infections, Stress and Fatigue Indicators of Dairy Ewes under Intensive Management?. *Pathogens*, 10(2), 232.
39. Al-Graibawi, M. A. A. & Yousif, A. A. (2021). Histopathological and Immunohistochemical Evaluation of Gangrenous Mastitis in Ewes. *Biochem. Cell. Arch*, 21, 483-490.
40. Sharma, G., Mutua, F., Deka, R. P., Shome, R., & Bandyopadhyay, S. (2020). A qualitative study on antibiotic use and animal health management in smallholder dairy farms of four regions of India. *Infection Ecology & Epidemiology*, 10, 1792033.
41. Altissimi, C., Noé-Nordberg, C., Ranucci, D. & Paulsen, P. (2023). Presence of Foodborne Bacteria in Wild Boar and Wild Boar Meat-A Literature Survey for the Period 2012-2022. *Foods*, 12, 1689.
42. Jabbar, A., Saleem, M. H., Iqbal, M. Z., Qasim, M., & Ashraf, M. (2020). Epidemiology and antibiogram of common mastitis-causing bacteria in Beetal goats. *Veterinary world*, 13, 2596.
43. Chung, L., Sahibzada, S., Annandale, H., Robertson, I., & Waichigo, F. (2021). Bacterial pathogens associated with clinical and subclinical mastitis in a Mediterranean pasture-based dairy production system of Australia. *Research in Veterinary Science*, 141, 103-109.
44. Li, X. X., Zhang, F. Q., Wang, S., Duan, X. C., & Hu, D. Y. (2023). *Streptococcus suis* prophage lysin as a new strategy

- for combating streptococci-induced mastitis and *Streptococcus suis* infection. *Journal of Antimicrobial Chemotherapy*, 78, 747-756.
45. Sharun, K., Dhama, K., Tiwari, R., Gugjoo, M. B., & Iqbal Yattoo, M. (2021). Advances in therapeutic and management approaches of bovine mastitis: a comprehensive review. *Veterinary Quarterly*, 41, 107-136.
 46. Sessitsch, A., Wakelin, S., Schloter, M., Maguin, E., & Cernava, T. (2023). Microbiome Interconnectedness throughout Environments with Major Consequences for Healthy People and a Healthy Planet. *Microbiology and Molecular Biology Reviews*, 87(3), e00212-22.
 47. Berg, G., Rybakova, D., Fischer, D., Cernava, T., & Vergès M. (2020). Microbiome Definition Re-visited: Old Concepts and New Challenges. *Microbiome*, 8(1), 103.
 48. Jalil, A., Abbasi, A. Z., Ain, Q., Hussain, Z., & Usmani, M. T. (2022). Prevalence & risk factors analysis of bovine mastitis in dairy herds of Rawalpindi district, Pakistan; a study to estimate severity & farmers' awareness about the disease.
 49. Ágredo-Campos, Á. S., Fernández-Silva, J. A., & Ramírez-Vásquez, N. F. (2023). *Staphylococcus aureus*, *Escherichia coli*, and *Klebsiella* spp. prevalence in bulk tank milk of Colombian herds and associated milking practices. *Veterinary World*, 16, 869.
 50. Romero, G., Peris, C., Fthenakis, G. C., & Diaz, J. R. (2020). Effects of machine milking on udder health in dairy ewes. *Small Ruminant Research*, 188, 106096.
 51. Girma, A., & Tamir, D. (2022). Prevalence of bovine mastitis and its associated risk factors among dairy cows in Ethiopia during 2005-2022: a systematic review and meta-analysis. *Veterinary Medicine International*.
 52. Zeleke, M. M. (2023). Investigation of palpable udder defects in non-dairy ewes: a thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Veterinary Science, Massey University, Palmerston North, New Zealand. Massey University.
 53. Pokludová, L. (2020). Prevention Is Better Than Cure. *Antimicrobials in Livestock 1: Regulation, Science, Practice: A European Perspective*, 125-165.
 54. Kubasova, T., Faldynova, M., Crhanova, M., Karasova, D., & Zeman, M. (2022). Succession, Replacement, and Modification of Chicken Litter Microbiota. *Applied and Environmental Microbiology*, 88, e01809-22.
 55. Rainard, P., Foucras, G., Fitzgerald, J. R., Watts, J., & Koop, G. (2018). Knowledge gaps and research priorities in *Staphylococcus aureus* mastitis control. *Transboundary and emerging diseases*, 65, 149-165.
 56. Piri-Gavgani, S., Ghanei, M., Fateh, A., Siadat, S. D., & Nematollahi, L. (2022). Identification of two neutralizing human single-chain variable fragment antibodies targeting *Staphylococcus aureus* alpha-hemolysin. *Iranian Journal of Basic Medical Sciences*, 25, 1207.
 57. Machado, G. P. (2018). Mastitis in small ruminants. *Anim. Husb. Dairy Vet. Sci*, 2(4), 1-9.
 58. Vermaak, P., Petzer, I. M., & Karzis, J. (2022). Effects of milking machine settings and teat liners on bovine udder health. *South African Journal of Animal Science*, 52(4), 421-432.
 59. Margastho, G., Quintas, H., Rodríguez-Estévez, V., & Simões, J. (2020). Udder morphometry and its relationship with intramammary infections and somatic cell count in Serrana goats. *Animals*, 10(9), 1534.
 60. Duheron, C. (2019). Effet du gène SOCS-2 sur la réponse inflammatoire systémique lors de mammites à *Staphylococcus aureus*, 73, 51-54.
 61. Tóth, T., Tóth, M. T., Abonyi-Tóth, Z., Silva, V., & Poeta, P., (2023). Ultrasound examination of the teat parameters of mastitis and healed udder quarters. *Veterinary and Animal Science*, 21, 100296.
 62. Notcovich, S. (2021). The physiology of the keratin plug formation in the teat canal of dairy cattle and its interaction with current and novel methods for prevention of intramammary infections: a thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Veterinary Science, Massey University, Turitea, Palmerston North, New Zealand. Massey University.
 63. Yan, Y., Zhu, K., Fan, M., Wan, W., & Zhao, X. (2022). Immunolocalization of antibacterial peptide S100A7 in mastitis goat mammary gland and lipopolysaccharide induces the expression and secretion of S100A7 in goat mammary gland epithelial cells via TLR4/NFκB signal pathway. *Animal Biotechnology*, 1-13.
 64. Rahal, A., Sohal, J., & Kumar, A. (2021). Revisiting Mastitis-An Immunopathophysiological View. *Indian Journals. Com*, 23(2), 75-91
 65. Stein, L. & Hollen, C. J. (2023). *Concept-Based Clinical Nursing Skills-E-Book: Fundamental to Advanced Competencies*, Elsevier Health Sciences.
 66. Nesaraj, J., Grinberg, A., Laven, R. & Biggs, P. (2023). Genomic epidemiology of bovine mastitis-causing *Staphylococcus aureus* in New Zealand. *Veterinary Microbiology*, 282, 109750.
 67. Addis, M. F., Pisanu, S., Monistero, V., Gazzola, A., & Penati, M. (2022). Comparative secretome analysis of *Staphylococcus aureus* strains with different within-herd intramammary infection prevalence. *Virulence*, 13, 174-190.
 68. Wen, Q., Gu, F., Sui, Z., Su, Z., & Yu, T. (2020). The process of osteoblastic infection by *Staphylococcus aureus*. *International Journal of Medical Sciences*, 17, 1327.
 69. Côté-Gravel, J., & Malouin, F. (2019). Symposium review: features of *Staphylococcus aureus* mastitis pathogenesis that guide vaccine development strategies. *Journal of dairy science*, 102(5), pp.4727-4740.62.
 70. Kahinda C. T. (2021). *Mastitis in small ruminants*. London, UK: IntechOpen
 71. Birhanu, Gemechu. (2020). *Virulence Factors and Pathogenesis of Some Streptococcus Species*. 6, 1-10.
 72. Kumari, T., Bhakat, C., & Singh, A.K. (2020). Adoption of management practices by farmers to control sub-clinical mastitis in dairy cattle. *Journal of Entomology and Zoology Studies*, 8(2), 924-927.
 73. Aqib, A. I., Muzammil, I., Naseer, M. A., Shoaib, M., & Bakht, P. (2022). Pathological insights into camel mastitis. *Acta Tropica*, 231, 106415.
 74. Abdullah, O. M., Aslam, S., Khan, M. A., Mushtaq, H., & Hassan, M. (2023). Healthy, sub-clinical, and clinical mastitis in Holstein-Friesian cattle: A comparative echotextural and electrical conductivity study. *South African Journal of Animal Science*, 53(2), 221-30.
 75. Carrillo-Lopez, L. M., Garcia-Galicia, I. A., Tirado-Gal-

- legos, J. M., Sanchez-Vega, R., & Huerta-Jimenez, M. (2021). Recent advances in the application of ultrasound in dairy products: Effect on functional, physical, chemical, microbiological and sensory properties. *Ultrasonics Sonochemistry*, 73, 105467.
76. Gougoulis, D., Athanasiou, L. V., Vasileiou, N. G., Voulgarakis, N., & Dimoveli, K. (2023). Outbreak of Acute Clinical Mastitis in Primigravidae Ewes in the Immediately Pre-Partum Period. *Ruminants*, 3(2), 133-139.
 77. Turk, R., Rošić, N., Kuleš, J., Horvatić, A., & Gelemanovic, A. (2021). Milk and serum proteomes in subclinical and clinical mastitis in Simmental cows. *Journal of Proteomics*, 244, 104277.
 78. Le Page, T., Buczinski, S., Dubuc, J., Labonté, J., & Roy, J.P. (2023). Development of a Nomogram to Estimate the 60-Day Probability of Death or Culling Due to Severe Clinical Mastitis in Dairy Cows at First Veterinary Clinical Evaluation. *Veterinary Sciences*, 10(4), 268
 79. Rötzer, V., Wenderlein, J., Wiesinger, A., Versen, F., & Rauch, E. (2023). Bovine Udder Health: From Standard Diagnostic Methods to New Approaches-A Practical Investigation of Various Udder Health Parameters in Combination with 16S rRNA Sequencing. *Microorganisms*, 11, 1311.
 80. Malcata, F. C. B. (2021). Tools for targeted treatment of bovine clinical mastitis (Doctoral dissertation, University of Glasgow).
 81. Shinde, S., Mahesh, K. & Venkanna, B. (2022). Evaluation of surf field test and california mastitis test for diagnosis of sub clinical mastitis in Crossbred Cows. *Journal of Krishi Vigyan*, 11, 37-42.
 82. Ashraf, A., & Imran, M. (2020). Causes, types, etiological agents, prevalence, diagnosis, treatment, prevention, effects on human health and future aspects of bovine mastitis. *Animal health research reviews*, 21, 36-49.
 83. Saleh, N., Allam, T., Omran, A. & Mohamed, A. (2022). A Review of The Subclinical Mastitis in Cattle with Special Reference to The New Approaches of Its Diagnosis and Control. *Journal of Current Veterinary Research*, 4, 33-46.
 84. Shewafera, A., Abraha, B., and Wasihun, P. (2022). Isolation of Selected Bacterial Pathogens from Bovine Mastitis in Selected Dairy Farms Found in Dire Dawa Town, Eastern Ethiopia (Doctoral dissertation, Haramaya University).
 85. Vasileiou, N. G., Fthenakis, G. C., & Mavrogianni, V. S. (2022). Comparison of the efficacy of intramammary or injectable antibiotic administration against staphylococcal mastitis in ewes. *Pathogens*, 11(10), 1164.
 86. Dego, O. K. (2020). Current status of antimicrobial resistance and prospect for new vaccines against major bacterial bovine mastitis pathogens. *Animal Reproduction in Veterinary Medicine*, 78921.
 87. Okoko, I. M., Maina, N., Kiboi, D. & Kagira, J. (2020). β -lactam resistance in bacteria associated with subclinical mastitis in goats in Thika Subcounty, Kenya. *Veterinary World*, 13, 1448.
 88. Zigo, F., Vasil', M., Ondrašovičová, S., Výrostková, J., & Bujok, J. (2021). Maintaining optimal mammary gland health and prevention of mastitis. *Frontiers in veterinary science*, 8, 607311.
 89. Autio, T., Tuunainen, E., Nauholz, H., Pirkkalainen, H., & London, L. (2021). Overview of control programs for non-EU-regulated cattle diseases in Finland. *Frontiers in Veterinary Science*, 8, 688936.
 90. Lianou, D. T., Michael, C. K., Gougoulis, D. A., Cripps, P. J., & Vasileiou, N. G. (2022). High milk somatic cell counts and increased *Teladorsagia* burdens overshadow non-infection-related factors as predictors of fat and protein content of bulk-tank raw milk in sheep and goat farms. *Foods*, 11(3), 443.
 91. Coffie, I. S. M. A. I. L. (2020). Health management, reproductive hormones and productive performance of dairy cattle in hot-humid and coastal environments, Ghana (Doctoral dissertation, University of Education, Winneba), 23, 37-41.
 92. Bukar, B. A. & Isa, M. M. (2023). Common Diseases of Goats, Treatment and Preventive Measures.
 93. Almashhadany, D. A., Mohammed, H. I., Muslat, T. A. M., Hassan, R. R., & Rashid, R. F. (2022). Milk-Borne Diseases. In *Health Risks of Food Additives-Recent Developments and Trends in Food Sector*. IntechOpen, 23, 71.
 94. Sachan, S., Choudhary, D., & Kausar, R. (2022). Isolation and identification of microorganisms from cow milk. *J Exp Clin microbiol*, 6, 14.
 95. Ntuli, V., Sibanda, T., Elegbeleye, J. A., Mugadza, D. T., & Seifu, E. (2023). Dairy production: Microbial safety of raw milk and processed milk products. Present knowledge in food safety. Elsevier.
 96. Poyah, N. F. (2020). Characterization of Methicillin Resistant *Staphylococcus aureus* Isolated from Raw Milk Obtained from the Eastern Cape Province. University of Johannesburg (South Africa).
 97. Goncagul, G., Gunaydin, E., & Cokal, Y. (2021). Antimicrobial susceptibility of bacteria isolated from goats with subclinical mastitis in the Southern Marmara region of Turkey. *Medycyna Weterynaryjna*, 77, 258-263.
 98. Pal, M., Kerorsa, G. B., Marami, L. M., & Kandi, V. (2020). Epidemiology, pathogenicity, animal infections, antibiotic resistance, public health significance, and economic impact of *staphylococcus aureus*: a comprehensive review. *American Journal of Public Health Research*, 8(1), 14-21.
 99. Adem, J. (2022). Review of the Zoonotic Importance of Salmonellosis and Associated Risk Factors.
 100. Ame, N. Y., Mohammed, L. A. & Ame, M. M. (2022). Review on public health importance of Salmonellosis of poultry in Ethiopia. *Int. J. Adv. Multidiscip. Res*, 9, 78-95.
 101. García-Díez, J., Saraiva, S., Moura, D., Grispoldi, L., & Cenci-Goga, B. T. (2023). The Importance of the Slaughterhouse in Surveilling Animal and Public Health: A Systematic Review. *Veterinary Sciences*, 10, 167.
 102. Raji, M. A., Kazeem, H. M., Magyigbe, K. A., & Ahmed, A. O. (2021). *Salmonella* Serovars, antibiotic resistance, and virulence factors isolated from intestinal content of slaughtered chickens and ready-to-eat chicken gizzards in the ilorin metropolis, Kwara State, Nigeria. *International Journal of Food Science*.
 103. Castañeda-Salazar, R., del Pilar Pulido-Villamarín, A., Ángel-Rodríguez, G. L., Zafra-Alba, C. A., & Oliver-Espinoza, O. J. (2021). Isolation and identification of *Salmonella* spp. in raw milk from dairy herds in Colombia. *Brazilian Journal of Veterinary Research and Animal Science*, 58, e172805-e172805.
 104. Treier, A., Stephan, R., Stevens, M. J., Cernela, N., &

- Nüesch-Inderbinnen, M. (2021). High occurrence of Shiga toxin-producing *Escherichia coli* in raw meat-based diets for companion animals-A public health issue. *Microorganisms*, 9(8), 1556.
105. Sarowska, J., Futoma-Koloch, B., Jama-Kmiecik, A., Frej-Madrzak, M., & Ksiazczyk, M., et al. (2019). Virulence factors, prevalence and potential transmission of extraintestinal pathogenic *Escherichia coli* isolated from different sources: recent reports. *Gut pathogens*, 1-6.
106. Abebe, E., Gugsa, G., & Ahmed, M. (2020). Review on major food-borne zoonotic bacterial pathogens. *Journal of tropical medicine*.
107. Paswan, R., & Park, Y. W. (2020). Survivability of *Salmonella* and *Escherichia coli* O157: H7 pathogens and food safety concerns on commercial powder milk products. *Dairy*, 1(3), 189-201.
108. Meena, P. R., Yadav, P., Hemlata, H., Tejavath, K. K., & Singh, A. P. (2021). Poultry-origin extraintestinal *Escherichia coli* strains carrying the traits associated with urinary tract infection, sepsis, meningitis and avian colibacillosis in India. *Journal of Applied Microbiology*, 130(6), 2087-2101.
109. Deddefo, A., Mamo, G., Asfaw, M., & Amenu, K. (2023). Factors affecting the microbiological quality and contamination of farm bulk milk by *Staphylococcus aureus* in dairy farms in Asella, Ethiopia. *BMC microbiology*, 23(1), 1-13.
110. Borham, M., Oreiby, A., El-Gedawy, A., Hegazy, Y., & Khalifa, H. O., et al. (2022). Review on bovine tuberculosis: An emerging disease associated with multidrug-resistant *Mycobacterium* species. *Pathogens*, 11(7), 715.
111. Mpatswenumugabo, J. P., Mukasafari, M. A., Ndahetuye, J. B., Wredle, E., & Båge, R. A. (2023). systematic literature review of milk consumption and associated bacterial zoonoses in East Africa. *Journal of Applied Microbiology*, 134(4), lxad080.
112. Ramadan, A. E. S., & Alanazi, K. H. (2023). Healthcare-Associated Infections (HAIs) Outbreak Management Manual.
113. Pavord, I., Petousi, N., & Talbot, N. (2023). Respiratory Disease. In *Medicine for Finals and Beyond*. CRC Press, pp. 155-202.
114. Mccown, M. E. (2022). Zoonotic and Infectious Vector/Tick Borne Pathogen Surveillance in Military Working Dogs, Police Working Dogs and Canines in Colombia, South America, University of Florida.
115. González-Espinoza, G., Arce-Gorvel, V., Mémet, S., & Gorvel, J. P. (2021). *Brucella*: Reservoirs and Niches in Animals and Humans. *Pathogens*, 10, 186.
116. Hensel, M.E., Garcia-Gonzalez, D.G., Chaki, S.P., Hartwig, A., & Gordy, P. W., et al. (2020). Vaccine candidate *Brucella melitensis* 16M Δ vjbR is safe in a pregnant sheep model and confers protection. *Msphere*, 5(3), 10-1128.
117. Simonetti, M. A., Suárez, J. L., & Rizzari, P. (2020). Prevalence of caprine brucellosis on herds of toba communities in Villa Río Bermejito, Chaco, Argentina (October 2010). *Transboundary and Emerging Diseases*, 67, 5-8.