

Paracetamol and Cardiac Congenital Malformations in Prediabetic Pregnancy Women

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Submitted: 07 June 2018; Accepted: 20 June 2018; Published: 10 July 2018

Abstract

Hyperglycemia can influence the development of the fetal heart, affecting both its structure and its function. Prospective and retrospective cohort studies have demonstrated an increased risk of congenital abnormalities with gestational diabetes. This observation is probably related to the inclusion of women with unrecognized type 2 diabetes [1, 2].

Substantial literature indicates that diabetes in pregnant rats and mice induces embryo lethality, growth retardation and a variable incidence of birth defects. Then, the maintenance of normal concentrations of metabolites from all nutrient classes may be important for prevention of adverse fetal outcome in diabetic pregnancy.

Acetaminophen overdose is the most often cause of acute liver injury and obese women are in particular risk, because is able to induce mitochondrial oxidative stress [3]. Acetaminophen (Paracetamol) over doses decreased embryonic low-molecular-weight thiols (glutathione and cysteine), compounds that play a vital role in the detoxication of exogenous and endogenous chemicals [3-5].

The apparent safety of Paracetamol drug, a useful analgesic only (with no anti-inflammatory properties) is compromised by its widespread and extensive chronic use, particularly in Peruvian population, where its analgesic is doing without control [6-9]. In fact, paracetamol though considered safe at a considerable low dose, especially in women could cause kidney derangement and cardiac malformations during pregnant state if the drug is ingested in the first trimester [8]. Major congenital malformations, including those affecting the cardiovascular system, remain the leading cause of mortality and morbidity in infants of diabetic mothers [10]. Thus, there is overcome potential maternal acetaminophen (paracetamol) toxicity [11].

Hyperglycemia and Cardiac Malformations

Diabetes mellitus in pregnancy is associated with an increased incidence of various congenital anomalies that occur during organogenesis. Pregnancy itself is diabetogenic caused by increased insulin resistance [12, 13]. Presentational diabetes is a major risk factor of congenital heart defects (CHDs).

Animal and human data all point to a role for the diabetic state and elevated insulin resistance in inducing congenital fetal malformations, and free radicals excess and abnormal gene activation have been compromised by week 8 of gestation: the major congenital malformations associated with non-physiological insulin resistance in pregnancy are caudal regression and renal, cardiac and central nervous system abnormalities [14].

Pregestational diabetes resulted in CHDs in 58% of the offspring, including ventricular septal defect (VSD), atrial septal defect (ASD), atrioventricular septal defects (AVSD), transposition of great arteries (TGA), double outlet right ventricle (DORV) and tetralogy of Fallot (TOF). Treatment with NAC in drinking water in presentational

diabetic mice completely eliminated the incidence of AVSD, TGA, and TOF and significantly diminished the incidence of ASD and VSD [15].

It's been demonstrated that maternal hyperglycemia caused a dilation of late-gestation fetal ventricular chambers, a reduction of total ventricular myocardial area, and an increase in transversal ascending thoracic aortic area. Glutathione, severely depleted in diabetes is probably, the main causality factor in cardiomyocyte apoptosis, in addition the hyperglycemia [15-17].

Intensive care of the pregnant mother with diabetes has dramatically decreased the incidence of diabetic fetopathy and clinical reports seem to link facial malformations to an increased incidence of sacral-caudal malformations in human diabetic pregnancy [18, 19]. Congenital malformations caused by experimental diabetes can be prevented by antioxidants in vivo that replace the glutathione depletion in mother [20, 21]. Improving the embryonic capability to scavenge oxygen radicals, either by increasing superoxide dismutase activity or by supplying a rate-limiting precursor (N-acetyl cysteine)

for the enhanced synthesis of reduced glutathione, blocks the embryonic dysmorphogenesis [22].

Available data in the literature are limited but support the idea that hyperglycemia may increase adult heart myocardial apoptosis [23]. There has been extensive evidence: maternal hyperglycemia is an inducer of birth defects that include a high incidence of cardiovascular malformations [21, 23].

Maternal diabetes mellitus may cause teratogenesis

There is a significant correlation between maternal hyperglycemia in early pregnancy and the risk of fetal abnormalities in pregnant women with diabetes mellitus, particularly type 1 and non-known-type 2 diabetes mellitus. Fetal abnormalities are strongly associated with higher levels of Hb_{A1c} in the first trimester of pregnancy [24].

Maternal diabetes mellitus is associated with increased teratogenesis, which can occur in pregestational type 1 and type 2 diabetes. Cardiac defects and with neural tube defects are the most common malformations observed in fetuses of pregestational diabetic mothers [25]. It's demonstrated a strong association between maternal obesity and risk of congenital heart defects. This association was present not only for congenital heart defects as a group, but for numerous individual defects. The overall risk increased with increasing BMI [26].

Whereas the sequelae of maternal pregestational diabetes, such as modulating insulin levels, altered fat levels, and increased reactive oxygen species, may play a role in fetal damage during diabetic pregnancy, hyperglycemia is thought to be the primary teratogen, causing particularly adverse effects on cardiovascular development. Fetal cardiac defects are associated with raised maternal glycosylated hemoglobin levels and are up to five times more likely in infants of mothers with pregestational diabetes compared with those without diabetes. The resulting anomalies are varied and include transposition of the great arteries, mitral and pulmonary atresia, double outlet of the right ventricle, tetralogy of Fallot, and fetal cardiomyopathy [25].

Despite improvements in prenatal care, the incidence of congenital malformations in diabetic pregnancies is still 3-4 times higher than in normal pregnancies. These defects could be attributed to alterations of intrauterine environment due to disorder of the maternal metabolism [27]. Glutathione (GSH), a tripeptide implicated in cellular protection against reactive oxygen species, is involved in diabetes-related embryo toxicity interestingly; teratogenicity of maternal serum in diabetic pregnancy is not mediated exclusively by increased concentrations of glucose and ketone bodies as it's demonstrated by Wentzel [28]. Both genetic background and obesity appear to influence the severity of fetal abnormalities in animals [25].

Previous studies in vivo and in vitro have suggested that the oxidative metabolism of the embryo may have a role in the teratogenicity of diabetic pregnancy. In particular, the production of reactive oxygen species by the embryonic mitochondria has been implicated in the teratological process. The induction of congenital malformations by the diabetic milieu occurs during the early embryonic development [29]. Hyperglycemia during organogenesis has a primary deleterious effect on yolk sac function with resultant embryopathy [19].

Diabetes-induced malformations have been often related, both

in vivo and in vitro studies, to morphological and physiological alterations of the yolk sac, the principal organ for the passage of nutrients from the mother to the rodent embryo [30]. The embryos explanted from diabetic mothers showed signs of developmental retardation and 16% were morphologically abnormal [31]. Therefore, reduction in embryonic GSH could reduce the protection against the oxidative stress condition described in diabetic pathology.

Norbert Freinkel suggested that the altered fuel mixture offered to the growing conceptus may be the key to most of the changes in the embryogenesis of diabetic pregnancy. He coined the term fuel-mediated teratogens. In vitro and in vivo a high glucose concentration causes embryonic dysmorphogenesis by generation of free oxygen radicals [32]. An enhanced production of such radicals in embryonic tissues is directly related to an increased risk of congenital malformations in occult diabetic pregnancy.

An abnormal handling of reactive oxygen species (ROS) is involved in diabetes-induced dysmorphogenesis in vivo. Indeed, an increased concentration of lipid peroxides, indicating damage caused by ROS, was found in fetuses of diabetes rats. In addition, embryos of diabetic rats had low concentrations of the antioxidant vitamin E compared to control embryos [33].

On the whole, in vivo and in vitro experiments indicate that hyperglycemia itself is not a major factor in producing diabetic embryopathies, but several depletion of tissues glutathione does it herself [30]. Therefore, the proved anti-teratogenic effects of supplementation of superoxide dismutase and N-acetylcysteine in rat embryo culture are supported [22].

It's proved that increased glucose levels caused embryonic maldevelopment in both normal and diabetic serum, and that despite normalization of the diabetic state, the serum from the insulin-treated diabetic rats caused more growth retardation than the no diabetic control serum [28]. Therefore, the pathogenesis of fetal malformations in diabetic pregnancy is multifactorial. Thus, maintaining metabolites from all nutrient classes at a normal level may be important in preventing adverse fetal outcome [34].

In Late Gestation

The rate of heart myocardial apoptosis may increase in adult mice under a hyperglycemic environment, and it's observed both increased apoptosis and necrosis in myocytes, endothelial cells and fibroblasts of the human adult diabetic heart [35-37]. In the fetal context, Gutierrez et al. detected ventricular chamber dilation and myocardial reduction in late gestation fetal hearts, collected from hyperglycemic pregnant CD-1 mouse dams [16, 23].

Paracetamol Danger

Teratogenic potential of diabetes may consist of two components; one associated with 'direct' teratogens perturbing developmental processes in embryos at a 'critical moment' in organogenesis, and a second component, associated with a direct or indirect influence of the diabetic environment on developmental processes in the preimplantation embryos. Thus, there is a threshold glucose level associated with a clear increase of the number of litters with severely malformed fetuses in diabetics' animals [38]. Maternal hyperglycemia altered morphology of the late-gestation fetal mouse heart [16].

Mitochondrial alterations produced by oxidative stress have been described in embryos developing in a diabetic environment. Interestingly, it's been demonstrated recently, that Paracetamol in normal doses may be able to induce mitochondrial oxidative stress [3]. Furthermore, oxygen radicals-scavenging enzymes can reduce the embryo toxic effects induced by diabetic conditions [31].

Acetaminophen overdose is the most often cause of acute liver injury. The toxic mechanism is linked to formation of an active metabolite that reacts with glutathione generating acetaminophen-glutathione conjugate (APAP-SG). This compound has been recognized to be non-toxic generally. Recent studies showed, however, that APAP-SG could possess a toxic effect too, particularly in obese women [3].

Liver glutathione stores become depleted with paracetamol overdoses –chronic use- so that the liver is unable to deactivate the toxic metabolite (NAPQ1) [39]. The paracetamol induced renal damage in pregnant women results from a mechanism similar to that which is responsible for hepatotoxicity; this mechanism is probably participate in cardiac malformations in an early pregnant state [40].

In this context, the worldwide use of paracetamol as a household analgesic, including during pregnancy may be dangerous: in fact, fetal tissues (and maternal) can be adversely affected by paracetamol and are potentially dangerous in the presence of chronic abdominal obesity (pathological insulin resistance) where liver and tissues antioxidant glutathione is reduced [6, 41].

Prevention

Animal models have been used to study the expression patterns of many genes that contribute to structural defects in the heart, although <10% of these underlie congenital cardiac defects in humans [42-44]. Intrauterine under- or over nutrition alters offspring cardiac structure and function. The expression of cardiac-specific genes is likely altered reflecting impaired cardiac insulin signaling that contributes to cardiac insulin resistance that often precedes cardiovascular disease [44].

Congenital malformations occur despite good glycemic control, thereby confirming a role for reactive oxygen species ROS and inflammatory and oxidant environment, or a lack of antioxidant protection enough, as glutathione, Vitamin E and Selenium [33, 45].

Daily oral natural vitamin E (gammatocopherol) may improve insulin action because it restores more appropriate plasma reduced glutathione, improving antioxidant cellular status and improving pancreatic b cells response to glucose in healthy and prediabetic subjects [46]. Thus, enhancing synthesis of reduced glutathione blocks the embryonic dysmorphogenesis, but not in cardiac malformations, because fetal heart was found to be hypertrophic (resorption rate in fetal organs tended to be decreased with the increased dosage of vitamin E) [20, 22]. Then, maternal administration of vitamin E can prevent congenital malformations in mid and late pregnancy, not in the early pregnancy. In comparison, Selenium may offer other additional advantages: its powerful antioxidant properties preserve reduced glutathione because its role in the major intracellular antioxidant enzyme the glutathione peroxidase. But Peruvian mothers' don't consume neither diary selenium nor vitamin D.

In addition, it is been demonstrated that acute glutathione depletion causes severe hypertension in normal rats: Since Paracetamol causes physiological reduction of glutathione, we don't surprise that this drug may increase blood pressure in advanced insulin resistant humans. Therefore, pregnant women must be caution before taking this drug, but never without medical indication. A recent investigation has found transcriptomic changes in full-genome human miRNA expression and immune modulating effects and oxidative stress responses to paracetamol even at low doses [47-50].

Most severe malformations occurs during organogenesis, period that is completed by sixth week after conception, and thus, the critical period for malformations is almost over by the time pregnancy becomes clinically apparent: in fact, organogenesis is completed before women recognize that they are pregnant; therefore, the precounseling and strict planning of pregnancy too in prediabetic women or gestational diabetes, become an urgent need.

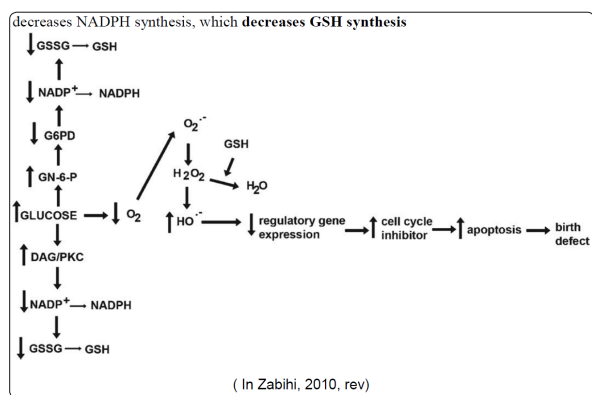
Conclusion

Because a reduction of glutathione levels in cells has been found to increase the risks for diseases and poisoning, Paracetamol is potentially dangerous in pregnant women. Pre-gestational maternal diabetes is associated with strong teratogenic effects on the kidney, urinary tract, and heart, and strongly associated with multiple congenital abnormalities [51]. Pre-conception recognition of women at high risk for type 2 diabetes and optimal glycemic control may reduce the risk of congenital anomalies. A healthy diet and regular exercise and reduction in paracetamol-toxicity in a subclinical way, may help optimize pre-pregnancy weight and reduce the risk of congenital anomalies [52, 1].

Paracetamol may increase the risk of cryptorchidism and asthma during childhood as well as preeclampsia, maternal phlebothrombosis and pulmonary embolism as it's recently reported [53]. In the presence of established diabetes, It is must be stress the importance of a strict metabolic control, started well before conception, to prevent excess rates of congenital malformation, and the intensive insulin therapy must be considered as an first option in this regard [54].

In diabetic women who is thinking to pregnant, an optimal metabolic control must been established, and the danger of paracetamol use must be informed [55-60]. Maternal hyperglycemia influence the development of the fetal heart, affecting both its structure and its function, and Paracetamol may be aggravated. Acetaminophen (Paracetamol) is one of the most common causes of poisoning worldwide, in particular in the patients with low amount of the hepatic glutathione, that is, insulin resistant and diabetic patients [60-66].

Biochemical/molecular pathway by which maternal hyperglycemia may cause birth defects Excess glucose transported to the embryo is metabolized. Increased glycolytic flux stimulates glucosamine-6-PO4 synthesis (GN-6-P), which inhibits G6PD activity, which decreases NADPH synthesis, Which decreases GSH synthesis



References

1. Armson BA, Wilson RD, Allen VM, Blight C, Gagnon A, et al. (2007) Society of Obstetricians and Gynecologists of Canada Teratogenicity associated with pre-existing and gestational diabetes. *J Obstet Gynaecol Can* 29: 927-934.
2. Passarella G, Trifirò G, Gasparetto M, Moreolo GS, Milanese O (2013) Disorders in glucidic metabolism and congenital heart diseases: detection and prevention. *Pediatr Cardiol* 34: 931-937.
3. Roušar T, Nýdlová E, Česla P, Staňková P, Kučera O, et al. (2012) Purified acetaminophen-glutathione conjugate is able to induce oxidative stress in rat liver mitochondria. *Physiol Res* 2: 103-109.
4. Mitchell JR, Jollow DJ, Potter WZ, Gillette JR, Brodie BB (1973) Acetaminophen-induced hepatic necrosis. IV. Protective role of glutathione. *J Pharmacol Exp Ther* 187: 211-217.
5. Beck MJ, McLellan C, Lightle RL, Philbert MA, Harris C (2001) Spatial glutathione and cysteine distribution and chemical modulation in the early organogenesis-stage rat conceptus in utero. *Toxicol Sci* 62: 92-102.
6. Neto JA, Oliveira-Filho RM, Simões MJ, Soares JM Jr, Kulay L Jr (2004) Long-term acetaminophen (paracetamol) treatment causes liver and kidney ultra-structural changes during rat pregnancy. *Clin Exp Obstet Gynecol* 31: 221-4.
7. Hamlyn AN, Douglas AP, James O (1978) the spectrum of paracetamol (acetaminophen) overdose: clinical and epidemiological studies. *Postgrad Med J* 54: 400-404.
8. Ucheya RE, Igweh JC (2006) Histological changes in kidney structure following a long-term administration of paracetamol (acetaminophen) in pregnant Sprague Dawley rats Niger. *J Physiol Sci* 21: 77-81.
9. Bessems JG, Vermeulen NP (2001) Paracetamol (acetaminophen)-induced toxicity: molecular and biochemical mechanisms, analogues and protective approaches. *Crit Rev Toxicol* 31: 55-138.
10. Pinter E, Haigh J, Nagy A, Madri JA (2001) Hyperglycemia-induced vasculopathy in the murine conceptus is mediated via reductions of VEGF-A expression and VEGF receptor activation. *Am J Pathol* 158: 1199-1206.
11. Horowitz RS, Dart RS, Jarvie DR, Bearer CF, Gupta U (1997) Placental Transfer of N-Acetylcysteine Following Human Maternal Acetaminophen Toxicity *J Toxicol Clin Toxicol* 35: 447-451.
12. Wu PY (1996) Infant of diabetic mother: a continuing challenge for perinatal-neonatal medicine. *Zhonghua Min Guo Xiao Er Ke Yi Xue Hui Za Zhi* 37: 312-319.
13. De Lorgeril M, Salem P, Accominotti M, Cadau M, Steghens JP, et al. (2001) Dietary and Blood Antioxidants in Patients with Chronic Heart Failure: Insights into the Potential Importance of Selenium in Heart Failure. *European Journal of Heart Failure* 3: 661-669.
14. Ryan EA (1998) Prevention and Treatment of Diabetes and Its Complications. *Medical Clinics of North America* 4: 824-842.
15. Moazzen H, Lu X, Ma NL, Velenosi TJ, Urquhart BL, et al. (2014) N-Acetylcysteine prevents congenital heart defects induced by pregestational diabetes. *Cardiovasc Diabetol* 13: 46.
16. Gutierrez JC, Hrubec TC, Prater MR, Smith BJ, Freeman LE, et al. (2007) Aortic and ventricular dilation and myocardial reduction in gestation day 17 ICR mouse fetuses of diabetic mothers *Birth Defects Res. A Clin Mol Teratol* 79: 459-464.
17. Ghosh S, Pulinkunnil T, Yuen G, Kewalramani G, An D, et al. (2005) Cardiomyocyte apoptosis induced by short-term diabetes requires mitochondrial GSH depletion. *Am J Physiol Heart Circ Physiol* 289: 768-776.
18. Eriksson UJ, Styrd J (1985) congenital malformations in diabetic pregnancy: the clinical relevance of experimental animal studies. *Acta Paediatr Scand Suppl* 320: 72-78.
19. Pinter E, Reece EA, Leranath CZ, Sanyal MK, Hobbins JC, et al. (1986) Yolk sac failure in embryopathy due to hyperglycemia: ultrastructural analysis of yolk sac differentiation associated with embryopathy in rat conceptuses under hyperglycemic conditions. *Teratology* 33: 73-84.
20. Simán CM, Eriksson UJ (1997) Vitamin E decreases the occurrence of malformations in the offspring of diabetic rats *Diabetes* 46: 1054-1061.
21. Reece EA, Wu YK (1997) Prevention of diabetic embryopathy in offspring of diabetic rats with use of a cocktail of deficient substrates and an antioxidant. *Am J Obstet Gynecol* 176: 790-798.
22. Wentzel P, Thunberg L, Eriksson UJ (1997) Teratogenic effect of diabetic serum is prevented by supplementation of superoxide dismutase and N-acetylcysteine in rat embryo culture. *Diabetologia* 40: 7-14.
23. Gutierrez JC, Prater MR, Smith BJ, Freeman LE, Mallela MK, et al. (2009) Late-gestation ventricular myocardial reduction in fetuses of hyperglycemic CD1 mice is associated with increased apoptosis. *Birth Defects Res B Dev Reprod Toxicol* 86: 409-415.
24. Todorova K, Mazneikova V, Ivanov S, Genova M (2005) the frequency of mild and severe fetal malformations in diabetic women with high values of glycosilated hemoglobin in early pregnancy. *Akush Ginekol (Sofia)* 44: 3-10.
25. Corrigan N, Brazil DP, McAuliffe F (2009) fetal cardiac effects of maternal hyperglycemia during pregnancy *Birth Defects Res. A Clin Mol Teratol* 85: 523-530.
26. Mills JL, Troendle J, Conley MR, Carter T, Druschel CM (2010) maternal obesity and congenital heart defects: a population-based study. *Am J Clin Nutr* 91: 1543-1549.
27. Giavini E, Broccia ML, Prati M, Domenico Roversi G (1991) Diet composition modifies embryo toxic effects induced by experimental diabetes in rats. *Biol Neonate* 59: 278-286.
28. Wentzel P, Eriksson UJ (1996) Insulin treatment fails to abolish the teratogenic potential of serum from diabetic rats. *Eur J Endocrinol* 134: 459-466.
29. Yang X, Borg LA, Eriksson UJ (1995) Altered mitochondrial morphology of rat embryos in diabetic pregnancy. *Anat Rec*

- 241: 255-267.
30. Menegola E, Broccia ML, Prati M, Ricolfi R, Giavini E (1996) Glutathione status in diabetes-induced embryopathies. *Biol Neonate* 69: 293-297.
 31. Lee AT, Reis D, Eriksson UJ (1999) Hyperglycemia-induced embryonic dysmorphogenesis correlates with genomic DNA mutation frequency in vitro and in vivo. *Diabetes* 48:371-376.
 32. Simán M (1997) congenital malformations in experimental diabetic pregnancy: aetiology and ant oxidative treatment. Minireview based on a doctoral thesis *Ups. J Med Sci* 102: 61-98.
 33. Giavini E (1993) Diabetes in pregnancy: experimental aspects. *Ann IST Super Sanita* 29: 27-34.
 34. Styrud J, Thunberg L, Nybacka O, Eriksson UJ (1995) Correlations between maternal metabolism and deranged development in the offspring of normal and diabetic rats. *Pediatr Res* 37: 343-353.
 35. Frustaci A, Kajstura J, Chimenti C, Jakoniuk I, Leri A, et al. (2000) Myocardial cell death in human diabetes. *Circ Res* 87: 1123-1132.
 36. Fiordaliso F, Leri A, Cesselli D, Limana F, Safai B, et al. (2001) Hyperglycemia activates p53 and p53-regulated genes leading to myocyte cell death. *Diabetes* 50: 2363-2375.
 37. Cai L, Li W, Wang G, Guo L, Jiang Y, et al. (2002) Hyperglycemia-induced apoptosis in mouse myocardium: mitochondrial cytochrome C-mediated caspase-3 activation pathway *Diabetes* 51: 1938-1948.
 38. Torchinsky A, Toder V, Carp H, Orenstein H, Fein A (1997) In vivo evidence for the existence of a threshold for hyperglycemia-induced major fetal malformations: relevance to the etiology of diabetic teratogenesis. *Early Pregnancy* 3: 27-33.
 39. Kaneo Y, Ogawa K, Tanaka T, Fujihara Y, Iguchi S (1994) A protective effect of glutathione-dextran macromolecular conjugates on acetaminophen-induced hepatotoxicity dependent on molecular size *Biol Pharm Bull* 17: 1379-1384.
 40. Sule AA, Tai DY, Tze CC, Deepa B, Leow MK (2006) potentially fatal paracetamol overdose and successful treatment with 3 days of intravenous N-acetylcysteine regime--a case report. *Ann Acad Med Singapore* 35: 108-111.
 41. Reed DJ, Fariss MW (1984) Glutathione Depletion and Oxidant Susceptibility. *Pharmacol Revi* 2: 255-335.
 42. Andersen TA, Troelsen Kde L, Larsen LA (2014) of mice and men: molecular genetics of congenital heart disease. *Cell Mol Life Sci* 71: 1327-1352.
 43. Richards AA, Garg V (2010) Genetics of congenital heart disease. *Curr Cardiol Rev* 6: 91-97.
 44. Govindsamy A, Naidoo S, Cerf ME (2018) Cardiac Development and Transcription Factors: Insulin Signalling, Insulin Resistance, and Intrauterine Nutritional Programming of Cardiovascular Disease. *J Nutr Metab* 2018: 8547976.
 45. Mill JL, Knopp RH, Simpson JL, Jovanovic-Peterson L, Metzger BE, et al. (1988) Lack of relation of increased malformation rates in infants of diabetic mothers to glycemic control during organogenesis. *N Engl J Med* 318: 671-676.
 46. Paolisso G, D'Amore A, Giugliano D, Ceriello A, Varricchio M, et al. (1993) Pharmacologic doses of vitamin E improve insulin action in healthy subjects and non-insulin-dependent diabetic patients. *Am J Clin Nutr* 57: 650-656.
 47. Vaziri ND, Wang XQ, Oveisi F, Rad B (2000) Induction of oxidative stress by glutathione depletion causes severe hypertension in normal rats *Hypertension* 36: 142-146.
 48. Sudano I, Flammer AJ, Periat D, Enseleit F, Hermmann M, et al. (2010) Acetaminophen Increases Blood Pressure in Ambulatory Patients With Coronary Artery Disease. *Circulation* 122: 1789-1796.
 49. Da Silva MH, da Rosa EJ, de Carvalho NR, Dobrachinski F, da Rocha JB, et al. (2012) acute brain damage induced by acetaminophen in mice: effect of diphenyl diselenide on oxidative stress and mitochondrial dysfunction. *Neurotox Res* 21: 334-344.
 50. Jetten MJ, Gaj S, Ruiz-Aracama A, de Kok TM, van Delft JH, et al. (2012) Omics analysis of low dose acetaminophen intake demonstrates novel response pathways in humans. *Toxicol Appl Pharmacol* 259: 320-328.
 51. Honda Y, Kessoku T, Sumida Y, Kobayashi T, Kato T, et al. (2017) Efficacy of glutathione for the treatment of nonalcoholic fatty liver disease: an open-label, single-arm, multicenter, pilot study. *BMC Gastroenterol* 17: 96.
 52. Nielsen GL, Nørgard B, Puho E, Rothman KJ, Sørensen HT, et al. (2005) Risk of specific congenital abnormalities in offspring of women with diabetes. *Diabet Med* 22: 693-696.
 53. Burdan F, Starosławska E, Szumilo J (2012) prenatal tolerability of acetaminophen and other over-the-counter non-selective cyclooxygenase inhibitors. *Pharmacol Rep* 64: 521-527.
 54. Fuhrmann K, Reiher H, Semmler K, Glöckner E (1984) the effect of intensified conventional insulin therapy before and during pregnancy on the malformation rate in offspring of diabetic mothers. *Exp Clin Endocrinol* 83: 173-177.
 55. Baker L, Egler JM, Klein SH, Goldman AS (1981) meticulous control of diabetes during organogenesis prevents congenital lumbosacral defects in rats. *Diabetes* 30: 955-959.
 56. Bauer AZ, Kriebel D (2013) Prenatal and perinatal analgesic exposure and autism: an ecological link. *Environ Health* 12: 41.
 57. Botella-Llusiá J, Pereira A, DeIsla JL (1970) Latent diabetes complicating pregnancy as a cause of fetal wastage and infertility. *Acta Eur Fert* 2: 313-334.
 58. Carvalho NR, da Rosa EF, da Silva MH, Tassi CC, Dalla Corte CL, et al. (2013) new therapeutic approach: diphenyl diselenide reduces mitochondrial dysfunction in acetaminophen-induced acute liver failure. *PLOS One* 8: e81961.
 59. Eriksson U, Dahlström E, Larsson KS, Hellerström C (1982) Increased incidence of congenital malformations in the offspring of diabetic rats and their prevention by maternal insulin therapy. *Diabetes* 31: 1-6.
 60. Farquhar H, Stewart A, Mitchell E, Crane J, Eyers S, et al. (2010) The role of paracetamol in the pathogenesis of asthma. *Clin Exp Allergy* 40: 32-41.
 61. Fine EL, Horal M, Chang TI, Fortin G, Loeken MR (1999) Evidence that elevated glucose causes altered gene expression, apoptosis, and neural tube defects in a mouse model of diabetic pregnancy. *Diabetes* 48: 2454-2462.
 62. Larson AM, Polson J, Fontana RJ, Davern TJ, Lalani E, et al. (2005) Acetaminophen-induced acute liver failure: results of a United States multicenter, prospective study. *Hepatology* 42: 1364-1372.
 63. Lisowski LA, Verheijen PM, Copel JA, Kleinman CS, Wassink S, et al. (2010) Congenital heart disease in pregnancies complicated by maternal diabetes mellitus. An international clinical collaboration, literature review, and meta-analysis. *Herz* 35: 19-26.
 64. Whitcomb DC, Block GD (1994) Association of acetaminophen hepatotoxicity with fasting and ethanol use. *JAMA* 272: 1845-

1850.

65. Zabihi S1, Loeken MR (2010) Understanding diabetic teratogenesis: where are we now and where are we going? Birth Defects Res A Clin Mol Teratol 88: 779-790.

66. Zhou X, Lu X (2013) the role of oxidative stress in high glucose-induced apoptosis in neonatal rat cardiomyocytes. Exp Biol Med (Maywood) 238: 898-902.

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