Isolation and Identification of Gram Positive and Negative Bacteria from Dairy Department Waste Lines at Baraton University

Stella Wanyama

Abstract

Baraton University dairy farm is an environment that attracts a microbiologist to inquire the composition of bacteria that exist there in. The knowledge of bacteria has in repeatedly amazed the life scientist community that have invested to acquire more information in this microbiology world. The study engages fundamental tests such as gram stain, endospore stain, and assays for specific microbial activities & enzymes, susceptibility on disinfectant and antibiotic, utilization of specific substrate and culture characteristics. The two organisms (gram negative and positive) tested positive for sucrose & lactose fermentation, Indole & Methly red, catalase & Oxidase, were both facultative and motile. On contrary, gram positive bacteria had spores and had a gamma haemolysis on Blood Agar, while gram-negative bacteria haemolysed beta haemolysis. To draw a conclusion on the identity of the two organisms is that, the gram positive is a Bacillus, while gram negative is Escherichia coli.

Keywords: Microbial Classification, Grams Stain, Endospore Stain, Susceptibility, Substrate.

Introduction

Barton University dairy farm is a source of delicious and nutritious milk, one of its kinds. The milk is consumed as yoghurt, mala, ice cream and fresh milk by the students, residential faculty members and to the rest of the Barton Community. As a system, the farm intentionally inoculates bacteria for production of yoghurt and mala. While non-intention & by nature providence, some bacteria grow and carry out the metabolic activities.

The dairy department is composed of three major facilities: the processing, milking and cattle feeding & resting structures. Mainly the cattle feed on hay made of maize stalk from the farm. The drainage from these facilities meets at some point. The three units are sources of enteric bacteria, mycobacterium and other parasites that are associated with cattle breeding [3].

The study primarily is to establish a pure culture of gram positive and negative bacteria and use special staining, assays for specific activities & microbial enzymes, effects of disinfectants and antibiotic micro-organism, selective and differential media and culture characteristics such as oxygen requirement for organism growth, organism motility & haemolysis pattern to identify the taxonomical name of the bacteria found at the dairy department waste system.

Materials & Method

The aseptic technique was used in dipping a sterile swab into the specimen from the waste drainage and inoculated into TSB culture media. Three inoculated culture media were established; culture A and B were incubated at 370°C while culture C was kept at room temperatures to determine the growth of the organism at ambient temperature for 48 hours.

Three plates of TSA culture were obtained to establish colonies and a gram stain was done to acquire a gram positive and negative bacteria by heat fixing, application of crystal violet and using iodine solution as mordant, thereafter a decolourization was carried out and finally counter stained with safranin. The cell shape and arrangement were determined through the use of a microscope. Parts of the stained colonies were inoculated by use of quadrant streaking to create pure cultures of gram positive and negative of unknown microorganisms [1].

An acid fast stain was applied to both gram positive and negative bacteria by use of Kinyoun’s carbolfuchsins, decolorized with Acid – alcohol solution and counter stained with Leoffler’s methylene blue and exposed to steam & heat control. The applications of Acid fast stain to confirm the presence of mycobacterium spp. identified in the cause of Tuberculosis in both cattle and human [2, 5]. To ascertain the formation of endospore in bacteria vegetative cells such as Bacillus anthracis, schaeffer
Table 1: Show Casing Tests Results and Characteristics of Gram Positive and Negative Bacteria

<table>
<thead>
<tr>
<th>Tests and Morphological&amp;Culture characteristics</th>
<th>Gram Positive Bacteria</th>
<th>Gram Negative Bacteria</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acid Fast</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Endospore</td>
<td>positive</td>
<td>Negative</td>
</tr>
<tr>
<td>Lactose broth</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>Maconkey agar</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>Triple –sugar Iron</td>
<td>Acid and gas production (Lactose and sucrose fermentation)</td>
<td>Acid and gas production (Lactose and sucrose fermentation)</td>
</tr>
<tr>
<td>Mannitol salt agar</td>
<td>Negative</td>
<td>Negative</td>
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<tr>
<td>Gelatinase</td>
<td>Negative</td>
<td>Negative</td>
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<tr>
<td>Amylase</td>
<td>Negative</td>
<td>Negative</td>
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<td>urease</td>
<td>Negative</td>
<td>Negative</td>
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<tr>
<td>Catalase</td>
<td>Positive</td>
<td>Positive</td>
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<tr>
<td>Oxidase</td>
<td>Positive</td>
<td>Positive</td>
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<tr>
<td>Phenylalanine Deaminase</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Indole (Tryptophanase)</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>Methly red</td>
<td>Positive</td>
<td>Positive</td>
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<tr>
<td>Voges proskauer Test</td>
<td>Negative</td>
<td>Negative</td>
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<tr>
<td>Citrate Utilization</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Cell shape and arrangement</td>
<td>Large single rods</td>
<td>Short single rods</td>
</tr>
<tr>
<td>Oxygen requirement</td>
<td>Facultative</td>
<td>Facultative</td>
</tr>
<tr>
<td>Hemolysis pattern</td>
<td>Gamma</td>
<td>Beta</td>
</tr>
<tr>
<td>Motility</td>
<td>Show movement</td>
<td>Show movement</td>
</tr>
</tbody>
</table>

Special features such as oxygen requirement for organisms’ growth, their motility and haemolysis pattern were tested through growth analysis on thioglycollate broth, motility media and Blood Agar Plates.

The susceptibility to disinfectant of the two cultures was determined by exposure to 10% omo, 10% jik, 10% dettol and 70% ethyl alcohol. The Kirby bauer disk method was also carried to determine the susceptibility of ampicillin, lincomycin, penicillin, minocycline, erythromycin, chloramphenicol and co-trimoxazole.

Results

The conjunction point of the drainage constituted of green and turbid sewage that produced a strong odour. Apart from the cattle waste, a combat liquid soap was evident in the sewage. It was an attractive site to investigate the bacteria growth. Inoculation of the specimen in TSB produced a saturated TBS with bacteria growth. Three TSA plate displayed different colonies in size, colour and shape. Gram-positive presenting large single blue rods, while gram negative produced short single rods. The selections were picked out of the numerous colonies on TSA plates.
gram negative exhibited no inhibition zones with ethyl alcohol, L,P, and M. While demonstrated inhibitions zones with omo (1.7cm), jik (1cm), dettol (1.1cm), A(1cm),Mi(2.5), E(1.6cm), CO(1cm), and C(2.6cm).

The organisms are motile, facultative and illustrated gamma haemolysis for gram positive and beta haemolysis for gram negative.

**Discussion and Conclusion**

The organisms’ habitat detects presence of gram positive and negative enterobacteriaeae group of bacteria, whereby most of these bacteria carry out mixed acid fermentation and grow at low PH detected in Methyl red tests. VP test is used widely to classify enterobacteriaeae strains, those bacteria that produce acetoin are VP test positive. In this case, acetoin increase the PH by reducing the acid in the environment or medium [8, 21]. Some cultures, with prolonged time of growth (to seven days) have tested VP positive. As a result, acidity is reduced in the media [7]. Barry and Feeney declares that acetoin can be detected a little faster if creative was added to MR VP broth followed by Barrett reagents [9].

Endospore formation on gram-positive cells has an ability to create resistance to some antibiotics and detergents as described in the results, there were no inhibition zones. The spores and endospore were quite evident in both Endospore and Acid Fast Staining. Acid Fast Staining of gram-positive bacteria demonstrated refractive bodies of endospore and free spores [6].

Ampicillin is used as plasmid maintaining agent during *Escherichia coli* MG 1655 cultivation [7]. Ampicillin does not necessitate limitation of growth but plasmid transfer as demonstrated by lack of inhibition zones in gram positive and negative. Lactic acid bacteria can be used as antibiotic against *E. coli* and *Klebsiella spp.* *Escherichia coli* is the most resistant bacteria and therefore lactic acid bacteria antibiotic activities is a solution to antibiotic resistant that is transmitted to Human population [10]. The susceptibility to antibiotic greatly depends on the environment, therefore to determine the resistance of bacteria on host, requires an environment resembling the host [13]. Pre exposure to some antibiotics or detergents can create resistance of bacteria towards other antibiotics or detergents [15].

*Escherichia coli* are used as a control for detection of tryptophanase. According to Newton and Shell tryptophanase in *Escherichia coli* catalyses this reaction L-Tryptophan + H20 - indole + pyruvate + NH3 [10, 14].

In normal circumstances and anatomical structure of *Escherichia coli* do not support acetoin production and citrate utilization. Therefore, VP and citrate tests are supposable negative. The transfers of plasmid in Isolates of different species and environment have proved utilization of citrate. Generation of acetoin by *Escherichia coli* strains in vegetables and fruits due to acquisition of *budAB* gene alter the normal functionality of *Escherichia coli*. These two phenomena leads to misdiagnosis of *E. coli* [7, 17].

Presence of catalase reinforces the facultative ability demonstrated the thioglycollate broth as most bacteria from the family of Enterobacteriaceae that are either aerobic or facultative produce catalase enzymes [11]. *Escherichia coli* exhibit a number of cytochrome oxidase such as cytochrome c oxidase; cytochrome bed oxidase and cytochrome boo oxidase that aid it to survive in different habitat. Cytochrome bed oxidase has proved to be pathogenetic enhancer when exposed to NO and enhances respiration in hydrogen sulphide environment [12, 16]. Tryptophan utilization by tryptophans generates sulphide in *Escherichia coli* that triggers cytochrome bed oxidase mechanism [16].

In the experiment, the two bacteria culture indicated movement, their growth extended away from the inoculation line. *E. coli* have capabilities of swimming and swarming through the environment by use of flagella and as a colony in search of energy and detecting of some substances [19, 20].

The results presented two types of haemolysis; Gamma haemolysis for gram-positive bacteria, which indicates the absence of red blood, cells lysing due to lack of toxin production. Beta haemolysis in gram-negative bacteria is due to toxin production. *E. coli* strains such as *Escherichia coli* O157 enter pathogenic *Escherichia coli* and Shiga Toxin-Producing *E. coli* are sources of toxin that lyses red blood cells [22-24].

**Author’s Contribution**

Stella carried out the experiments and authored this article.

**Conflict of Interest**

There is no conflict of interest.

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**References**


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