

## Ameliorating Effect of Zinc Oxide Nanoparticles against Hematotoxicity Induced by Cyclophosphamide in Male Albino Rats

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### Abstract

**Background:** Cyclophosphamide (CP) is a drug with a wide spectrum of clinical uses. Zinc oxide is the most widely used nanoparticles. Nanoparticles could induce oxidative stress that eventually leads to cell toxicity, inflammation and hemolysis.

**Objectives:** The objective of this study was to evaluate the hematological changes induced by Zn-O nano-particles and/or Cyclophosphamide in male rats.

**Materials and Methods:** Twenty four adult male rats (Sprague Dawley) were grouped randomly into four groups of six rats each. Group I. Control group: Received 0.2 ml saline /day i.p. injection for 14 days (day by day), group II (CP group): Received CP 20 mg/kg/day body weight (b.w.) day by day for 14 days by intraperitoneal injection, Group III (nZnO group): Received nZnO (5 mg/kg)/day b.w., intraperitoneally for 14 days. Group IV (CP + ZnO NPs group): Received nZnO group: Received nZnO (5 mg/kg/day) b.w., intraperitoneally for 14 days, plus CP 20 mg/kg/day body weight (b.w.) day by day for 14 days by intraperitoneal injection. At the end of the experimental period, rats were anesthetized using light ether. Blood samples were taken for hematological evaluation.

**Results:** Red blood cells count, hemoglobin concentration, and white blood cells count were significantly decline in rats treated with CP in comparison to control group, while combination of nZnO with CP reduced changes in red blood cells and hemoglobin values. Neutrophils, lymphocytes, eosinophils, and monocytes count were significantly decreased in CP-immunosuppressed group when compared with the control group. In CP-immunosuppressed animals treated with nZnO, these parameters were improved when compared with CP treated groups.

**Conclusion:** It can be concluded that CP induced changes in the hematological parameters. Treatment of rats with zinc oxide nano-particles and CP together ameliorated the toxicity induced by CP. These results may provide further visions into proper treatment of patients by improving side effects of chemotherapy. However further studies are necessary to establish optimal doses of nZnO and receive the best safety profile.

**Keywords:** Zinc Oxide Nanoparticles, Cyclophosphamide, Hematotoxicity, Hematological changes, RBCs, WBCs, White blood cells differential count

### Introduction

Cyclophosphamide (CP) is a drug with a wide spectrum of clinical uses. Its effectiveness in the treatment of cancer (lymphoma, acute and chronic leukemias, multiple myeloma) and non-malignant diseases such as rheumatoid arthritis and vasculitis has been well established [1]. However, this drug also induces acute inflammation of the urinary bladder, renal damage and liver damage [2-5]. CP use is limited due to myelotoxicity, and its marked organ toxicity associated with increased oxidative stress and inflammation [6, 7]. CP is a prodrug with its two active metabolites, phosphoramidate mustard (PM) and acrolein. PM is an anticancer metabolite whereas acrolein is a toxic metabolite, responsible for hematological and myelotoxicity [6, 8].

Biosafety issues of manufactured nanomaterials have recently attracted significant public attention and research interest. Zinc oxide (ZnO) nanoparticles (NPs) are mass-produced nanomaterials and have been widely used in the industrial production of sunscreen, antibacterial reagents, rubber additives and pigments [9]. ZnO NPs have also found applications in the production of solar cells, photocatalysts, transparent conductive films and ultraviolet photodetectors [10-13]. Previous studies have demonstrated that ZnO NPs are toxic to microorganisms, cells, plants, aquatic biota and rodents [14-19]. The red blood cells (RBCs) are susceptible to oxidative stress damage. Nanoparticles could induce oxidative stress that eventually leads to cell toxicity and hemolysis of RBCs at certain doses [20, 21]. Smaller nanosized particles caused granulocyte activation and hemolysis and induced inflammation and hemolysis in human blood samples [21-23]. Zinc oxide is the most widely used nanoparticles among various nanomaterials. Recently, these nanoparticles have been shown to specifically kill cancerous cells;

therefore, it is believed that these nanoparticles may be used as an alternative antitumor agent. However, it is also known that these nanoparticles pose several deleterious effects to living beings [24]. Many previous studies have suggested that the high solubility of ZnO NPs (i.e. the ability to release a large number of free zinc ions) might play an important role in the cytotoxicity. ZnO NPs are one of the most abundantly used nanomaterials in consumer products and biomedical applications. As a result, human exposure to these NPs is highly frequent and they have become an issue of concern to public health. Although toxicity of ZnO NPs has been extensively studied and they have been shown to affect many different cell types and animal systems, there is a significant lack of toxicological data for ZnO NPs.

### Objectives

The objective of this study was to evaluate the hematological changes induced by Zn-O nano-particles and/or Cyclophosphamide in male rats.

### Materials and Methods

Experiment was carried out at Department of Environmental Studies, Institute of Graduate Studies and Research, Alexandria University laboratories

### Chemicals

All reagents were of the highest quality. Zinc oxide as nanoparticles with an average size 67 nm is a gift from Dr. Eman El-Trass. Cyclophosphamide (200 mg/ampoule) was purchased from Sigma-Aldrich.

### Animals

24 male rats with average body weight of 150±30 g were obtained from faculty of agriculture, Alexandria University, and acclimatized for two weeks before starting the experiment. The rats were maintained under temperature (25°C). They were fed with standard food and had free access to water. Animals were randomly divided into 4 groups and housed in Universal galvanized wire cages.

### Experimental design

Adult male rats (Sprague Dawley) were randomly divided into four groups (six rats each) as follows:

**Group I.** Control group: Received 0.2 ml/day saline i.p. injection for 14 days (day by day).

**Group II.** CP group: Received CP 20 mg/kg/day body weight (b.w.) (i.p. injection of saline) day by day for 14 days by intraperitoneal injection [25].

**Group III.** nZnO group: Received nZnO (5 mg/kg/day) b.w., intraperitoneally for 14 days [26].

**Group IV.** CP + nZnO group: Received nZnO (5 mg/kg/day) b.w., intraperitoneally for 14 days, plus CP 20 mg/kg/day b.w., day by day for 14 days by intraperitoneal injection.

After 24-hr from last treatments all the animals were anesthetized using light ether. Blood samples were taken from the heart within 1

min. Tubes was used to compile blood drawn from the heart directly; 1 ml were collected on sodium heparin for hematological studies.

### Hematological Measurements

One milliliters of blood collected into heparinized sample bottles were analyzed for hematological parameters; was used for complete blood picture analysis by Coulter (Hemat 8 analyser; SEAC).

### Statistical Analysis

The values are expressed as mean±SE. The results were computed statistically using statistical Package for Social Sciences (SPSS software package, version 25) using one-way analysis of variance (ANOVA). Post hoc testing was performed for inter-group comparison on using the LSD. P<0.05 was considered as significant [27].

### Results and Discussion

#### Effects of zinc oxide nanoparticles, cyclophosphamide and their combination on blood RBCs count, hemoglobin concentration, WBCs count

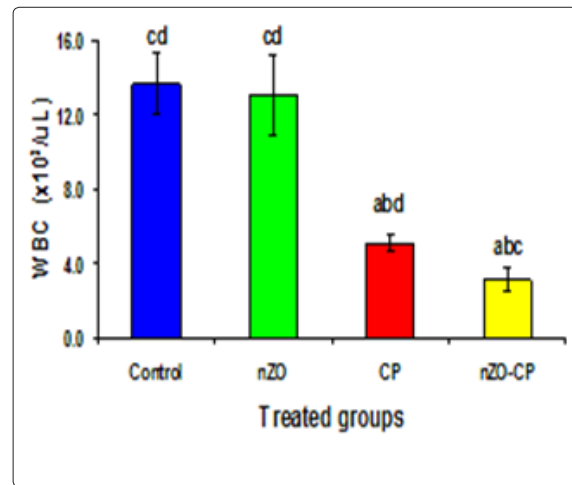
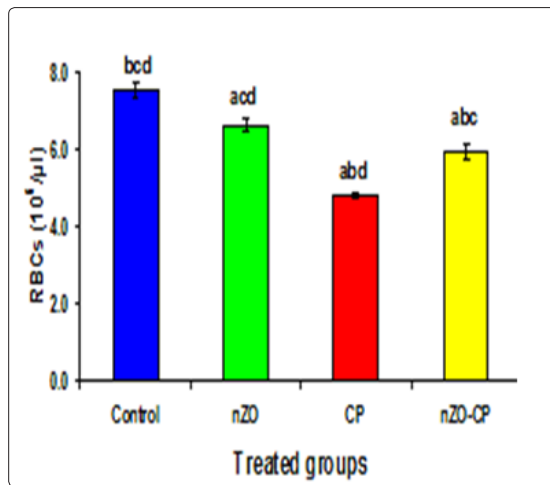
In the present experimental study, it has been observed that red blood cell counts, hemoglobin and white blood cells count were significantly decline in rats treated with CP in comparison to control group (Tables 1, 2 and Figures 1-3). Our results may suggest that CP suppresses bone marrow ability to produce new ones, resulting in lowering of blood cells counts which results in decrease of Hb percentage in the blood. In connection to this, cyclophosphamide altered liver and kidney functions by modulating liver enzymes [28, 29]. However, the activity of this enzyme is not limited only to the liver as it is also present in the brain, muscle and red blood cells [30]. The cyclophosphamide-induced changes in the level of blood cells in this study were similar to those observed in humans during treatment with this drug. The most common abnormalities found in humans are leukopenia and anemia. Similar results were obtained in rats after a single intraperitoneal dose of CP (50 mg/kg): Significant decreases in WBC count and Hb concentration were noted [31]. Chemotherapy-induced anemia is one of the most common side effects experienced by cancer patients, occurring in approximately 70-90% of those undergoing treatment for the disease [32].

The response to stress is characterized by hormone change (catecholamine and corticosteroid) that induced alteration on the haematological parameter. Increase on the volume of the plasma, loss of water in the RBCs and haemolysis of RBCs in the blood stream. It has already been proved that the organs where the Zn levels are of prime importance, the deficiency caused disturbance in the redox balance and oxidative stress leading to the cellular damages [33]. Initial experiment on Zn provides the evidence that at appropriate concentration Zn can protect CP-induced damage in the urinary bladder and blood of rat [34]. It has also been suggested that further experiments can explore the underlying mechanisms of Zn protection against CP-induced toxicity. Pharmacological modulation of metallothionein was used as one of the strategies to overcome the toxic issues of several drugs, especially with anticancer agents [35, 36]. Previous studies have reported that pharmacological increase in metallothionein levels during anticancer drug regime protects the normal cells from the toxic insults of the drug [35].

**Table 1: Effects of treatment of rats with zinc oxide nanoparticles or cyclophosphamide, and their mixture on RBCs count and hemoglobin concentration**

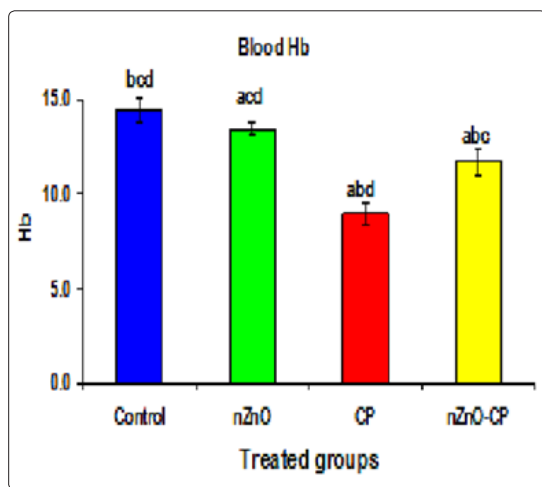
Groups	Groups			
	Control	nZnO	CP	nZnO + CP
Parameters	Mean±SE	Mean± SE	Mean± SE	Mean± SE
RBCs Count (x10 <sup>6</sup> /μl)	7.57 ± 0.22 <sup>bcd</sup>	6.62 ± 0.18 <sup>acd</sup>	4.82 ± 0.10 <sup>abd</sup>	5.96 ± 0.22 <sup>abc</sup>
Hb (g/dl)	14.42 ± 0.66 <sup>bcd</sup>	13.44 ± 0.33 <sup>acd</sup>	8.94 ± 0.63 <sup>abd</sup>	11.68 ± 0.716 <sup>abc</sup>

Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup> Comparison of nZnO + CP and other groups.



**Figure 1:** Effect of treatment of rat with zinc oxide nanoparticles (nZnO) and/or cyclophosphamide (CP) on RBCs count (x10<sup>6</sup>/μL). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.

**Figure 3:** Effect of treatment of rat with zinc oxide nanoparticles (nZnO) and/or cyclophosphamide (CP) on RBCs count (x10<sup>6</sup>/μL). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.



**Figure 2:** Effect of treatment of rat with zinc oxide nanoparticles (ZnONPs) and/or cyclophosphamide (CP) on Haemoglobin Concentration (Hb, g/dl). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.

### Effects of Zinc Oxide Nanoparticles, Cyclophosphamide and Their Combination on Blood Neutrophils, Lymphocytes, Eosinophils, and Monocytes Count

History has shown that CP is one of the most potent immunosuppressive drugs. In the present study, we found that CP induced leukopenia, neutropenia and lymphocytopenia in rats, these results parallel to the result of winkelstein who demonstrated that CP is an effective inhibitor of cell mediated immune response and leads to a depletion of lymphocytes in the peripheral blood and tissue [37].

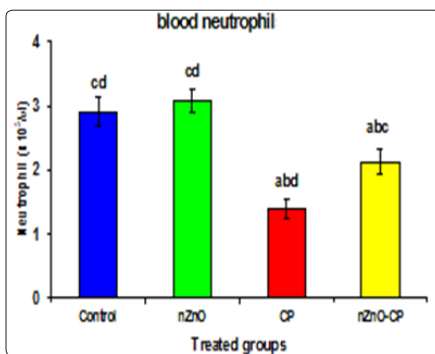
In the present work, regarding to the result of neutrophils, lymphocytes, eosinophils and monocyte were significantly decreased in CP - immunosuppressed group when compared with the control group. In CP- immunosuppressed animals treated with nZnO, these parameters were improved when compared with CP treated groups (Tables 2 and Figures 4-7).

Chemotherapy-induced anemia is one of the most common side effects experienced by cancer patients, occurring in approximately 70-90 % of those undergoing treatment for the disease [32]. This is because chemotherapeutic drugs kill rapidly dividing cells in the body, including cancer cells and normal cells which include red blood cells, at the same time suppress bone marrow ability to produce new ones, resulting in decrease of blood Hb level.

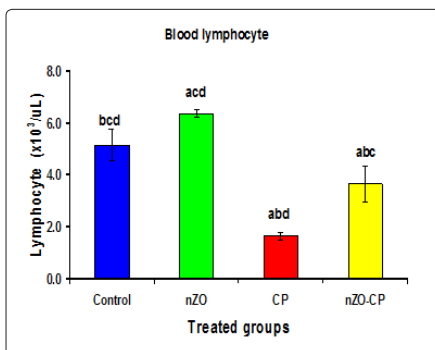
The greater efficacy shown by the pretreatment group might be the result of the compound providing some added protection to the target cells before exposure to the chemotherapeutic agent. From this observation, it is possible to conclude that ZnO NPs its chemoprotective effect against CP-induced cellular toxicity in part by scavenging the free radicals generated by CP reactive metabolites. Because CP does not act through oxidative stress mechanism, it is expected that the Zn would be beneficial to the host but without reducing the efficiency of the treatment.

**Table 2: Effects of treatment of rats with zinc oxide nanoparticles or cyclophosphamide, and their mixture on total WBCs, neutrophils, lymphocytes, eosinophils, and monocytes count**

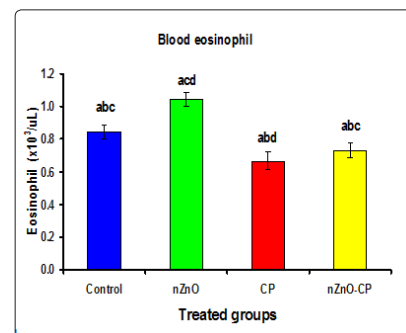
Parameters	Groups			
	Control	nZnO	CP	nZnO + CP
WBCs Count ( $\times 10^3/\mu\text{L}$ )	13.64 $\pm$ 1.65 <sup>cd</sup>	13.04 $\pm$ 2.20 <sup>cd</sup>	5.10 $\pm$ 0.50 <sup>abd</sup>	3.10 $\pm$ 0.66 <sup>abc</sup>
Neutrophils Count ( $\times 10^3/\mu\text{L}$ )	2.90 $\pm$ 0.22 <sup>cd</sup>	3.08 $\pm$ 0.18 <sup>cd</sup>	1.40 $\pm$ 0.15 <sup>abd</sup>	2.13 $\pm$ 0.19 <sup>abc</sup>
Lymphocytes Count ( $\times 10^3/\mu\text{L}$ )	5.14 $\pm$ 0.866 <sup>bcd</sup>	6.34 $\pm$ 0.20 <sup>acd</sup>	1.62 $\pm$ 0.15 <sup>abd</sup>	3.62 $\pm$ 0.31 <sup>abc</sup>
Eosinophils Count ( $\times 10^3/\mu\text{L}$ )	0.844 $\pm$ 0.03 <sup>bcd</sup>	1.04 $\pm$ 0.05a <sup>cd</sup>	0.67 $\pm$ 0.04 <sup>abd</sup>	0.73 $\pm$ 0.04 <sup>abc</sup>



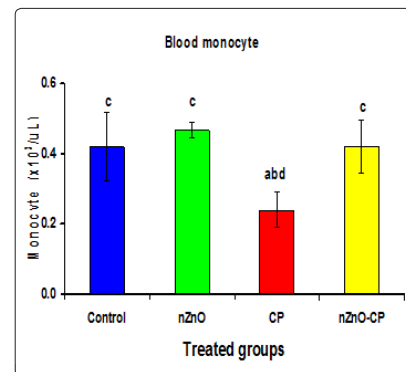
**Figure 4:** Effect of treatment of rat with zinc oxide nanoparticles (nZnO) and/or cyclophosphamide (CP) on neutrophils count ( $\times 10^6/\mu\text{L}$ ). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.



**Figure 5:** Effect of treatment of rat with zinc oxide nanoparticles (ZnONPs) and/or cyclophosphamide (CP) on lymphocytes count ( $\times 10^6/\mu\text{L}$ ). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.



**Figure 6:** Effect of treatment of rat with zinc oxide nanoparticles (nZnO) and/or cyclophosphamide (CP) on eosinophils count ( $\times 10^6/\mu\text{L}$ ). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.



**Figure 7:** Effect of treatment of rat with zinc oxide nanoparticles (ZnONPs) and/or cyclophosphamide (CP) on monocytes count ( $\times 10^6/\mu\text{L}$ ). Significance at  $P > 0.05$ . <sup>a</sup>Comparison of control and other groups; <sup>b</sup>Comparison of nZnO and other groups; <sup>c</sup>Comparison of CP and other groups; <sup>d</sup>Comparison of nZnO + CP and other groups.

## Conclusion

It can be concluded that CP induced changes in the hematological parameters. Treatment of rats with zinc oxide nano-particles and CP together ameliorated the toxicity induced by CP. These results may provide further visions into proper treatment of patients by improving side effects of chemotherapy. However further studies are necessary to establish optimal doses of nZnO and receive the best safety profile.

## References

1. Dollery A (1999) Cyclophosphamide In: Dollery C Editor Therapeutic drugs Edinburgh: Churchill Livingstone pp 349-353.
2. Walker RD, Sommerkamp H (1998) Hemorrhagic cystitis after high dose chemotherapy. An interdisciplinary problem. *Urologic A* 37: 516-521.
3. Kopečna L (2001) Late effects of anticancer therapy on kidney function in children with acute lymphoblastic leukemia. *Bratisl Lek Listy* 102: 357-360.
4. Shaunak S, Munro JM, Weinbren K, Walport MJ, Cox TM, et al. (1988) Cyclophosphamide induced liver necrosis: a possible interaction with azathioprine. *Q J Med* 67: 309-317.
5. Gustafsson LL, Eriksson L, Dahl, S, Eleborg, L (1996) Cyclophosphamide induced acute liver failure requiring transplantation in a patient with genetically deficient debrisoquine metabolism: a causal relationship? *J Int Med* 240: 311-314.
6. Iqbal A, Syed MA, Haque MM, Najmi AK, Ali J, et al. (2020) Effect of nerolidol on cyclophosphamide-induced bone marrow and hematologic toxicity in Swiss albino mice. *Exper Hematol* 82: 24-32.
7. Nafees S, Rashid S, Ali N, Hasan SK, Sultana S, et al. (2015) Rutin ameliorates cyclophosphamide induced oxidative stress and inflammation in Wistar rats: Role of NFκB/MAPK pathway. *Chem Biol Interact* 231: 98-107.
8. Brock N, Hohorst HJ (1967) Metabolism of cyclophosphamide. *Cancer* 20: 900-904.
9. Kotecha M, Veeman W, Rohe B, Tausch M (2006) NMR investigations of silane-coated nano-sized ZnO particles. *Microporous Mesoporous. Mat* 95: 66-75.
10. Beek WJE, Wienk MM, Janssen RAJ (2004) Efficient hybrid solar cells from zinc oxide nanoparticles and a conjugated polymer. *Adv Mater* 16: 1009-1013.
11. Berber M, Bulto V, Kliss R, Hahn H (2005) Transparent nanocrystalline ZnO films prepared by spin coating. *Scr Mater* 53: 547-551.
12. Jing LQ, Wang BQ, Xin BF, Li SD, Shi KY, et al. (2004) Investigations on the surface modification of ZnO nanoparticle photocatalyst by depositing Pd. *J Solid State Chem* 177: 4221-4227.
13. Jun JH, Seong H, Cho K, Moon BM, Kim S, et al. (2009) Ultraviolet photodetectors based on ZnO nanoparticles. *Ceram Int* 35: 2797-2801.
14. Brayner R, Ferrari Iliou R, Brivois N, Djediat S, Benedetti MF, et al. (2006) Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Lett* 6: 866-870.
15. Lin DH, Xing BS (2007) Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth. *Environ Pollut* 150: 243-250.
16. Premanathan M, Karthikeyan K, Jeyasubramanian K, Manivannan G (2011) Selective toxicity of ZnO nanoparticles toward Gram-positive bacteria and cancer cells by apoptosis through lipid peroxidation. *Nanomedicine: Nanotech. Biol Med* 7: 184-192.
17. Reddy KM, Feris K, Bell J, Wingett DG, Hanley C, et al. (2007) Selective toxicity of zinc oxide nanoparticles to prokaryotic and eukaryotic systems. *Appl Phys Lett* 90: 2139021-2139023.
18. Wang B, Feng WY, Wang M, Wang TC, Gu YQ, et al. (2008) Acute toxicological impact of nano- and submicro-scaled zinc oxide powder on healthy adult mice. *J Nanopart Res* 10: 263-276.
19. Zhu XS, Wang JX, Zhang XZ, Chang Y, Chen YS, et al. (2009) The impact of ZnO nanoparticle aggregates on the embryonic development of zebrafish (*Danio rerio*). *Nanotechnol* 20: 195103.
20. Koochi MK, Hejazy M, Najafi D, Sajadi SM (2017) Investigation of hematotoxic effect of nano ZnO, nano Fe<sub>3</sub>O<sub>4</sub> and nano SiO<sub>2</sub> in vitro. *Nanomed Res J* 2: 93-99.
21. Yahya RAM, Attia AM, El-Banna SG, El-Trass EE, Azab AE, et al. (2019) Hematotoxicity induced by copper oxide and/or zinc oxide nanoparticles in male albino rats. *J Biotechnol Bioeng* 3: 1-7.
22. Mayer A, Vadon M, Rinner B, Novak A, Wintersteiger R, et al. (2009) The role of nanoparticle size in hemocompatibility. *Toxicol* 258: 139-147.
23. Chang YN, Zhang M, Xia L, Zhang J, Xing G, et al. (2012) The toxic effects and mechanisms of CuO and ZnO nanoparticles. *Materials* 5: 2850-2871.
24. Roy R, Das M, Dwivedi PD (2015) Toxicological mode of action of ZnO nanoparticles: Impact on immune cells. *Mol Immunol* 63: 184-192.
25. Fahmey MA, Hassan NHA, El Fiky SA, Elalfy HG (2015) A mixture of honey bee products ameliorates the genotoxic side effects of cyclophosphamide. *Asian Pac J Trop Dis* 5: 638-644.
26. Badkoobeh P, Parivar K, Kalantar SM, Hosseini SD, Salabat A, et al. (2013) Effect of nano-zinc oxide on doxorubicin- induced oxidative stress and sperm disorders in adult male Wistar rats. *Iran J Reprod Med* 11: 355-364.
27. Howell DC (1995) Fundamental statistics for the behavioral sciences (3rd ed) Duxbury press. An imprint of Wads Worth publishing company Belmont. California 1995: 163-166.
28. Davila JC, Lenherr A, Acosta D (1989) Protective effect of flavonoids on drug-induced hepatotoxicity in vitro. *Toxicol* 57: 267-286.
29. Abraham P, Indirani K, Sugumar E (2007) Effect of cyclophosphamide treatment on selected lysosomal enzymes in the kidney of rats. *Exp Toxicol Pathol* 59: 143-149.
30. Ballantyne B (1988) Xenobiotic-induced rhabdomyolysis. In: Ballantyne B (Ed), *Perspectives in Basic and Applied Toxicology*, Butterworth and Co Ltd: pp70-153.
31. Chang MS, Kim do R, Ko EB, Choi BJ, Park SY, et al. (2009) Treatment with Astragal radix and Angelicae radix enhances erythropoietin gene expression in the cyclophosphamide-induced anemic rat. *J Med Food* 12: 637-642.
32. Groopman JE, Utri LM (1999) Chemotherapy-induced anemia in adults: Incidence and treatment. *J Natl Cancer Inst* 91: 1616-1634.
33. Orlova M, Orlov A (2011) Role of zinc in an organism and its influence on processes leading to apoptosis, *Br. J Med Med Res* 1: 239-305.
34. Ayhanci A, Uyar R, Aral E, Kabadere S, Appak S, et al.

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- (2008) Protective effect of zinc on cyclophosphamide-induced hematotoxicity and urotoxicity, *Biol Trace Elem Res* 126: 186-193.
35. Doz F, Roosen N, Rosenblum ML (1993) Metallothionein and anticancer agents: the role of metallothionein in cancer chemotherapy. *J Neurooncol* 17: 123-129.
36. Suzuki CAM, Ohta H, Albores A, Koropatnick J, Cherian MG, et al. (1990) Induction of metallothionein synthesis by zinc in cadmium pretreated rats. *Toxicol* 63: 273-284.
37. Winkelstein A (1973) Mechanisms of immunosuppression: Effects of cyclophosphamide on cellular immunity. *Blood* 41: 273-284.

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