

Amyloid, A Jekyll and Hyde Molecule, Induces Neuronal Decline and Cognitive Dysfunction but is also a Unique Molecular Template used in Nano-Electronics, Light Capture Photovoltaics, Biosensors and in Neuromorphic Computing

Margaret M Smith^{1,2} and James Melrose^{1,3,4*}

¹Raymond Purves Bone and Joint Research Laboratory, Kolling Institute, St. Leonards, NSW 2065, Australia

²Arthroparm/Sylvan Scientific Pty Ltd, Bondi Junction, Sydney, NSW 2022, Australia

³School of Medical Sciences, Faculty of Medicine and Health, The University of Sydney at Royal North Shore Hospital, St. Leonards, NSW 2065, Australia

⁴Graduate School of Biomedical Engineering, Faculty of Engineering, University of New South Wales, Sydney, NSW 2052, Australia

*Corresponding Author

Prof. James Melrose, Raymond Purves Laboratory, Level 10, Kolling Institute of Medical Research B6, Royal North Shore Hospital, St. Leonards, NSW 2065, Australia.

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Abstract

This study, examined the self-assembly of amyloid peptides β -40 or β -42 to form neurotoxic plaque and neurofibrillary tangles, neuronal dysfunction and diseases of cognitive decline (e.g. Alzheimer's, Parkinson's and Huntington's disease). However, not all amyloids are toxic, functional amyloids are non-toxic ordered structures that have been used in tissue engineering applications. Long and hollow amyloid fibres and flattened tube and spiral ribbon-like structures are diverse self-assembling structures that have found application in the development of next generation computers and biosensors, ultracapacitors, memristors, actuators, molecular switches, artificial synapses and in photoelectric photon capture light harvesting technologies in nano-photoelectronics, photovoltaics and photo-pharmacological drug regulation. Hybrid $A\beta(16-22)$ - α -synuclein amyloid fibrils also exhibits light-harvesting and electron-transfer properties. Engineered amyloid assemblies are thus driving futuristic developments in nanotechnology. With a better understanding of amyloid fibril assembly clues are also being uncovered as to how the toxic build up of amyloid deposits in brain tissues and resultant diseases of cognitive decline may be prevented. The elevated incidence of neurological diseases in the ageing general global population, points to a clear need to find a remedy for these debilitating conditions.

Keywords: Amyloid Fibrils, Nano-Electronics, Memristors, Bio-Sensors, Actuators, Molecular Switches, Photovoltaics, Artificial Synapses, Functional Amyloids, Neuromorphic Computing

1. Introduction

The aim of this study was to review the roles of amyloid protein aggregates in neurodegeneration and contrast this with beneficial aspects of its structural organization which have found application in nanobiology in several innovative areas of tissue engineering.

1.1. Definition of Amyloid

Amyloid is an insoluble protein aggregate formed from misfolded proteins containing β -pleated sheets and can be stained with Congo red, exhibiting a characteristic green-yellow birefringence under polarized light [1-6]. These misfolded proteins exhibit specific features that facilitate auto-assembly of ordered repeat structures in insoluble amyloid deposits [2-4]. $A\beta$ peptides released from

appican (APP) in brain tissues are assembled into dimers, trimers, oligomers and distinctive fibrillar structures. Amyloid deposits occur in many tissues in the human body other than the brain, leading to organ and tissue dysfunction [7,8]. Amyloid refers to the insoluble protein aggregates that accumulate in the aging brain in diseases of cognitive decline such as Alzheimer's (AD) and Parkinson's (PD). Amyloid can arise from over 36 proteins in various parts of the body in different human diseases [7-9]. Amyloid deposits in the brain display distinctive morphologies formed by β -40 or β -42 peptides. Amyloid plaques are visible by light microscopy using a variety of staining procedures, including silver stains, Congo red, Thioflavin, cresyl violet and periodic acid-Schiff (PAS) procedures which stain different components of amyloid plaques, with variable sensitivity [7-10]. Immunolocalization of amyloid plaques with specific antibodies to A β epitopes and to other amyloid-associated components has also been used to visualize amyloid formations [11]. Examination of human autopsy samples and tissues from experimental models of AD have identified the biochemical, cytological, and inflammatory processes involved in the generation of these amyloid plaques [7]. Amyloid fibrils are diverse molecular structures, ENTAIL and PARROT are two information systems developed for the classification of amyloid fibril biodiversity [12-14]. Ultrasensitive, new generation amyloid biosensors have also been developed for the detection of amyloid peptides in tissues, plasma and cerebrospinal fluid [15-22]. These show amyloid plaque formation may be linked to trauma of the brain microvascular and immune system [23]. Chronic brain inflammation and immune dysfunction may accelerate amyloid deposition [24,25].

A β 40 and A β 42 oligomers of A β peptides in AD only differ by two C-terminal residues, however A β 42 aggregates assemble much faster than A β 40 structures and is more toxic, forming pore-like structures in cell membranes [26-28]. The additional C-terminal residues in A β 42 allow electrostatic interactions which stabilize the β -hairpin promoting dimer formation and oligomerization through intermolecular β -bridge formation [29,30]. A combination of hydrophobic and charged amino acids in A β peptides contribute to aggregate formation; glycosaminoglycans (GAGs) also act at the earliest stages of fibril formation, namely during amyloid-beta nucleation [31]. The amyloid plaques in Alzheimer's brains consist mainly of A β 42 and some plaques contain only A β 42, even though A β 40 concentrations in cerebrospinal fluids is several-fold higher than A β 42 in the brain.

1.2. Amyloid Aggregates Promote Neurodegeneration and Cognitive Decline, However Functional Amyloids are Non-Toxic and have Beneficial Properties

The amyloid cascade hypothesis proposes that dysfunctional metabolism of amyloid precursor protein (APP) initiates the pathogenesis of AD [32]. This results in aggregation of A β , formation of neuritic plaques which cause pathological changes through formation of neurofibrillary tangles that disrupt normal synaptic connections and compromised neuronal synaptic activity, neuronal cell death, cognition, memory problems and

development of dementia and cognitive decline in AD and PD [32,33-36]. However, not all amyloids are toxic nor do they disrupt normal organ functions [37]. So-called functional amyloids have beneficial properties through unique structural features and have been receiving considerable attention in tissue engineering applications in highly innovative areas of nanobiology [38]. From an engineering perspective, the self-assembling regular and tight packing structure of amyloids provides strength, and are useful molecular templates amenable to structural modification and application in tissue engineering. Such self-assembled networks are highly suited to biomaterial scaffold developments [39-42]. Amyloids are some of the strongest protein structures ever identified in nature [43,44]. Spider-web drag-line silk is composed of repeat peptide modules with a high β -sheet content on a weight for weight basis web silk has a strength exceeding that of high quality structural steel [45-47].

2. Amyloid Precursor Protein and Its Bioactive Fragments

Appican is a transmembrane chondroitin sulfate-proteoglycan precursor, containing an embedded amyloid precursor protein (APP) module within its core protein and is a 110-130kDa type 1 transmembrane glycoprotein [48,49]. APP is also a component of APP-like proteins 1 and 2 (APLP-1, 2) and these constitute a family of mammalian membrane proteins. However, unlike APP, the APLPs lack the A β sequence, and do not give rise to the AD [50]. APP and APLP regulate synaptic transmission, plasticity, and calcium homeostasis, and are important cell messengers and regulators of neural activity in neurotransduction and are neuroprotective, soluble sAPP α counteracts the deleterious effects of the APP-derived A β 40 and 42 peptides that lead to amyloid deposition [51-54].

2.1. The Enigmatic and Perplexing Story of APP Processing in Brain Tissues

APP is a type I transmembrane glycoprotein notorious for its involvement in the pathogenesis of AD through β A40 and β A42 peptides generated by an amyloidogenic protease pathway in the brain involving β -secretase (BACE-1, β -site APP cleaving enzyme, mepapsin 2) and γ -secretase [55]. β A40 and β A42 peptides self-assemble to form insoluble plaques and neurofibrillary tangles which detrimentally impact on synaptic function, neuron viability and cognitive processes in the brain, leading to dementia [56-58]. APP, however, is also processed by α -secretase via a non-amyloidogenic pathway, generating the soluble amyloid precursor sAPP α peptide [55]. sAPP α promotes neuroprotection, synaptic plasticity, memory formation, neurogenesis, neuritogenesis, and reduces amyloid and tau pathology [59-61]. Several studies also suggest that sAPP α regulates the trafficking of APP and its processing by proteases and may decrease the risk of developing AD [62]. A β monomers share similar properties to sAPP α however, when self-assembled into A β oligomers, they become neurotoxic [61]. The precise physiological functions of full length APP and its proteolytic fragments is thus a complex story, A β production is not deleterious per se but when assembled into oligomers β A is toxic [63-66].

3. Identification of Proteins with Amyloid Aggregative Potential

The propensity of a peptide to form an amyloid fibril is dependent on factors such as polypeptide charge, sequence, hydrophobicity and peptide secondary structure and not by a specific “amyloid” amino-acid sequence per se [67-73]. Amyloidogenic peptides self-assemble into repetitive stacked structures that attract further proteins to propagate fibril formation [74]. Predictive algorithms have been developed to assess peptides that display a propensity to form amyloid fibrils in web-based software that predicts aggregation-prone protein sequences [75-77]. AMYLPRED2 (<http://biophysics.biol.uoa.gr/AMYLPRED2>) is a public web tool for the prediction of amyloidogenic determinants in 'aggregation-prone' peptide sequences within proteins [76]. Development of methods to produce controlled amyloid fibrils *in vitro* is an important innovation and opens the door for the design of new biomaterials exploiting the superior structural properties of amyloid fibrils and the inherent diversity of peptide sequences [78]. A number of bioinformatics and computational studies have examined the peptide sequences of amyloid proteins to better understand the aggregation process and to determine peptide sequences that initiate and propagate assembly of fibrillar material [79-97].

4. The Attributes of Functional Amyloids

In nature, amyloids have a range of functions across mammals, bacteria, fungi and marine organisms in beneficial physiological processes such as regulation of pigment formation, storage and controlled release of peptide hormones, memory, fertilization of oocytes by sperm, antimicrobial responses, regulated necrosis, cellular responses to stress and powerful adhesive properties [98-103]. Naturally occurring amyloids are also associated with a number of disease processes (Table 1A). Amyloid fibrils are found in barnacle and mussel bioadhesives with powerful adhesive properties that have inspired the development of tissue adhesives of potential application in highly specialized surgical procedures [104]. The unique architectural assembly processes and exceptional mechanical strength of amyloid fibrils makes these structures of interest in innovative applications in organic micro-circuitry in nano-electronics, in the development of actuators, molecular switches, memristors and microcomputers [105-114]. The low power requirements and ultra-high speed signal transmittance capability of memristors is revolutionising development of neuromorphic circuits that are used in synthetic neural networks, switching devices and low-power sensors in microcomputing [115-117]. Nano-wires prepared in hollow amyloid fibril casting templates have been used in bio-sensing,

optoelectronics and photovoltaics and show potential in the development of synthetic synapses in highly innovative bio-nanotechnological applications [118-120]. Amyloid fibrils have found a number of applications in tissue engineering (Table 1B) [121]. Photobiomodulation therapy, using near infra-red 700-1400 nm low-level laser phototherapy, reduces the deposition of β A in the AD brain, ameliorating neuroinflammation and oxidant stress, supporting mitochondrial homeostasis to elicit a healing or regenerative response [122]. The surface chemistry of engineered amyloid fibrils can be modified depending on the amino acids used and bacterial expression systems used in their assembly; fibrils can also be coated with chemicals that modify their responsiveness to specific chemical microenvironments and their light capture properties [123-125]. Amyloid fibrils can also be coated with gold nanomaterials modified with peptides or other chemicals. Gold is chemically inert but highly conductive and, when modified with additional components, can fine-tune the surface interactivity of amyloid fibrils. Some chemicals (PEDOT-S, Poly(3,4-ethylenedioxythiophene) polystyrene sulfonate; PPF, luminescent polyfluorene; APFO, Ammonium pentadecafluoroctanoate) can improve the light harvesting and electron transfer properties of fibrils to improve the efficiency of photobiomodulation therapy [126]. Amyloid fibrils have UV-visible–near-infrared optical light capture properties that have been applied in pharmaceutical healthcare applications [127,128]. Light capture technologies can be used to activate photoswitchable drugs in the regulation of neural pain generation. Systemic pharmacotherapeutic neuroinhibitory medications like antiseizure drugs, used to treat epilepsy and neuropathic pain also have unwanted side-effects, there is an urgent need to develop pain alleviating drugs that do not display such unwanted side-effects. Photopharmacology uses light-activated drugs illuminated locally at specific tissue target sites to provide specificity of action. Photoswitchable derivatives (carbazopine-1, carbamazepine) of the antiseizure drug carbamazepine (tegretol) are used to treat tonic-clonic seizures and bipolar disorder and to relieve intense, stabbing, electric shock-like pain caused by trigeminal neuralgia (douloureux, Fothergill disease). Light emitting semi-conductor diodes (LEDs) that deliver light of 400-590nm can be used to activate carbamazepine, the photo-switched drug has been shown to provide specific analgesic mechanical and thermal pain relief profiles in a rat model of neuropathic pain [129]. Amyloid fibril optical biosensors and smart Trojan-horse technology has improved tissue light delivery precision to photoactivate pain alleviating drugs in photomedicine and have also been examined for the eradication of α -amyloid fibril deposition in tissues [128,130-135].

Disease process	Amyloid precursor protein	Amyloid monomer
AD	APP	A β
Atrial amyloidosis	Atrial natriuritic protein	Amyloid ATF
Spongiform encephalopathy	Prion protein PrP c	PrP s c
Primary systemic amyloidosis	Ig L and H chains	AL, AH
Secondary systemic amyloidosis	Apo serum amyloid A	SAA
Familial amyloid polyneuropathy I	Transthyretin	ATTR
Familial amyloid polyneuropathy II	ApoA	AApoA
Haemodialysis amyloidosis	β 2-microglobulin	A α β 2M
Hereditary systemic amyloidosis	Lysozyme	ALys
Diabetes type II	ProIAPP	APP/amylin
Insulin injection amyloidosis	Insulin	AIns
Cerebral amyloid angiopathy	Cystatin C	ACys
Finnish hereditary systemic amyloidosis	Gelsolin	AGel
Age associated pituitary prolactinomas	Prolactin	APro
Familial amyloidosis	Fibrinogen α A chain	AFib

Table 1 A: Naturally occurring amyloids associated with specific disease processes [9,38,136]

Amyloid	Applications	Reference
Transthyretin peptide (105-115)	Cell adhesive properties	137
Gonadotropin releasing hormone	Long acting peptide/protein drug depot/delivery	138
<i>Candida albicans</i> Killer decapeptide	Auto-delivery of therapeutic peptides	139
Yeast Sup35p NM prion domain	Nanowire development	140
α -synuclein fibrils	Enzyme entrapment hydrogel	141
insulin fibril semi-conductor oligoelectrolyte	Optoelectronic nanowire assembly	142
PPF coated Insulin fibrils	Polymer light emitting diode fibrils	143
APFO-12 coated Insulin fibrils	Optical nanowires	144
PEDOT-S coated Insulin fibrils	Conductive nanowires	145
β 2-microglobulin	Nanoporous cell support matrix	146

Table 1B: Functional Amyloids used in bio-nanotechnology applications

Abbreviations: PEDOT-S, Poly(3,4-ethylenedioxythiophene) polystyrene sulfonate; PPF, luminescent polyfluorene; APFO, Ammonium pentadecafluoroctanoate.

5. Amyloid Fibrils as Building Blocks for Innovative Biomaterials

Protein assemblies based on amyloid core structures display diverse biological functions with potential application in futuristic self-assembling biomaterials [41,126,136,147-150]. The roles of amyloid fibrils in a range of human diseases of cognitive decline are well known, however, so-called functional amyloids have low toxicity and beneficial properties in a range of physiologic processes (eg, long-term memory formation, gradual release of stored peptide hormones) and roles in cell fate and cell-signalling cascades in mammals, bacteria and fungi [74,151-154]. Functional amyloids display many features that make them attractive candidates for tissue engineering forming self-assembled regularly organised structures with impressive biophysical properties in

extended straight filaments, tapes, twisted ribbons, and hollow tubes [45,121,155]. Fibrils are typically 5–20 nm in diameter with a length in the micron size range irrespective of the many different proteins from which they are assembled. Fibrillar assembly occurs over a wide range of temperature, pH and solvent conditions providing flexibility in the procedures that may be utilised in the laboratory in engineering applications to assemble fibrils. Introduction of point mutations in fibrillar structures can be used to design fibrils with variable chemical surface and electrostatic characteristics with diverse binding properties and unique environmental responsiveness [150,156]. Amyloid fibrils possess a Young's Modulus in the GPa scale and a strength comparable to steel. High-resolution data gathered from X-ray diffraction and

NMR images, demonstrate the presence of extensive cross β -sheet structure within the core of the fibril which forms an expansive hydrogen-bonding network spanning the length of the fibril. It is this cooperative intermolecular hydrogen-bonding network in the fibril that confers stability and unique material properties [43,157].

Hollow ~100 nm nanotubes have been developed using self-assembly of amyloid-like fibrillar structures to form templates within which silver nanowires can be cast of 10-nm width and lengths ranging from 60 to 100 microns with the central nanowire recovered by proteolytic dissolution of the peptide shell [109,110,158]. Multi-layered co-axial nano-wire assemblies have also been prepared decorated on the exterior of the nanowire with metallic gold, nanowire structures have been confirmed by TEM and energy dispersive X-ray analysis [108]. Gold or silver nanowires with widths of ~100 nm have demonstrated high conductivities and low resistances (~80 Ω), their use in the microcircuitry of nanoelectronics have revolutionised development of next generation computers and biosensors [159]. Ultracapacitors have also been developed using an external magnetic field to orient horizontal and vertical arrangements of nanotubes which display enhanced capacitance relative to carbon and carbon nanotube-modified electrodes [107].

5.1. Amyloid Cell Attachment Matrices and Hydrogels for Cell Delivery in Tissue Repair Strategies

The ECM has important instructive properties over cellular activity. Instructive communication between the cells and ECM is encoded in peptide epitopes of structural and signaling components that act as molecular directors of cellular activity [160]. Peptide epitopes can be incorporated into synthetic biomatrices to mimic the specific communication that occurs between cells and tissues to control cell adhesion, differentiation, immunomodulation and ECM turnover to achieve tissue homeostasis. Self-assembled amyloid peptide biomatrices have been prepared with RGD motifs to improve cellular attachment [161]. Lysozyme amyloid micronetworks support chondrocyte viability and ECM deposition, while α -synuclein and β -lactoglobulin matrices maintain cell viability [160]. Synthetic amyloid biomatrices with specific added proteins hold promise in cartilage regeneration [162]. Amyloid scaffolds represent versatile biomaterials for cell adhesion and tissue engineering applications [41].

In tissues, amyloid fibrils also promote neural proliferation and neurite outgrowth through interaction with ECM components [65,163]. Amyloid fibril 3D scaffolds have been used to culture neurons and used to develop a model of AD [164]. Amyloid fibril bioscaffolds have been used to culture neural progenitor cells to assess if an A $\beta\beta$ sheet environment could be used to guide differentiation of cultured neural progenitor cells simulating conditions found in brain tissues in which amyloid deposition was evident producing neural progenitor cells of a phenotype evident in amyloid containing AD tissues [165].

Amyloid fibril hydrogels have also been prepared and shown to

be suitable as cell delivery vehicles for cultured stem cells which have undergone cellular differentiation [166,167]. Such hydrogels are smart materials capable of transitioning to altered rheological properties in response to spatiotemporal changes in temperature, pH, ionic strength, and external stresses and are also suitable for drug delivery and stem cells in regenerative procedures [137,168].

5.2. Application of Amyloid Fibrils in Nanobiology and Organic Microelectronics

Amyloid fibrils have found application in photoelectric and light harvesting technologies [169]. A peptide fragment of transerythrin (TTR 105-115) spontaneously forms highly aligned uniform fibrils in which fluors can be incorporated with little disruption in fibril structure to provide an amyloid fibril with the ability to capture photons and harvest light [170]. A β (16–22)- α -synuclein hybridised amyloid fibrils also exhibit light-harvesting and electron-transfer properties [171].

While naturally occurring amyloids are robust self-assembled nanometer-sized fibril biomaterials, it is now possible using in-vitro methods to assemble large 10-20 μm diameter amyloid fibers several mm in length [172]. Using a short, hydrophobic director α -helical template peptide and mixtures of peptides it is possible to self-assemble large amyloid fibers, encoded by micron-sized self-assembled structures at the genetic level, with tailored rectangular or cylindrical cross-sectional morphologies and robust material properties (modulus 0.1-2.5 GPa) [173,174]. A template peptide expressed in *E. coli* has been used to grow customized 200-500 nm amyloid fibrils and 7-20 μm wide fibers for tissue engineering.

5.3. Amyloid Fibrillar Assemblies and Neuromorphic Computing

Quantum computers offer the computational power required to drive neuromorphic hardware in neural network dynamic simulations [175]. Machine learning and artificial intelligence algorithms running on neuromorphic hardware by quantum computers are being developed to assist in data analysis in artificial synapse modular supercomputing developments. Self-assembling amyloid fibrillar structures can be modeled to provide neuromorphic hardware due to varied fibril surface chemistry and their responsiveness to specific electrochemical microenvironments. This may be useful in the development of electrochemical random-access memory using ionic neurotransistors, leading to neuromorphic computing networks that drive sensory intelligent perception systems [176,177].

5.4. HS-Amyloid Interactions in Pathological Neuronal Tissue

While deposition of amyloid as neuritic plaques and fibrillary tangles in AD is a self-assembly process an ongoing debate over the last 30 years over whether HSPGs also have roles to play in amyloid aggregation has finally been resolved and it is now accepted that HSPGs also promote the amyloid aggregation process and this affects the functional status of brain tissues [178-180]. HS has multiple roles in the regulation of amyloidogenesis, a range of synthetic HS oligosaccharides have been prepared to examine amyloid-HS interactions [181-185]. HS from porcine

mucosa has also been shown to promote amyloid-beta clearance in APP/PS1 mice alleviating the pathological features of AD in the mouse [186]. Brain HSPGs are differentially expressed in AD and a HS code has been established for neurodegenerative changes that accompany amyloid deposition [187,188]. Dystroglycan-HSPG interactions also regulate synaptic plasticity and specificity maintaining normal neuronal functional activity [189]. A further HS code has been shown to independently regulate synuclein aggregation dynamics [190]. 6-O-Phosphorylated HS oligosaccharides have also been synthesized and shown to inhibit amyloid β aggregation suggesting the charge status of GAGs are important in such regulatory processes [191]. KS is another highly charged instructive GAG in neuronal tissues and has neuroregulatory instructive properties however it is not known if it regulates amyloid aggregative processes or specifically regulates neuronal activity in pathological tissues in AD [192-194].

GAGs have been proposed as attractive platforms for energy devices and flexible electronics and may be particularly relevant to the electro-responsive neuron [195]. A range of proteoglycans and hyaluronan organize the neuronal micro-environment ensuring that ion fluxes, gradients and micro-niches are maintained in brain tissues ensuring optimal neuronal activity and viability [196,197].

6. Application of Engineered Amyloid Polymers in Biomedicine

A number of studies have reviewed the diverse range of biomedical applications that have been developed using functional amyloids in biomedicine [198-201]. These applications range from development of cell-culture bioscaffolds and hydrogels for tissue engineering, real-time bidirectional control systems between living brains and actuators of motor and sensory neuromorphic drug delivery systems, immune therapies, biosensor development and $A\beta$ bio-imaging applications, anti-cancer therapies, photopharmacology, light-capture photovoltaics, nanoelectronics, nano-photoelectronics, ultracapacitors, memristors, artificial synapses, molecular switches and implantable neural communication interface technologies [198-201].

7. Concluding Remarks and Future Studies

This review has illustrated the toxic features of amyloid plaques and neurotangles in brain tissues which lead to neurodegeneration, impaired neuron synaptic function and consequential diseases of cognitive decline. The beneficial properties of functional amyloids are also reviewed and engineering applications that have been developed using the amyloid fibril as a molecular template which can be manipulated in a number of innovative applications in significant advancements in nano-technology. These have made important contributions to neuro-signaling in neural repair strategies and in areas of bio-regulation using nano-electronics. Development of biosensors, actuators, ultracapacitors, memristors, molecular switches and next generation micro-computing technologies are examples of these innovative applications that have been applied in bioregulation. The high conductance of nano-wires cast in hollow amyloid fibril molecular templates, low resistance and ultra-high signaling capability of such structures makes them

particularly useful in nano-electronic applications. Furthermore, engineered amyloid fibril assemblies have also found application in photoelectric and photon capture light harvesting technologies of application in innovative nano-photoelectronics and photovoltaic applications which promise to revolutionise specific areas of optical neuro-regulatory processes. The application of AI and quantum computer methodologies in brain-interface technologies offers particularly exciting possibilities in the improvement of real-time bidirectional control systems between living brains and actuators in motor and sensory neuromorphic applications and have had notable clinical success in the treatment of paralyzed patients' and expanded the mobility of disabled patients [202]. We have entered an exciting era in bioregulation of neuroregulatory processes.

References

1. Benson, M. D., Buxbaum, J. N., Eisenberg, D. S., Merlini, G., Saraiva, M. J., Sekijima, Y., ... & Westermark, P. (2018). Amyloid nomenclature 2018: recommendations by the International Society of Amyloidosis (ISA) nomenclature committee. *Amyloid*, 25(4), 215-219.
2. Ke, P. C., Zhou, R., Serpell, L. C., Riek, R., Knowles, T. P., Lashuel, H. A., ... & Mezzenga, R. (2020). Half a century of amyloids: past, present and future. *Chemical Society Reviews*, 49(15), 5473-5509.
3. Buxbaum, J. N., & Linke, R. P. (2012). A molecular history of the amyloidoses. *Journal of molecular biology*, 421(2-3), 142-159.
4. Puchtler, H., & Sweat, (1966). A review of early concepts of amyloid in context with contemporary chemical literature from 1839 to 1859. *Journal of Histochemistry & Cytochemistry*, 14(2), 123-134.
5. Sipe, J. D., & Cohen, A. S. (2000). History of the amyloid fibril. *Journal of structural biology*, 130(2-3), 88-98.
6. Yakupova, E. I., Bobyleva, L. G., Vikhlyantsev, I. M., & Bobylev, A. G. (2019). Congo Red and amyloids: history and relationship. *Bioscience reports*, 39(1), BSR20181415.
7. Walker, L. C. (2020). $A\beta$ plaques. *Free neuropathology*, 1, 1-31.
8. Westermark, G. T., Fändrich, M., Lundmark, K., & Westermark, P. (2018). Noncerebral amyloidoses: aspects on seeding, cross-seeding, and transmission. *Cold Spring Harbor perspectives in medicine*, 8(1), a024323.
9. Fowler, D. M., Koulov, A. V., Alory-Jost, C., Marks, M. S., Balch, W. E., & Kelly, J. W. (2006). Functional amyloid formation within mammalian tissue. *PLoS biology*, 4(1), e6.
10. Lamy, C., Duyckaerts, C., Delaere, P., Payan, C. H., Fermanian, J., Poulain, V., & Hauw, J. J. (1989). Comparison of seven staining methods for senile plaques and neurofibrillary tangles in a prospective series of 15 elderly patients. *Neuropathology and applied neurobiology*, 15(6), 563-578.
11. Menter, T., Bachmann, M., Grieshaber, S., & Tzankov, A. (2016). A more accurate approach to amyloid detection and subtyping: combining *in situ* Congo red staining and immunohistochemistry. *Pathobiology*, 84(1), 49-55.

12. Li, D., & Liu, C. (2020). Structural diversity of amyloid fibrils and advances in their structure determination. *Biochemistry*, 59(5), 639-646.
13. Auriemma Citarella, A., Di Biasi, L., De Marco, F., & Tortora, G. (2022). ENTAIL: yEt aNoTher amyloid fibrils cLassifier. *BMC bioinformatics*, 23(1), 517.
14. Griffith, D., & Holehouse, A. S. (2021). PARROT is a flexible recurrent neural network framework for analysis of large protein datasets. *Elife*, 10, e70576.
15. Zhang, Y., Ren, B., Zhang, D., Liu, Y., Zhang, M., Zhao, C., & Zheng, J. (2020). Design principles and fundamental understanding of biosensors for amyloid- β detection. *Journal of Materials Chemistry B*, 8(29), 6179-6196.
16. Kaushik, A., Jayant, R. D., Tiwari, S., Vashist, A., & Nair, M. (2016). Nano-biosensors to detect beta-amyloid for Alzheimer's disease management. *Biosensors and bioelectronics*, 80, 273-287.
17. Gao, H., Chen, J., Huang, Y., & Zhao, R. (2024). Advances in targeted tracking and detection of soluble amyloid- β aggregates as a biomarker of Alzheimer's disease. *Talanta*, 268, 125311.
18. Sharma, P. K., Kim, E. S., Mishra, S., Ganbold, E., Seong, R. S., Kim, Y. M., ... & Kim, N. Y. (2022). Ultrasensitive probeless capacitive biosensor for amyloid beta (A β 1-42) detection in human plasma using interdigitated electrodes. *Biosensors and Bioelectronics*, 212, 114365.
19. Zheng, Y., Zhang, L., Zhao, J., Li, L., Wang, M., Gao, P., ... & Wang, W. (2022). Advances in aptamers against A β and applications in A β detection and regulation for Alzheimer's disease. *Theranostics*, 12(5), 2095.
20. Jeong, D., Kim, J., Chae, M. S., Lee, W., Yang, S. H., Kim, Y., ... & Hwang, K. S. (2018). Multifunctionalized reduced graphene oxide biosensors for simultaneous monitoring of structural changes in amyloid- β 40. *Sensors*, 18(6), 1738.
21. Wang, X., Li, L., Gu, X., Yu, B., & Jiang, M. (2021). Switchable electrochemical aptasensor for amyloid- β oligomers detection based on triple helix switch coupling with AuNPs@ CuMOF labeled signaling displaced-probe. *Microchimica Acta*, 188, 1-11.
22. Xing, Y., & Xia, N. (2015). Biosensors for the determination of amyloid-beta peptides and their aggregates with application to Alzheimer's disease. *Analytical Letters*, 48(6), 879-893.
23. Bu, X. L., Xiang, Y., Jin, W. S., Wang, J., Shen, L. L., Huang, Z. L., ... & Wang, Y. J. (2018). Blood-derived amyloid- β protein induces Alzheimer's disease pathologies. *Molecular psychiatry*, 23(9), 1948-1956.
24. Heppner, F. L., Ransohoff, R. M., & Becher, B. (2015). Immune attack: the role of inflammation in Alzheimer disease. *Nature Reviews Neuroscience*, 16(6), 358-372.
25. Jorfi, M., Maaser-Hecker, A., & Tanzi, R. E. (2023). The neuroimmune axis of Alzheimer's disease. *Genome Medicine*, 15(1), 6.
26. Brody, D. L., Magnoni, S., Schwetye, K. E., Spinner, M. L., Esparza, T. J., Stocchetti, N., ... & Holtzman, D. M. (2008). Amyloid- β dynamics correlate with neurological status in the injured human brain. *Science*, 321(5893), 1221-1224.
27. Serra-Batiste, M., Ninot-Pedrosa, M., Bayoumi, M., Gairí, M., Maglia, G., & Carulla, N. (2016). A β 42 assembles into specific β -barrel pore-forming oligomers in membrane-mimicking environments. *Proceedings of the National Academy of Sciences*, 113(39), 10866-10871.
28. Kim, W., & Hecht, M. H. (2005). Sequence determinants of enhanced amyloidogenicity of Alzheimer A β 42 peptide relative to A β 40. *Journal of Biological Chemistry*, 280(41), 35069-35076.
29. Itoh, S. G., Yagi-Utsumi, M., Kato, K., & Okumura, H. (2022). Key residue for aggregation of amyloid- β peptides. *ACS Chemical Neuroscience*, 13(22), 3139-3151.
30. Hsu, F., Park, G., & Guo, Z. (2018). Key residues for the formation of A β 42 amyloid fibrils. *ACS omega*, 3(7), 8401-8407.
31. McLaurin, J., Franklin, T., Zhang, X., Deng, J., & Fraser, P. E. (1999). Interactions of Alzheimer amyloid- β peptides with glycosaminoglycans: Effects on fibril nucleation and growth. *European journal of biochemistry*, 266(3), 1101-1110.
32. Hardy, J. A., & Higgins, G. A. (1992). Alzheimer's disease: the amyloid cascade hypothesis. *Science*, 256(5054), 184-185.
33. Hardy, J., & Allsop, D. (1991). Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends in pharmacological sciences*, 12, 383-388.
34. O'brien, R. J., & Wong, P. C. (2011). Amyloid precursor protein processing and Alzheimer's disease. *Annual review of neuroscience*, 34(1), 185-204.
35. Chiellini, G. (2021). Understanding Amyloid Structures and Disease: A Continuing Challenge in Health Research. *International Journal of Molecular Sciences*, 22(12), 6620.
36. Dos Santos, H. M., Bertollo, A. G., Mingoti, M. E. D., Grolli, R. E., Kreuz, K. M., & Ignácio, Z. M. (2024). Dementia and depression: biological connections with amyloid β protein. *Basic & Clinical Pharmacology & Toxicology*, 134(5), 563-573.
37. Jackson, M. P., & Hewitt, E. W. (2017). Why are functional amyloids non-toxic in humans?. *Biomolecules*, 7(4), 71.
38. Rubel, M. S., Fedotov, S. A., Grizel, A. V., Sopova, J. V., Malikova, O. A., Chernoff, Y. O., & Rubel, A. A. (2020). Functional mammalian amyloids and amyloid-like proteins. *Life*, 10(9), 156.
39. Reynolds, N. P. (2019). Amyloid-like peptide nanofibrils as scaffolds for tissue engineering: Progress and challenges. *Biointerphases*, 14(4), 040801.
40. Mankar, S., Anoop, A., Sen, S., & Maji, S. K. (2011). Nanomaterials: amyloids reflect their brighter side. *Nano reviews*, 2(1), 6032.
41. Das, S., Jacob, R. S., Patel, K., Singh, N., & Maji, S. K. (2018). Amyloid fibrils: Versatile biomaterials for cell adhesion and tissue engineering applications. *Biomacromolecules*, 19(6), 1826-1839.
42. Binaymotlagh, R., Chronopoulou, L., & Palocci, C. (2023). Peptide-based hydrogels: template materials for tissue engineering. *Journal of Functional Biomaterials*, 14(4), 233.

43. Knowles, T. P., & Buehler, M. J. (2011). Nanomechanics of functional and pathological amyloid materials. *Nature nanotechnology*, 6(8), 469-479.
44. Sweers, K. K., Bennink, M. L., & Subramaniam, V. (2012). Nanomechanical properties of single amyloid fibrils. *Journal of Physics: Condensed Matter*, 24(24), 243101.
45. Li, J., & Zhang, F. (2021). Amyloids as building blocks for macroscopic functional materials: designs, applications and challenges. *International journal of molecular sciences*, 22(19), 10698.
46. Fukuma, T., Mostaert, A. S., & Jarvis, S. P. (2006). Explanation for the mechanical strength of amyloid fibrils. *Tribology Letters*, 22, 233-237.
47. Sawaya, M. R., Hughes, M. P., Rodriguez, J. A., Rick, R., & Eisenberg, D. S. (2021). The expanding amyloid family: Structure, stability, function, and pathogenesis. *Cell*, 184(19), 4857-4873.
48. Tsuchida, K., Shioi, J., Yamada, S., Boghosian, G., Wu, A., Cai, H., ... & Robakis, N. K. (2001). Appican, the proteoglycan form of the amyloid precursor protein, contains chondroitin sulfate E in the repeating disaccharide region and 4-O-sulfated galactose in the linkage region. *Journal of Biological Chemistry*, 276(40), 37155-37160.
49. Pangalos, M. N., Shioi, J., Efthimiopoulos, S., Wu, A., & Robakis, N. K. (1996). Characterization of appican, the chondroitin sulfate proteoglycan form of the Alzheimer amyloid precursor protein. *Neurodegeneration*, 5(4), 445-451.
50. Müller, U. C., Deller, T., & Korte, M. (2017). Not just amyloid: physiological functions of the amyloid precursor protein family. *Nature Reviews Neuroscience*, 18(5), 281-298.
51. Heftner, D., Ludewig, S., Draguhn, A., & Korte, M. (2020). Amyloid, APP, and electrical activity of the brain. *The Neuroscientist*, 26(3), 231-251.
52. Turner, P. R., O'Connor, K., Tate, W. P., & Abraham, W. C. (2003). Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. *Progress in neurobiology*, 70(1), 1-32.
53. Korte, M., Herrmann, U., Zhang, X., & Draguhn, A. (2012). The role of APP and APLP for synaptic transmission, plasticity, and network function: lessons from genetic mouse models. *Experimental brain research*, 217, 435-440.
54. Russell, C. L., Semerdjieva, S., Empson, R. M., Austen, B. M., Beesley, P. W., & Alifragis, P. (2012). Amyloid- β acts as a regulator of neurotransmitter release disrupting the interaction between synaptophysin and VAMP2. *PLoS ONE* 7(8): e43201.
55. Al-Kuraishi, H. M., Jabir, M. S., Al-Gareeb, A. I., Albuhadily, A. K., Albukhaty, S., Sulaiman, G. M., & Batiha, G. E. S. (2023). Evaluation and targeting of amyloid precursor protein (APP)/amyloid beta (A β) axis in amyloidogenic and non-amyloidogenic pathways: A time outside the tunnel. *Ageing Research Reviews*, 92, 102119.
56. Orobets, K., Karamyshev, AL. (2023) Amyloid Precursor Protein and Alzheimer's Disease. . *Int J Mol Sci* 24, 14794.
57. Kuo, C. C., Chiang, A. W., Baghdassarian, H. M., & Lewis, N. E. (2021). Dysregulation of the secretory pathway connects Alzheimer's disease genetics to aggregate formation. *Cell systems*, 12(9), 873-884.
58. Sun, J., & Roy, S. (2018). The physical approximation of APP and BACE-1: A key event in alzheimer's disease pathogenesis. *Developmental neurobiology*, 78(3), 340-347.
59. Tackenberg, C., & Nitsch, R. M. (2019). The secreted APP ectodomain sAPP α , but not sAPP β , protects neurons against A β oligomer-induced dendritic spine loss and increased tau phosphorylation. *Molecular brain*, 12, 1-4.
60. Baratchi, S., Evans, J., Tate, W. P., Abraham, W. C., & Connor, B. (2012). Secreted amyloid precursor proteins promote proliferation and glial differentiation of adult hippocampal neural progenitor cells. *Hippocampus*, 22(7), 1517-1527.
61. Dar, N. J., & Glazner, G. W. (2020). Deciphering the neuroprotective and neurogenic potential of soluble amyloid precursor protein alpha (sAPP α). *Cellular and Molecular Life Sciences*, 77, 2315-2330.
62. Habib, A., Sawmiller, D., & Tan, J. (2017). Restoring Soluble Amyloid Precursor Protein α Functions as a Potential Treatment for Alzheimer's Disease. *Journal of neuroscience research*, 95(4), 973-991.
63. Castro, M. A., Hadziselimovic, A., & Sanders, C. R. (2019). The vexing complexity of the amyloidogenic pathway. *Protein Science*, 28(7), 1177-1193.
64. Nalivaeva, N. N., & Turner, A. J. (2013). The amyloid precursor protein: a biochemical enigma in brain development, function and disease. *FEBS letters*, 587(13), 2046-2054.
65. Sosa, L. J., Cáceres, A., Dupraz, S., Oksdath, M., Quiroga, S., & Lorenzo, A. (2017). The physiological role of the amyloid precursor protein as an adhesion molecule in the developing nervous system. *Journal of neurochemistry*, 143(1), 11-29.
66. Chasseigneaux, S., & Allinquant, B. (2012). Functions of A β , sAPP α and sAPP β : similarities and differences. *Journal of neurochemistry*, 120, 99-108.
67. López de la Paz, M., & Serrano, L. (2004). Sequence determinants of amyloid fibril formation. *Proceedings of the National Academy of Sciences*, 101(1), 87-92.
68. DuBay, K. F., Pawar, A. P., Chiti, F., Zurdo, J., Dobson, C. M., & Vendruscolo, M. (2004). Prediction of the absolute aggregation rates of amyloidogenic polypeptide chains. *Journal of molecular biology*, 341(5), 1317-1326.
69. Pawar, A. P., Dubay, K. F., Zurdo, J., Chiti, F., Vendruscolo, M., & Dobson, C. M. (2005). Prediction of "aggregation-prone" and "aggregation-susceptible" regions in proteins associated with neurodegenerative diseases. *Journal of molecular biology*, 350(2), 379-392.
70. Zibaee, S., Makin, O. S., Goedert, M., & Serpell, L. C. (2007). A simple algorithm locates β -strands in the amyloid fibril core of α -synuclein, A β , and tau using the amino acid sequence alone. *Protein Science*, 16(5), 906-918.
71. Fernandez-Escamilla, A. M., Rousseau, F., Schymkowitz, J., & Serrano, L. (2004). Prediction of sequence-dependent and mutational effects on the aggregation of peptides and proteins. *Nature biotechnology*, 22(10), 1302-1306.
72. Palato, L. M., Pilcher, S., Oakes, A., Lamba, A., Torres,

- J., Ledesma Monjaraz, L. I., ... & Moffet, D. A. (2019). Amyloidogenicity of naturally occurring full-length animal IAPP variants. *Journal of Peptide Science*, 25(8), e3199.
73. Conchillo-Solé, O., de Groot, N. S., Avilés, F. X., Vendrell, J., Daura, X., & Ventura, S. (2007). AGGRESCAN: a server for the prediction and evaluation of "hot spots" of aggregation in polypeptides. *BMC bioinformatics*, 8, 1-17.
74. Rambaran, R. N., & Serpell, L. C. (2008). Amyloid fibrils: abnormal protein assembly. *Prion*, 2(3), 112-117.
75. Castillo, V., Graña-Montes, R., Sabate, R., & Ventura, S. (2011). Prediction of the aggregation propensity of proteins from the primary sequence: aggregation properties of proteomes. *Biotechnology journal*, 6(6), 674-685.
76. De Groot, N. S., Castillo, V., Graña-Montes, R., & Ventura, S. (2012). AGGRESCAN: method, application, and perspectives for drug design. In: Baron, R. (eds) Computational Drug Discovery and Design. Methods in Molecular Biology, vol 819, pp 199-220. Springer, New York, NY.
77. Tsolis, A. C., Papandreou, N. C., Iconomidou, V. A., & Hamodrakas, S. J. (2013). A consensus method for the prediction of 'aggregation-prone' peptides in globular proteins. *PloS one*, 8(1), e54175.
78. Cherny, I., & Gazit, E. (2008). Amyloids: not only pathological agents but also ordered nanomaterials. *Angewandte Chemie International Edition*, 47(22), 4062-4069.
79. Zhang, Z., Chen, H., & Lai, L. (2007). Identification of amyloid fibril-forming segments based on structure and residue-based statistical potential. *Bioinformatics*, 23(17), 2218-2225.
80. Bondarev, S. A., Uspenskaya, M. V., Leclercq, J., Falgarone, T., Zhuravleva, G. A., & Kajava, A. V. (2024). AmyloComp: a bioinformatic tool for prediction of amyloid co-aggregation. *Journal of Molecular Biology*, 168437.
81. Wojciechowski, J. W., Szczurek, W., Szulc, N., Szefczyk, M., & Kotulska, M. (2023). PACT-Prediction of amyloid cross-interaction by threading. *Scientific Reports*, 13(1), 22268.
82. Emily, M., Talvas, A., & Delamarche, C. (2013). MetAmyl: a METa-predictor for AMYloid proteins. *PloS one*, 8(11), e79722.
83. Charoenkwan, P., Kanthawong, S., Nantasesamat, C., Hasan, M. M., & Shoombuatong, W. (2021). iAMY-SCM: Improved prediction and analysis of amyloid proteins using a scoring card method with propensity scores of dipeptides. *Genomics*, 113(1), 689-698.
84. Niu, M., Li, Y., Wang, C., & Han, K. (2018). RFamyloid: a web server for predicting amyloid proteins. *International journal of molecular sciences*, 19(7), 2071.
85. Conchillo-Solé, O., de Groot, N. S., Avilés, F. X., Vendrell, J., Daura, X., & Ventura, S. (2007). AGGRESCAN: a server for the prediction and evaluation of "hot spots" of aggregation in polypeptides. *BMC bioinformatics*, 8, 1-17.
86. Hamodrakas, S. J. (2011). Protein aggregation and amyloid fibril formation prediction software from primary sequence: towards controlling the formation of bacterial inclusion bodies. *The FEBS journal*, 278(14), 2428-2435.
87. Charoenkwan, P., Ahmed, S., Nantasesamat, C., Quinn, J. M., Moni, M. A., Lio', P., & Shoombuatong, W. (2022). AMYPred-FRL is a novel approach for accurate prediction of amyloid proteins by using feature representation learning. *Scientific reports*, 12(1), 7697.
88. Nair, S. S. K., Reddy, N. S., & Hareesa, K. S. (2012). AmylPepPred: amyloidogenic peptide prediction tool. *Bioinformation*, 8(20), 994.
89. Tian, J., Wu, N., Guo, J., & Fan, Y. (2009). Prediction of amyloid fibril-forming segments based on a support vector machine. *BMC bioinformatics*, 10, 1-8.
90. Xiao, X., Robang, A. S., Sarma, S., Le, J. V., Helmicki, M. E., Lambert, M. J., ... & Hall, C. K. (2022). Sequence patterns and signatures: Computational and experimental discovery of amyloid-forming peptides. *PNAS nexus*, 1(5), pgac263.
91. Morris, K. L., Rodger, A., Hicks, M. R., Debulpaep, M., Schymkowitz, J., Rousseau, F., & Serpell, L. C. (2013). Exploring the sequence-structure relationship for amyloid peptides. *Biochemical Journal*, 450(2), 275-283.
92. López de la Paz, M., & Serrano, L. (2004). Sequence determinants of amyloid fibril formation. *Proceedings of the National Academy of Sciences*, 101(1), 87-92.
93. Al-Garawi, Z. S., Morris, K. L., Marshall, K. E., Eichler, J., & Serpell, L. C. (2017). The diversity and utility of amyloid fibrils formed by short amyloidogenic peptides. *Interface Focus*, 7(6), 20170027.
94. Thompson, M. J., Sievers, S. A., Karanicolas, J., Ivanova, M. I., Baker, D., & Eisenberg, D. (2006). The 3D profile method for identifying fibril-forming segments of proteins. *Proceedings of the National Academy of Sciences*, 103(11), 4074-4078.
95. Galzitskaya, O. V., Garbuzynskiy, S. O., & Lobanov, M. Y. (2006). Prediction of amyloidogenic and disordered regions in protein chains. *PLoS computational biology*, 2(12), e177.
96. Família, C., Dennison, S. R., Quintas, A., & Phoenix, D. A. (2015). Prediction of peptide and protein propensity for amyloid formation. *PloS one*, 10(8), e0134679.
97. Belli, M., Ramazzotti, M., & Chiti, F. (2011). Prediction of amyloid aggregation in vivo. *EMBO reports*, 12(7), 657-663.
98. Mostaert, A. S., Crockett, R., Kearn, G., Cherny, I., Gazit, E., Serpell, L. C., & Jarvis, S. P. (2009). Mechanically functional amyloid fibrils in the adhesive of a marine invertebrate as revealed by Raman spectroscopy and atomic force microscopy. *Archives of histology and cytology*, 72(4+5), 199-207.
99. Siddiqi, M. K., Majid, N., Malik, S., Alam, P., & Khan, R. H. (2019). Amyloid oligomers, protofibrils and fibrils. *Macromolecular Protein Complexes II: Structure and Function*, 471-503.
100. Dovidchenko, N. V., Leonova, E. I., & Galzitskaya, O. V. (2014). Mechanisms of amyloid fibril formation. *Biochemistry (Moscow)*, 79, 1515-1527.
101. Guyonnet, B., Egge, N., & Cornwall, G. A. (2014). Functional amyloids in the mouse sperm acrosome. *Molecular and cellular biology*, 34(14), 2624-2634.
102. Whelly, S., Johnson, S., Powell, J., Borchardt, C., Hastert, M. C., & Cornwall, G. A. (2012). Nonpathological extracellular

- amyloid is present during normal epididymal sperm maturation. *PloS one*, 7(5), e36394.
103. Whelly, S., Muthusubramanian, A., Powell, J., Johnson, S., Hastert, M. C., & Cornwall, G. A. (2016). Cystatin-related epididymal spermatogenic subgroup members are part of an amyloid matrix and associated with extracellular vesicles in the mouse epididymal lumen. *MHR: Basic science of reproductive medicine*, 22(11), 729-744.
104. Melrose, J. (2022). High performance marine and terrestrial bioadhesives and the biomedical applications they have inspired. *Molecules*, 27(24), 8982.
105. Buchanan, J. A., Varghese, N. R., Johnston, C. L., & Sunde, M. (2023). Functional amyloids: Where supramolecular amyloid assembly controls biological activity or generates new functionality. *Journal of Molecular Biology*, 435(11), 167919.
106. Reches, M., & Gazit, E. (2006). Molecular self-assembly of peptide nanostructures: mechanism of association and potential uses. *Current Nanoscience*, 2(2), 105-111.
107. Adler-Abramovich, L., Aronov, D., Beker, P., Yevnin, M., Stempler, S., Buzhansky, L., ... & Gazit, E. (2009). Self-assembled arrays of peptide nanotubes by vapour deposition. *Nature nanotechnology*, 4(12), 849-854.
108. Carny, O., Shalev, D. E., & Gazit, E. (2006). Fabrication of coaxial metal nanocables using a self-assembled peptide nanotube scaffold. *Nano Letters*, 6(8), 1594-1597.
109. Reches, M., & Gazit, E. (2003). Casting metal nanowires within discrete self-assembled peptide nanotubes. *Science*, 300(5619), 625-627.
110. Reches, M., & Gazit, E. (2006). Controlled patterning of aligned self-assembled peptide nanotubes. *Nature nanotechnology*, 1(3), 195-200.
111. Tiwari, O. S., & Gazit, E. (2024). Characterization of amyloid-like metal-amino acid assemblies with remarkable catalytic activity. *Peptide Catalysts, Including Catalytic Amyloids*, 697, 181.
112. Gazit, E. (2007). Self-assembled peptide nanostructures: the design of molecular building blocks and their technological utilization. *Chemical Society Reviews*, 36(8), 1263-1269.
113. Hauser, C. A., Maurer-Stroh, S., & Martins, I. C. (2014). Amyloid-based nanosensors and nanodevices. *Chemical Society Reviews*, 43(15), 5326-5345.
114. Taheri, R. A., Akhtari, Y., Tohidi Moghadam, T., & Ranjbar, B. (2018). Assembly of gold nanorods on HSA amyloid fibrils to develop a conductive nanoscaffold for potential biomedical and biosensing applications. *Scientific reports*, 8(1), 9333.
115. Miranda, E., & Suñé, J. (2020). Memristors for neuromorphic circuits and artificial intelligence applications. *Materials*, 13(4), 938.
116. Fu, J., Wang, J., He, X., Ming, J., Wang, L., Wang, Y., ... & Ling, H. (2023). Pseudo-transistors for emerging neuromorphic electronics. *Science and Technology of Advanced Materials*, 24(1), 2180286.
117. Barraj, I., Mestiri, H., & Masmoudi, M. (2024). Overview of Memristor-Based Design for Analog Applications. *Micromachines*, 15(4), 505.
118. Jariwala, D., Sangwan, V. K., Lauhon, L. J., Marks, T. J., & Hersam, M. C. (2013). Carbon nanomaterials for electronics, optoelectronics, photovoltaics, and sensing. *Chemical Society Reviews*, 42(7), 2824-2860.
119. Li, C., & Mezzenga, R. (2013). The interplay between carbon nanomaterials and amyloid fibrils in bio-nanotechnology. *Nanoscale*, 5(14), 6207-6218.
120. Terrones, H., Terrones, M., López-Urías, F., Rodríguez-Manzo, J. A., & Mackay, A. L. (2004). Shape and complexity at the atomic scale: the case of layered nanomaterials. *Philosophical Transactions of the Royal Society of London. Series A: Mathematical, Physical and Engineering Sciences*, 362(1823), 2039-2063.
121. Mankar, S., Anoop, A., Sen, S., & Maji, S. K. (2011). Nanomaterials: amyloids reflect their brighter side. *Nano reviews*, 2(1), 6032.
122. Ramanishankar, A., Begum, R. F., Jayasankar, N., Nayeem, A., Prajapati, B. G., & Nirenjen, S. (2024). Unleashing light's healing power: an overview of photobiomodulation for Alzheimer's treatment. *Future Science OA*, 10(1), FSO922.
123. Huang, Y., Chang, Y., Liu, L., & Wang, J. (2021). Nanomaterials for modulating the aggregation of β -amyloid peptides. *Molecules*, 26(14), 4301.
124. Ender, A. M., Kaygisisiz, K., Räder, H. J., Mayer, F. J., Synatschke, C. V., & Weil, T. (2021). Cell-instructive surface gradients of photoresponsive amyloid-like fibrils. *ACS Biomaterials Science & Engineering*, 7(10), 4798-4808.
125. Nilsson, M. R., & Dobson, C. M. (2003). Chemical modification of insulin in amyloid fibrils. *Protein science*, 12(11), 2637-2641.
126. Morris, R., MacPhee, C. (2013). Amyloid Protein Biomaterials. In: Roberts, G.C.K. (eds) Encyclopedia of Biophysics. Springer, Berlin, Heidelberg.
127. Pansieri, J., Josserand, V., Lee, S. J., Rongier, A., Imbert, D., Sallanon, M. M., ... & Forge, V. (2019). Ultraviolet-visible-near-infrared optical properties of amyloid fibrils shed light on amyloidogenesis. *Nature photonics*, 13(7), 473-479.
128. Kim, S., Kim, J. H., Lee, J. S., & Park, C. B. (2015). Beta-sheet-forming, self-assembled peptide nanomaterials towards optical, energy, and healthcare applications. *small*, 11(30), 3623-3640.
129. Camerin, L., Maleeva, G., Gomila, A. M., Suárez-Pereira, I., Matera, C., Prischich, D., ... & Gorostiza, P. (2024). Photoswitchable Carbamazepine Analogs for Non-Invasive Neuroinhibition In Vivo. *Angewandte Chemie*, e202403636.
130. Arrue, L., & Ratjen, L. (2017). Internal targeting and external control: phototriggered targeting in nanomedicine. *ChemMedChem*, 12(23), 1908-1916.
131. Marcus, D. J., & Bruchas, M. R. (2023). Optical Approaches for Investigating Neuromodulation and G Protein-Coupled Receptor Signaling. *Pharmacological Reviews*, 75(6), 1119-1139.
132. Shamsipur, M., Ghavidast, A., & Pashabadi, A. (2023). Phototriggered structures: Latest advances in biomedical

- applications. *Acta Pharmaceutica Sinica B*, 13(7), 2844-2876.
133. Son, G., Lee, S. H., Wang, D., & Park, C. B. (2018). Thioflavin T-Amyloid Hybrid Nanostructure for Biocatalytic Photosynthesis. *Small*, 14(40), 1801396.
134. Balasco, N., Diaferia, C., Rosa, E., Monti, A., Ruvo, M., Doti, N., & Vitagliano, L. (2023). A comprehensive analysis of the intrinsic visible fluorescence emitted by peptide/protein amyloid-like assemblies. *International journal of molecular sciences*, 24(9), 8372.
135. Aziz, A. A., Siddiqui, R. A., & Amtul, Z. (2020). Engineering of fluorescent or photoactive Trojan probes for detection and eradication of β -Amyloids. *Drug Delivery*, 27(1), 917-926.
136. Fowler, D. M., Koulov, A. V., Balch, W. E., & Kelly, J. W. (2007). Functional amyloid—from bacteria to humans. *Trends in biochemical sciences*, 32(5), 217-224.
137. Gras, S. L., Tickler, A. K., Squires, A. M., Devlin, G. L., Horton, M. A., Dobson, C. M., & MacPhee, C. E. (2008). Functionalised amyloid fibrils for roles in cell adhesion. *Biomaterials*, 29(11), 1553-1562.
138. Maji, S. K., Schubert, D., Rivier, C., Lee, S., Rivier, J. E., & Riek, R. (2008). Amyloid as a depot for the formulation of long-acting drugs. *PLoS biology*, 6(2), e17.
139. Pertinhez, T. A., Conti, S., Ferrari, E., Magliani, W., Spisni, A., & Polonelli, L. (2009). Reversible self-assembly: a key feature for a new class of autodelivering therapeutic peptides. *Molecular pharmaceutics*, 6(3), 1036-1039.
140. Scheibel, T., Parthasarathy, R., Sawicki, G., Lin, X. M., Jaeger, H., & Lindquist, S. L. (2003). Conducting nanowires built by controlled self-assembly of amyloid fibers and selective metal deposition. *Proceedings of the National Academy of Sciences*, 100(8), 4527-4532.
141. Bhak, G., Lee, S., Park, J. W., Cho, S., & Paik, S. R. (2010). Amyloid hydrogel derived from curly protein fibrils of α -synuclein. *Biomaterials*, 31(23), 5986-5995.
142. Herland, A., Björk, P., Nilsson, K. P. R., Hammarström, P., Inganäs, O., & Konradsson, P. (2005). Electroactive luminescent self-assembled bio-organic nanowires: integration of semiconducting oligoelectrolytes within amyloidogenic proteins. *Advanced Materials*, 17(12), 1466-1471.
143. Tanaka, H., Herland, A., Lindgren, L. J., Tsutsui, T., & Andersson, M. R. (2008). Enhanced current efficiency from bio-organic light-emitting diodes using decorated amyloid fibrils with conjugated polymer. *Nano letters*, 8(9), 2858-2861.
144. Herland, A., Thomsson, D., Mirzov, O., Scheblykin, I. G., & Inganäs, O. (2008). Decoration of amyloid fibrils with luminescent conjugated polymers. *Journal of Materials Chemistry*, 18(1), 126-132.
145. Hamed, M., Herland, A., Karlsson, R. H., & Inganäs, O. (2008). Electrochemical devices made from conducting nanowire networks self-assembled from amyloid fibrils and alkoxy sulfonate PEDOT. *Nano letters*, 8(6), 1736-1740.
146. Ahn, M., Kang, S., Koo, H. J., Lee, J. H., Lee, Y. S., & Paik, S. R. (2010). Nanoporous protein matrix made of amyloid fibrils of β 2-microglobulin. *Biotechnology progress*, 26(6), 1759-1764.
147. Reynolds, N. P. (2019). Amyloid-like peptide nanofibrils as scaffolds for tissue engineering: Progress and challenges. *Biointerphases*, 14(4).
148. Peña-Díaz, S., Olsen, W. P., Wang, H., & Otzen, D. E. (2024). Functional amyloids: The biomaterials of tomorrow?. *Advanced Materials*, 36(18), 2312823.
149. Aggeli, A., Boden, N., & Zhang, S. (Eds.). (2001). *Self-assembling peptide systems in biology, medicine, and engineering*. Dordrecht: Kluwer Academic Publishers.
150. Zhang, S. (2003). More than just bare scaffolds: towards multi-component and decorated fibrous biomaterials. *Nat Biotechnol* 21, 1171-1178.
151. Eisenberg, D., & Jucker, M. (2012). The amyloid state of proteins in human diseases. *Cell*, 148(6), 1188-1203.
152. Greenwald, J., & Rick, R. (2010). Biology of amyloid: structure, function, and regulation. *Structure*, 18(10), 1244-1260.
153. Pham, C. L., Kwan, A. H., & Sunde, M. (2014). Functional amyloid: widespread in Nature, diverse in purpose. *Essays in biochemistry*, 56, 207-219.
154. Syed, A. K., & Boles, B. R. (2014). Fold modulating function: bacterial toxins to functional amyloids. *Frontiers in microbiology*, 5, 401.
155. Morris, R. J., & MacPhee, C. E. (2013). Amyloid protein biomaterials. *Encyclopedia of Biophysics*, 76-81.
156. Collier, J. H., & Messersmith, P. B. (2004). Self-Assembling Polymer-Peptide Conjugates: Nanostructural Tailoring. *Advanced Materials*, 16(11), 907-910.
157. Knowles, T. P., Fitzpatrick, A. W., Meehan, S., Mott, H. R., Vendruscolo, M., Dobson, C. M., & Welland, M. E. (2007). Role of intermolecular forces in defining material properties of protein nanofibrils. *science*, 318(5858), 1900-1903.
158. Scheibel, T., Parthasarathy, R., Sawicki, G., Lin, X. M., Jaeger, H., & Lindquist, S. L. (2003). Conducting nanowires built by controlled self-assembly of amyloid fibers and selective metal deposition. *Proceedings of the National Academy of Sciences*, 100(8), 4527-4532.
159. Willner, I., & Katz, E. (Eds.). (2006). *Bioelectronics: from theory to applications*. John Wiley & Sons.
160. Ligorio, C., & Mata, A. (2023). Synthetic extracellular matrices with function-encoding peptides. *Nature reviews bioengineering*, 1(7), 518-536.
161. Deidda, G., Jonnalagadda, S. V. R., Spies, J. W., Ranella, A., Mossou, E., Forsyth, V. T., ... & Mitraki, A. (2017). Self-assembled amyloid peptides with Arg-Gly-Asp (RGD) motifs as scaffolds for tissue engineering. *ACS Biomaterials Science & Engineering*, 3(7), 1404-1416.
162. van Dalen, M. C., Karperien, M., Claessens, M. M., & Post, J. N. (2023). Choice of Protein, Not Its Amyloid-Fold, Determines the Success of Amyloid-Based Scaffolds for Cartilage Tissue Regeneration. *ACS omega*, 8(27), 24198-24209.
163. Koo, E. H., Park, L., & Selkoe, D. J. (1993). Amyloid beta-protein as a substrate interacts with extracellular matrix to

- promote neurite outgrowth. *Proceedings of the National Academy of Sciences*, 90(10), 4748-4752.
164. Ranjan, V. D., Qiu, L., Lee, J. W. L., Chen, X., Jang, S. E., Chai, C., ... & Zeng, L. (2020). A microfiber scaffold-based 3D in vitro human neuronal culture model of Alzheimer's disease. *Biomaterials Science*, 8(17), 4861-4874.
165. Mathes, T. G., Monirizad, M., Ermis, M., de Barros, N. R., Rodriguez, M., Kraatz, H. B., ... & Falcone, N. (2024). Effects of amyloid- β -mimicking peptide hydrogel matrix on neuronal progenitor cell phenotype. *Acta Biomaterialia*, 183, 89-100.
166. Jacob, R. S., Ghosh, D., Singh, P. K., Basu, S. K., Jha, N. N., Das, S., ... & Maji, S. K. (2015). Self healing hydrogels composed of amyloid nano fibrils for cell culture and stem cell differentiation. *Biomaterials*, 54, 97-105.
167. Xuan, Q., Wang, Y., Chen, C., & Wang, P. (2021). Rational biological interface engineering: Amyloidal supramolecular microstructure-inspired hydrogel. *Frontiers in Bioengineering and Biotechnology*, 9, 718883.
168. Yanlian, Y., Ulung, K., Xiumei, W., Horii, A., Yokoi, H., & Shuguang, Z. (2009). Designer self-assembling peptide nanomaterials. *Nano Today*, 4(2), 193-210.
169. Liang, Y., Guo, P., Pingali, S. V., Pabit, S., Thiagarajan, P., Berland, K. M., & Lynn, D. G. (2008). Light harvesting antenna on an amyloid scaffold. *Chemical communications*, (48), 6522-6524.
170. Channon, K. J., Devlin, G. L., & MacPhee, C. E. (2009). Efficient energy transfer within self-assembling peptide fibers: a route to light-harvesting nanomaterials. *Journal of the American Chemical Society*, 131(35), 12520-12521.
171. Choi, Y. S., Kim, J., Bhak, G., Lee, D., & Paik, S. R. (2011). Photoelectric Protein Nanofibrils of α -Synuclein with Embedded Iron and Phthalocyanine Tetrasulfonate. *Angewandte Chemie*, 123(27), 6194-6198.
172. Ridgley, D. M., Ebanks, K. C., & Barone, J. R. (2011). Peptide mixtures can self-assemble into large amyloid fibers of varying size and morphology. *Biomacromolecules*, 12(10), 3770-3779.
173. Ridgley, D. M., & Barone, J. R. (2013). Evolution of the amyloid fiber over multiple length scales. *ACS nano*, 7(2), 1006-1015.
174. Ridgley, D. M., Claunch, E. C., & Barone, J. R. (2012). The effect of processing on large, self-assembled amyloid fibers. *Soft Matter*, 8(40), 10298-10306.
175. Diaz-Pier, S., & Carloni, P. (2024). Impact of quantum and neuromorphic computing on biomolecular simulations: Current status and perspectives. *Current Opinion in Structural Biology*, 87, 102817.
176. Liu, R., Liu, T., Liu, W., Luo, B., Li, Y., Fan, X., ... & Teng, Y. (2024). SemiSynBio: A new era for neuromorphic computing. *Synthetic and Systems Biotechnology*, 9(3), 594-599.
177. Sun, Y., Wang, H., & Xie, D. (2024). Recent Advance in Synaptic Plasticity Modulation Techniques for Neuromorphic Applications. *Nano-Micro Letters*, 16(1), 1-32.
178. Taylor, A. I., & Staniforth, R. A. (2022). General principles underpinning amyloid structure. *Frontiers in Neuroscience*, 16, 878869.
179. Snow, A. D., Cummings, J. A., & Lake, T. (2021). The unifying hypothesis of Alzheimer's disease: Heparan sulfate proteoglycans/glycosaminoglycans are key as first hypothesized over 30 years ago. *Frontiers in Aging Neuroscience*, 13, 710683.
180. Melrose, J., & Smith, M. M. (2024). Heparan sulfate proteoglycans in pathological protein aggregation and brain functionality. *Advanced Neurology*, 3(3), 3812.
181. Ozsan McMillan, I., Li, J. P., & Wang, L. (2023). Heparan sulfate proteoglycan in Alzheimer's disease: aberrant expression and functions in molecular pathways related to amyloid- β metabolism. *American Journal of Physiology-Cell Physiology*, 324(4), C893-C909.
182. Nguyen, D. L., Okolicsanyi, R. K., & Haupt, S. L. M. (2024). Heparan sulfate proteoglycans: Mediators of cellular and molecular Alzheimer's disease pathogenic factors via tunnelling nanotubes?. *Molecular and Cellular Neuroscience*, 103936.
183. Schultheis, N., Becker, R., Berhanu, G., Kapral, A., Roseman, M., Shah, S., ... & Selleck, S. (2023). Regulation of autophagy, lipid metabolism, and neurodegenerative pathology by heparan sulfate proteoglycans. *Frontiers in Genetics*, 13, 1012706.
184. Gyimesi, M., Okolicsanyi, R. K., & Haupt, L. M. (2024). Beyond amyloid and tau: rethinking Alzheimer's disease through less explored avenues. *Open Biology*, 14(6), 240035.
185. Wang, P., Zhao, J., Hossaini Nasr, S., Otieno, S. A., Zhang, F., Qiang, W., ... & Huang, X. (2021). Probing Amyloid β Interactions with Synthetic Heparan Sulfate Oligosaccharides. *ACS chemical biology*, 16(10), 1894-1899.
186. Wu, L., Jiang, W., Zhao, N., & Wang, F. (2022). Heparan sulfate from porcine mucosa promotes amyloid-beta clearance in APP/PS1 mice and alleviates Alzheimer's pathology. *Carbohydrate Polymers*, 285, 119205.
187. Lorente-Gea, L., García, B., Martín, C., Ordiales, H., García-Suárez, O., Piña-Batista, K. M., ... & Fernández-Vega, I. (2020). Heparan sulfate proteoglycans undergo differential expression alterations in Alzheimer disease brains. *Journal of Neuropathology & Experimental Neurology*, 79(5), 474-483.
188. Yamada, M., & Hamaguchi, T. (2018). The sulfation code for propagation of neurodegeneration. *Journal of Biological Chemistry*, 293(27), 10841-10842.
189. Melrose, J. (2024). Dystroglycan-HSPG interactions provide synaptic plasticity and specificity. *Glycobiology*, 34(10), cwae051.
190. Roy, A., Chalapathi, A. V., & Balagurunathan, K. (2022). Investigating the Roles of Heparan Sulfate Structures in Alpha-Synuclein Aggregation in Cell Culture Models. *Glycosaminoglycans: Methods and Protocols*, 807-820.
191. Uchimura, K., Nishitsuji, K., Chiu, L. T., Ohgita, T., Saito, H., Allain, F., ... & Hung, S. C. (2022). Design and Synthesis of 6-O-Phosphorylated Heparan Sulfate Oligosaccharides to Inhibit Amyloid β Aggregation. *Chembiochem*, 23(15), e202200191.

192. Melrose, J. (2024). Keratan sulfate, an electrosensory neurosentient bioresponsive cell instructive glycosaminoglycan. *Glycobiology*, 34(3), cwae014.
193. Melrose, J. (2019). Keratan sulfate (KS)-proteoglycans and neuronal regulation in health and disease: the importance of KS-glycodynamics and interactive capability with neuroregulatory ligands. *Journal of Neurochemistry*, 149(2), 170-194.
194. Caterson, B., & Melrose, J. (2018). Keratan sulfate, a complex glycosaminoglycan with unique functional capability. *Glycobiology*, 28(4), 182-206.
195. Santos, F. M., Nunes, S. C., & de Zea Bermudez, V. (2024). Looking beyond biology: glycosaminoglycans as attractive platforms for energy devices and flexible electronics. *Energy Advances*, 3(8), 1766-1843.
196. Melrose, J. (2024). CNS/PNS proteoglycans functionalize neuronal and astrocyte niche microenvironments optimizing cellular activity by preserving membrane polarization dynamics, ionic microenvironments, ion fluxes, neuronal activation, and network neurotransductive capacity. *Journal of Neuroscience Research*, 102(7), e25361.
197. Melrose, J. (2023). Hyaluronan hydrates and compartmentalises the CNS/PNS extracellular matrix and provides niche environments conducive to the optimisation of neuronal activity. *Journal of Neurochemistry*, 166(4), 637-653.
198. Chowdhury, S., & Sarkar, N. (2024). Exploring the potential of amyloids in biomedical applications: A review. *Biotechnology and Bioengineering*, 121(1), 26-38.
199. Yadav, S. S., Padhy, P. K., Singh, A. K., Sharma, S., Fatima, S., Sinha, A., ... & Priya, S. (2024). Advancements in amyloid-based biological materials for healthcare, environmental and sensing applications. *Materials Advances*, 5(10), 4078-4090.
200. Rashid, M. H., Sen, P. (2024). Recent Advancements in Biosensors for the Detection and Characterization of Amyloids: A Review. *Protein J* 43, 656-674.
201. Kaur, K. K., Allahbadia, G., Singh, M. (2023). Biosensors in Tissue Engineering: An Exhaustive Review with Future Therapeutic Potentials -With Further Updates in Electrochemical Sensors in Medical Practice. *Int J Nanotechnol Nanomed*, 8(1), 197-221.
202. Zhang, X., Ma, Z., Zheng, H., Li, T., Chen, K., Wang, X., ... & Lin, H. (2020). The combination of brain-computer interfaces and artificial intelligence: applications and challenges. *Annals of translational medicine*, 8(11), 712.

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